

# Endocrine response to high intensity barbell squats performed with constant movement tempo and variable training volume

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## Abstract

**OBJECTIVE:** Research indicates that among the many elements of resistance exercise protocols, training volume and total training load are the key factors for post-exercise increase in the secretion of testosterone (T), growth hormone (GH), insulin-like growth factor (IGF-1) and cortisol (C). The aim of this study was to determine the effects of resistance exercises with variable volume and constant intensity and movement tempo on post-exercise concentrations of selected anabolic and catabolic hormones.

**MATERIALS AND METHODS:** 28 experienced powerlifters ( $27.8 \pm 2.9$  years, with  $6.64 \pm 1.29$  years of training experience, average body mass of  $85.3 \pm 3.3$  kg and body height of  $165.8 \pm 10.3$  cm) who compete at the national and international level performed three repetitions of barbell squats with a constant external load of 90% 1RM and variable volume (3, 6 and 12 sets of squats) in three stages (pre-exercise, immediately post exercise, and 1h after exercise) over three consecutive weeks. Venous blood samples (10ml) were collected from the antecubital vein, to determine pre- and post-exercise values of the following variables T, GH, IGF-1, C, at rest, immediately after the cessation of the last set of squats, and after 60 minutes of recovery.

**RESULTS:** The T test showed that performing 6 and 12 sets resulted in increases of post exercise GH ( $p < 0.01$ ). Performing 6 sets of squats resulted in post exercise decrease ( $p < 0.01$ ) in IGF-1 and C. Performing 3 sets of squats resulted in immediate post exercise decrease of IGF-1 ( $p < 0.01$ ), which was not maintained 1h after exercise. There were no other significant differences in analysed variables, with the training volume of three sets of three repetitions, confirming previous data suggesting that low volume is the limiting factor in increased post-exercise secretion.

**CONCLUSION:** This study demonstrated that in terms of endocrine response, the optimal volume of high intensity strength exercise is six sets. Therefore, intentionally high volume (12 sets) or low volume (3 sets) are not an effective stimuli for endocrine responses of trained individuals. The 6 sets of squats seems to drive

hormonal responses of GH, C and IGF-1, which may play a significant role in stimulating muscle growth and tissue regeneration.

## INTRODUCTION

Resistance exercises, especially those performed with high intensity, cause significant endocrine changes, both acute and chronic (Kraemer & Ratamess, 2005; Uchida et al. 2009; Crewther et al. 2011). The endocrine system is particularly sensitive to resistance exercise and changes in anabolic and catabolic hormones have been associated with the process of post-exercise rebuilding of damaged muscle cells, and thus the magnitude and rate of the post-exercise adaptation (Kraemer & Ratamess, 2005; Sedliak et al. 2007; Kadi, 2008). Hormones, particularly growth hormone (GH) and testosterone, have a significant effect on the rate of protein synthesis, and the type of substrates metabolized during and immediately after exercise (Uchida et al. 2009). These hormones also stimulate the activation and proliferation of satellite cells which facilitate myofibrillar hypertrophy (Kadi et al. 2005). GH has an anabolic effect on skeletal muscles, stimulates the synthesis of proteins, facilitates the transport of amino acids into skeletal muscles, thus affecting hypertrophy of both Type I and Type II muscle fibres (Hansen et al. 2001). Research has demonstrated that blocking the effects of anabolic hormones reduces the rate of adaptive changes and the effectiveness of weight training programs (Kvorning et al. 2006). These hormones play a significant role in mediating increases in muscle mass and muscle strength (Kadi, 2008).

Some research indicates that among the many variables of strength training protocols, training volume and total training load are key factors for post-exercise increase in the secretion of various hormones (Kraemer et al. 1991). Other research results indicate that duration of the force production and the length of rest periods between sets are the most significant factors stimulating plasma and serum cortisol (C). These hormonal responses are particularly intense in the case of high-intensity, medium or high volume training programs (Kraemer et al. 1991) and when the training protocol targets large muscle groups (Kraemer & Ratamess, 2005). Research on the effects of strength training on muscle hypertrophy showed an important role not only of anabolic hormones like testosterone (T), but also for growth factors, including the insulin-like growth factor-1 (IGF-1). Serum IGF-1 elevations are induced by strength training (Kraemer & Ratamess, 2005), but some studies suggested that this is the case only when resting concentrations are low (Kraemer et al. 1991). The divergent findings concerning the effect of strength training on the process of adaptation and response of the endocrine system may result from the fact that most procedures did not specify the movement speed for an exercise or the whole strength training session. Only a few publications have analysed the