

In children with autoimmune thyroiditis *CTLA4* and *FCRL3* genes – but not *PTPN22* – are overexpressed when compared to adults

Katarzyna WOJCIECHOWSKA-DURCZYNSKA^{1,2}, Kinga KRAWCZYK-RUSIECKA^{1,2},
Arkadiusz ZYGMUNT^{1,2}, Renata STAWERSKA², Andrzej LEWINSKI^{1,2}

¹ Department of Endocrinology and Metabolic Diseases, Medical University of Lodz, Poland

² Polish Mother's Memorial Hospital – Research Institute, Lodz, Poland

Correspondence to: Andrzej Lewiński MD., PhD.
Department of Endocrinology and Metabolic Diseases
Medical University of Lodz
Rzgowska 281/289, 93-338 Lodz, Poland.
TEL: +48 42 271 11 42; FAX: +48 42 271 11 40 ; E-MAIL: alewin@csk.umed.lodz.pl

Submitted: 2015-12-09 Accepted: 2016-01-19 Published online: 2016-02-28

Key words: autoimmune thyroiditis; children; gene expression

Neuroendocrinol Lett 2016; 37(1):65–69 PMID: 26994388 NEL370116A03 © 2016 Neuroendocrinology Letters • www.nel.edu

Abstract

BACKGROUND: Numerous genetic studies revealed several susceptibility genes of autoimmune thyroid diseases (AITD), including *CTLA4*, *PTPN22* and *FCRL3*. These immune-modulating genes are involved in genetic background of AITD among children and adult patients. However, possible age-related differences in overexpression of these genes remain unclear.

PURPOSE: The goal of this single centre cohort study was evaluation of expression levels of three (3) genes *CTLA4*, *PTPN22* and *FCRL3* in adult patients and children with autoimmune thyroiditis.

METHODS: A total of 47 patients – 24 adults (mean age – 47.7 years) and 23 children (mean age – 12.4 years) with autoimmune thyroiditis were assessed for the level of expression of *CTLA4*, *PTPN22* and *FCRL3* genes, utilizing ABI PRISM® 7500 Sequence Detection System (Applied Biosystem, Foster City, CA, USA).

RESULTS: The overexpression of *PTPN22* (mean RQ=2.988) and *FCRL3* (mean RQ=2.544) genes were confirmed in adult patients with autoimmune thyroiditis, at the same time the expression level of *CTLA4* gene was significantly decreased (mean RQ=0.899) ($p<0.05$). Similar discrepancies were not observed in children with autoimmune thyroiditis in whom overexpression of all three genes – *CTLA4*, *PTPN22* and *FCRL3* – was observed. Differences in *CTLA4* and *FCRL3* genes expression levels in patients with autoimmune thyroiditis were found depending on the age, with increased expression levels of *CTLA4* (mean RQ=3.451) and *FCRL3* (mean RQ=7.410) in children when compared to adults ($p<0.05$) (Mann-Whitney's U-test). There were moderate negative linear correlations between two genes in question (*CTLA4* and *FCRL3*) expression level and patients' age [correlation coefficient (r)=-0.529 ($p<0.0002$) and -0.423 ($p<0.0032$), respectively; Spearman's rank correlation test].

CONCLUSION: Our results are consistent with the hypothesis that there are few age-dependent genetic differences as regards autoimmune thyroiditis in adults and children. Accordingly, *CTLA4* and *FCRL3* genes overexpression may play an important role in children suffering from autoimmune thyroiditis.

INTRODUCTION

Autoimmune thyroid diseases (AITD) are multifactorial disorders in which autoimmunity toward thyroid autoantigens has a specific genetic background and is facilitated by exposure to environmental factors. Among many immune-related genes, which impair the self-tolerance to thyroid autoantibodies and determine the risk of AITD development, the role of *CTLA4*, *PTPN22* and *FCRL3* genes is especially highlighted.

Cytotoxic T lymphocyte-associated factor 4 (*CTLA4*, also known as CD152) has been shown to confer susceptibility to autoimmune diseases (Gough *et al.* 2005), including AITD (Hou *et al.* 2015). This protein is a negative regulator of T cell response and acts by delivering an inhibitory signal which can reverse the T cell receptor (TCR)-induced stop signal needed for interaction between T cell and antigen presenting cell (APC), thus reducing adhesion periods between these cells, which in – turn – decreases cytokine production and proliferation (Schneider *et al.* 2006; Downey *et al.* 2007, Schneider *et al.* 2008).

PTPN22 has already been recognized as a risk factor of autoimmunity and it is associated with Graves' disease (GD). It is a non-receptor type protein tyrosine phosphatase, expressed mainly in hematopoietic cells. *PTPN22* has been shown to attenuate the strength of TCR signals (Wu *et al.* 2006). There are reports indicating that human *PTPN22* also inhibits the activity of B cell antigen receptor (Rieck *et al.* 2007; Arechiga *et al.* 2009). Polymorphism of *PTPN22* resulted both in the gain and loss of *PTPN22* protein function in T cells in some reports (Rieck *et al.* 2007; Zhang *et al.* 2011).

Fc receptor-like 3 (FCRL3) gene is also involved in the pathogenesis of AITD, particularly in GD. It is an orphan cell surface receptor of unknown function with structural homology to classical receptors for immunoglobulin constant chains (Fc receptors). *FCRL3* plays a key role in the development, maturation and function of B-lymphocytes (Matesanz-Isabel *et al.* 2011). The pathogenic activation of *FCRL3* expression leads to down-regulation of B-cell receptor-mediated signalling,

incomplete anergy and deletion in autoreactive B-cells, and finally to breakdown of B-cell tolerance (Kochi *et al.* 2009). Presence of *FCRL3* was also demonstrated on the surface of a subset of Treg cells, characterized by lower relative response to antigenic stimulation and reduced suppressor activity (Swainson *et al.* 2010).

Emerging evidence suggests that these immunomodulating susceptibility genes are involved in pathogenesis of AITD in children (Pastuszek-Lewandoska *et al.* 2013), as well as in adult patients (Lee *et al.* 2015). However, any possible association between *CTLA4*, *PTPN22* and *FCRL3* genes overexpression and patients' age remains unknown. In contrast to other genetic studies evaluating children and adults patients with AITD separately, in this study, for the first time, a representative number of samples from patients of all ages with positive concentrations of antibodies against thyroperoxidase (anti-TPO) have been collected. Data have been analyzed and compared among others as regards the expression of these genes.

METHODS

Peripheral blood samples from patients with autoimmune thyroiditis, hospitalized in Department of Endocrinology and Metabolic Disease Medical University of Lodz, were obtained. Inclusion criteria for autoimmune thyroiditis were the presence of positive values of antibodies against thyroid autoantigens (anti-TPO and antithyroglobulin or anti-TPO alone) and negative value of antibodies against thyrotropin receptor (anti-TSHR). Patients were divided into two groups based on age: 23 children (aged 6–18 years; mean age 12.4) and 24 adults (aged 19–75 years; mean age 47.7). The study procedures were approved by the Local Ethical Committee of Medical University of Lodz and written informed consent was obtained from all participating adults individuals or – in case of children – from parents or legal guardians. Blood samples from the patients without thyroid autoimmunity served as a control for real-time PCR experiment (calibrator). Total RNA from the blood was extracted according to modi-

Tab. 1. Basic measures characterizing gene expressions in selected groups.

		N	Mean	SD	SEM	95% CI		Minimum	Maximum
						Lower CB	Upper CB		
CTLA4	children	23	3.451	4.584	0.955	1.468	5.433	0.263	17.168
	adults	24	0.899	1.364	0.278	0.322	1.475	0.039	6.305
	both	47	2.147	3.558	0.518	1.103	3.192	0.039	17.168
PTPN22	children	23	3.353	4.076	0.850	1.591	5.116	0.304	18.762
	adults	24	2.988	5.332	1.088	0.736	5.239	0.215	25.291
	both	47	3.167	4.711	0.687	1.783	4.550	0.215	25.291
FCRL3	children	23	7.410	9.751	2.033	3.193	11.626	0.949	37.045
	adults	24	2.544	2.979	0.608	1.286	3.803	0.337	14.569
	both	47	4.925	7.480	1.091	2.729	7.122	0.337	37.045

(SD – standard deviation; SEM – standard error of the mean; CI – confidence interval; CB – confidence bound).

fied Chomczynski and Sacchi's method. The purity of total RNA was assessed by NanoDrop® ND-100 spectrophotometer (data not presented). Total RNA was used in the first strand cDNA synthesis with TaqMan® Reverse Transcription Reagents (Applied Biosystem, Branchburg, New Jersey, USA) according to manufacturers' instruction. Real-time PCR was performed on the ABI PRISM® 7500 Sequence Detection System (Applied Biosystem, Foster City, CA, USA) by using TaqMan® Universal PCR Master Mix (Applied Biosystem) and TaqMan® Gene Expression Assays probe and primer mix (Applied Biosystem) according to the manufacturer's specification. The Assays Identification numbers were: CTLA4: Hs03044418_m1; PTPN22: Hs01587518_m1, FCRL3: Hs00364720_m1. Thermal cycler conditions were as follows: hold for 10 min. at 95 °C, followed by two-step PCR for 50 cycles of 95 °C for 15 s followed by 60 °C for 1 min. Amplification reactions, in triplicate for each sample, were performed and the results were normalized to the ACTB gene expression level. An analysis of relative gene expression data was performed, using the $2^{-\Delta\Delta CT}$ method on an ABI PRISM® 7500 Sequence Detection System Software. The calibrator was prepared as a cDNA mix from all cDNA samples (separately, 5 healthy adult controls and 2 healthy children controls). The fold change in studied gene expression, normalised to endogenous control, was calculated using: $RQ=2^{-\Delta\Delta CT}$ (Table 1).

All statistical calculations were performed using IBM SPSS Statistics for Windows, Version 20.0 (IBM Corp. Armonk, NY) with the level of statistical significance $p < 0.05$. Basic measures of location (i.e. mean), measures of dispersion (SD, SEM), and 95% confidence interval for the mean and minimum and maximum values were calculated to provide detailed descriptions of gene expressions in selected groups (Table 1). A *t*-test was used to evaluate CTLA4, PTPN22 and FCRL3 gene expression levels in children and adult groups. Subsequently, the data were statistically analyzed, using non-parametric Mann-Whitney's *U* test in order to compare expression level values (RQ) CTLA4, FCRL3 and PTPN22 between studied independent groups (adults vs. children). Furthermore, Spearman's rank correlation coefficient was used to describe the correlation between CTLA4, FCRL3 and PTPN22 genes expression levels and patient's age.

RESULTS

The overexpression of PTPN22 (mean RQ=2.988) and FCRL3 (mean RQ=2.544) genes were confirmed in adult patients with autoimmune thyroiditis, at the same time the expression level of CTLA4 gene was significantly decreased (mean RQ=0.899) ($p < 0.05$). Similar discrepancies were not observed in children with autoimmune thyroiditis in whom the overexpression of all three genes – CTLA4, PTPN22 and FCRL3 – was observed.

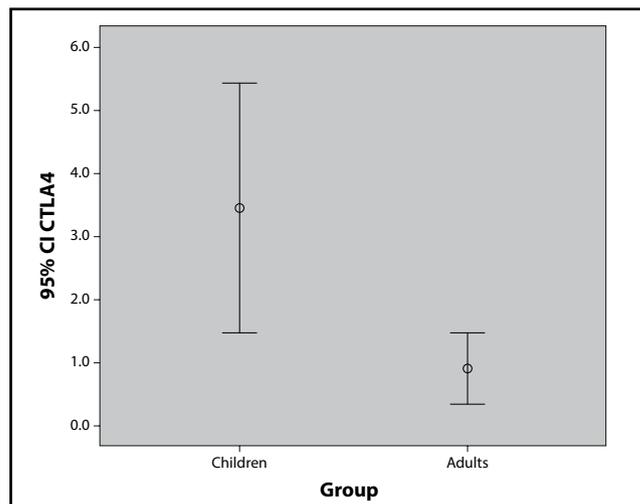


Fig. 1. The box-and-whisker plot diagram representing the different expression levels of CTLA4 in adults and children (data represent means \pm and 95% confidence interval).

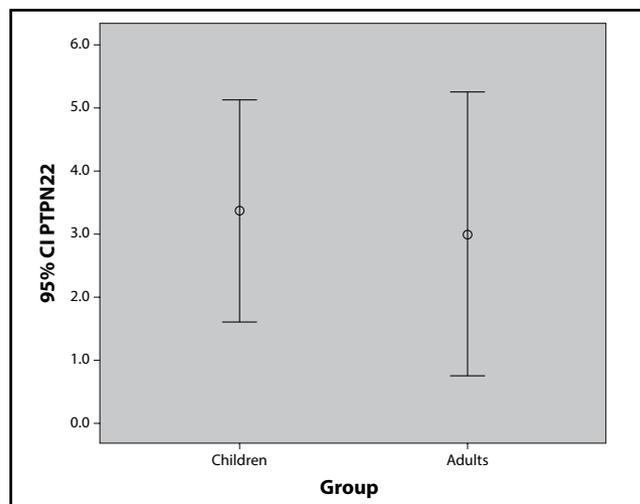


Fig. 2. The box-and-whisker plot diagram representing the different expression levels of PTPN22 in adults and children (data represent means \pm and 95% confidence interval).

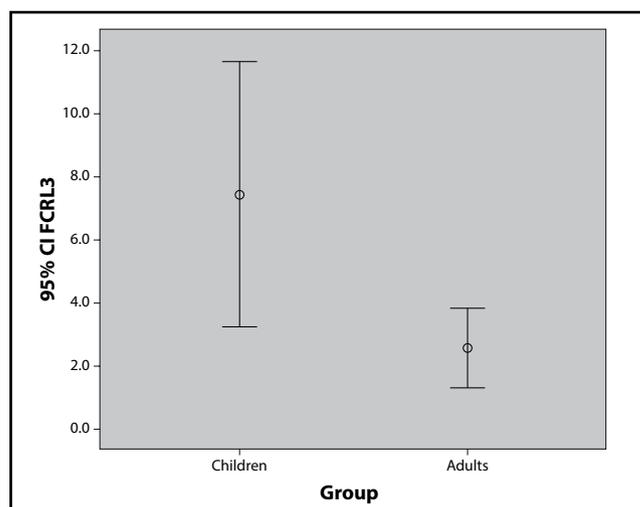


Fig. 3. The box-and-whisker plot diagram representing the different expression levels of FCRL3 in adults and children (data represent means \pm and 95% confidence interval).

Differences in *CTLA4* and *FCRL3* genes expression levels in patients with autoimmune thyroiditis were found depending on the age, with increased expression levels of *CTLA4* (mean RQ=3.451) and *FCRL3* (mean RQ=7.410) in children when compared to adults ($p<0.05$) (Mann-Whitney's U test). The box-and-whisker plot diagrams, representing the different expression levels of *CTLA4*, *PTPN22* and *FCRL3* genes in adults and children, are shown in Figures 1–3, respectively.

Accordingly, there were moderate negative linear correlations between two genes in question (*CTLA4* and *FCRL3*) expression level and patients' age [correlation coefficient (r)= -0.529 ($p<0.0002$) and -0.423 ($p<0.0032$), respectively; Spearman's rank correlation test]. Such relationship was not recorded for *PTPN22* gene ($r=-0.154$, $p>0.3$).

DISCUSSION

Autoimmune thyroid diseases arise due to complex interactions between environmental and genetic factors (Effraïmidis & Wiersinga 2014). Possible involvement of environmental insults, endocrine disruptors and duration of exposure are unquestioned risk factors for the development of diseases. That is confirmed by far less prevalence of AITD in children than in adults (Wiersinga 2014). However, latest studies demonstrate and highlight a strong genetic influence on the development of AITD, especially in children and adolescence (Pastuszak-Lewandoska *et al.* 2013; Wiersinga 2014). Accordingly, female dominance as regards autoimmune diseases – commonly observed in adulthood – is less marked in children and this phenomenon cannot be explained exclusively by shorter exposure to environmental factors. This fact possibly results from specific age-related genetic background in this group of patients (Wiersinga 2014). In accordance, AITD in children is quite frequently related to various genetic syndroms (Glick *et al.* 2013). Identification of AITD in childhood enhances risk of developing other autoimmune disorders with genetic background, including diabetes, celiac disease, etc. (Tolone *et al.* 2009; Glick *et al.* 2013; Riquetto *et al.* 2015). Significant progress has been made in finding the AITD susceptibility genes and understanding the mechanisms by which they confer risk for disease in both adults and children. On the other hand, little is known about particular, risk genetic factors involved in pathogenesis of autoimmune thyroiditis in childhood and adolescence.

In the present study, the increased expression of *CTLA4*, *PTPN22* and *FCRL3* genes in children with autoimmune thyroiditis has been proved. Comparing children and adults with autoimmune thyroiditis, significant differences in expression of *CTLA4* and *FCRL3* have been observed, with higher values in children (Figures 1–3). This relation has not been recorded for *PTPN22* gene. Our further statistical analysis has confirmed existence of moderate negative correlation between

CTLA4 or *FCRL3* gene expression level and patients' age. This observation speaks in favour of the hypothesis on age-specific genetic risk factors acting in childhood and precipitating autoimmune thyroiditis in children.

To our knowledge, there are only scarce reported data on the expression of *CTLA4*, *PTPN22* and *FCRL3* in pathogenesis of autoimmune thyroiditis in children. In contrast, the significant role of above genes in the pathogenesis of not only AITD, but also in several other autoimmune processes in adults has been demonstrated in many reports. According to the previous studies, a polymorphism in *FCRL3* is predisposing to rheumatoid arthritis, systemic lupus erythematosus and biliary cirrhosis (Kochi *et al.* 2005; Effraïmidis & Wiersinga 2014; Zhao *et al.* 2013) and interestingly, the increased expression of *FCRL3* by real-time PCR has also been confirmed in GD (Zhao *et al.* 2013) and endometriosis (Szczepańska *et al.* 2013).

Noteworthy, *PTPN22* gene was originally proved to be associated with type 1 diabetes mellitus (DM1) in children (Liu *et al.* 2015), subsequently it was shown to increase the risk of other autoimmune diseases, including GD, rheumatoid arthritis, juvenile idiopathic arthritis and autoimmune primary adrenal insufficiency (Lee *et al.* 2006), but it reduced the risk of Crohn's disease (Spalinger *et al.* 2013).

CTLA4 has been identified as the most important genetic factor in both GD and Hashimoto thyroiditis (HT) in adults (Hou *et al.* 2015). Results of studies by Kucharska *et al.* (2010, 2013) demonstrated that surface expression of *CTLA4* on T-cells is decreased in children with HT. Our present results are consistent with above cited data in group of adult persons, but not in children in whom expression of *CTLA4* has been increased. Our data speak for an essential role of *CTLA4* gene in development of childhood autoimmune thyroiditis.

Autoimmune thyroid diseases are often associated with DM1. It has been shown that both *PTPN22* and *CTLA4* genes are associated with co-occurrence of HT and DM1 in the same individual (Tomer *et al.* 2015). It is to be emphasized that increased expression of these genes in patients involved in our study could predispose them to another autoimmune disease in future.

Summing up, the pathogenesis of autoimmune thyroiditis is much more complex than formerly thought and relations gene-gene and gene-environment have hardly been researched. It is possible that there are many more still unknown susceptibility genes, each variant contributing just a little to the development of autoimmune thyroiditis. In our opinion, the application of knowledge about susceptibility genes is slowly entering clinical practice.

ACKNOWLEDGMENTS

This study was financially supported by the Medical University of Lodz, project No 502-12086 and statutory funds 503/1-107-03/503-11-001.

Competing interests: The authors declare that they have no competing interests.

Authors' contributions: KW-D designed the study, carried out the molecular genetic studies, participated in coordination of the study and in preparation of manuscript. KK-R, RS, AZ provided blood samples for analysis, collected from their patients. AL senior author, supervised the study and wrote the final version of manuscript. All authors have read and approved the final manuscript.

REFERENCES

- 1 Arechiga AF, Habib T, He Y, Zhang X, Zhang ZY, Funk A, *et al.* (2009). Cutting edge: the PTPN22 allelic variant associated with autoimmunity impairs B cell signalling. *J Immunol.* **182**: 3343–3347.
- 2 Downey J, Smith A, Schneider H, Hogg N, Rudd CE (2007). TCR/CD3 mediated stop-signal is decoupled in T-cells from *Ctla4* deficient mice. *Immunol Lett.* **115**: 70–72.
- 3 Effraimidis G, Wiersinga WM (2014). Mechanisms in endocrinology: autoimmune thyroid disease: old and new players. *Eur J Endocrinol.* **170**: R241–R252.
- 4 Glick AB, Wodzinski A, Fu P, Levine AD, Wald DN (2013). Impairment of regulatory T-cell function in autoimmune thyroid disease. *Thyroid.* **23**: 871–878.
- 5 Gough SC, Walker LS, Sansom DM (2005). CTLA4 gene polymorphism and autoimmunity. *Immunol Rev.* **204**: 102–115.
- 6 Hou HF, Jin X, Sun T, Li C, Jiang BF, Li QW (2015). Cytotoxic T lymphocyte-associated antigen 4 gene polymorphisms and autoimmune thyroid diseases: an updated systematic review and cumulative meta-analysis. *Int J Endocrinol.* 2015: 747–816.
- 7 Kochi Y, Myouzen K, Yamada R, Suzuki A, Kurosaki T, Nakamura Y (2009). FCRL3, an autoimmune susceptibility gene, has inhibitory potential on B-cell receptor-mediated signaling. *J Immunol.* **183**: 5502–5510.
- 8 Kochi Y, Yamada R, Suzuki A, Harley JB, Shirasawa S, Sawada T, *et al.* (2005). A functional variant in FCRL3, encoding Fc receptor-like 3, is associated with rheumatoid arthritis and several autoimmunities. *Nat Genet.* **37**: 478–485.
- 9 Kucharska AM, Gorska E, Wasik M, Demkow U (2013). Expression of cytotoxic T lymphocyte antigen-4 in T cells from children with Hashimoto's thyroiditis. *Adv Exp Med Biol.* **756**: 163–168.
- 10 Kucharska AM, Gorska E, Wasik M, Pyrzak B (2010). Decreased CD4+CD152+ T cell subset and its correlation with the level of antithyroid antibodies in children with chronic autoimmune thyroiditis. *Eur J Med Res.* **15**: 72–75.
- 11 Lee HJ, Li CW, Hammerstad SS, Stefan M, Tomer Y (2015). Immunogenetics of autoimmune thyroid diseases: A comprehensive review. *J Autoimmun.* **64**: 82–90.
- 12 Lee YH, Rho YH, Choi SJ, Ji JD, Song GG, Nath SK, *et al.* (2006). The PTPN22 C1858T functional polymorphism and autoimmune diseases – a meta-analysis. *Rheumatology (Oxford).* **46**: 49–56.
- 13 Liu HW, Xu RY, Sun RP, Wang Q, Liu JL, Ge W, *et al.* (2015). Association of PTPN22 gene polymorphism with type 1 diabetes mellitus in Chinese children and adolescents. *Genet Mol Res.* **14**: 63–68.
- 14 Matesanz-Isabel J, Sintes J, Llinàs L, de Salort J, Lázaro A, Engel P (2011). New B-cell CD molecules. *Immunol Lett.* **134**: 104–112.
- 15 Pastuszek-Lewandoska D, Domańska D, Rudzińska M, Bossowski A, Kucharska A, Sewerynek E (2013). CTLA-4 polymorphisms (+49 A/G and -318 C/T) are important genetic determinants of AITD susceptibility and predisposition to high levels of thyroid autoantibodies in Polish children – preliminary study. *Acta Biochim Pol.* **60**: 641–646.
- 16 Rieck M, Arechiga A, Onengut-Gumuscu S, Greenbaum C, Concannon P, Buckner JH (2007). Genetic variation in PTPN22 corresponds to altered function of T and B lymphocytes. *J Immunol.* **179**: 4704–4710.
- 17 Riquetto AD, de Noronha RM, Matsuo EM, Ishida EJ, Vaidergorn RE, Soares Filho MD (2015). Thyroid function and autoimmunity in children and adolescents with type 1 diabetes mellitus. *Diabetes Res Clin Pract.* **110**: e9–11.
- 18 Schneider H, Downey J, Smith A, Zinselmeyer BH, Rush C, Brewer JM, *et al.* (2006). Reversal of the TCR stop signal by CTLA-4. *Science.* **313**: 1972–1975.
- 19 Schneider H, Smith X, Liu H, Bismuth G, Rudd CE (2008). CTLA-4 disrupts ZAP70 microcluster formation with reduced T cell/APC dwell times and calcium mobilization. *Eur J Immunol.* **38**: 40–47.
- 20 Spalinger MR, Lang S, Weber A, Frei P, Fried M, Rogler G, *et al.* (2013). Loss of protein tyrosine phosphatase nonreceptor type 22 regulates interferon- γ -induced signaling in human monocytes. *Gastroenterology.* **144**: 978–988.
- 21 Swainson LA, Mold JE, Bajpai UD, McCune JM (2010). Expression of the autoimmune susceptibility gene FcRL3 on human regulatory T cells is associated with dysfunction and high levels of programmed cell death-1. *J Immunol.* **184**: 3639–3647.
- 22 Szczepańska M, Wirstlein P, Holysz H, Skrzypczak J, Jagodziński PP (2013). The FCRL3 -169T>C polymorphism and the risk of endometriosis-related infertility in a Polish population. *Arch Gynecol Obstet.* **288**: 799–780.
- 23 Tolone C, Cirillo G, Papparella A, Tolone S, Santoro N, Grandone A (2009). A common CTLA4 polymorphism confers susceptibility to autoimmune thyroid disease in celiac children. *Dig Liver Dis.* **41**: 385–389.
- 24 Tomer Y, Dolan LM, Kahaly G, Divers J, D'Agostino RB Jr, Imperatore G *et al.* (2015). SEARCH for Diabetes in Youth Study. Genome wide identification of new genes and pathways in patients with both autoimmune thyroiditis and type 1 diabetes. *J Autoimmun.* **60**: 32–39.
- 25 Wiersinga WM (2014). Thyroid autoimmunity. *Endocr Dev.* **26**: 139–157.
- 26 Wu J, Katrekar A, Honigberg LA, Smith AM, Conn MT, Tang J, *et al.* (2006). Identification of substrates of human protein-tyrosine phosphatase PTPN22. *J Biol Chem.* **281**: 11002–11010.
- 27 Zhang J, Zahir N, Jiang Q, Miliotis H, Heyraud S, Meng X (2011). The autoimmune disease-associated PTPN22 variant promotes calpain-mediated Lyp/Pep degradation associated with lymphocyte and dendritic cell hyperresponsiveness. *Nat Genet.* **43**: 902–907.
- 28 Zhao SX, Liu W, Zhan M, Song ZY, Yang SY, Xue LQ *et al.* (2013). China Consortium for the Genetics of Autoimmune Thyroid Disease. *PLoS One.* **8**: e57758.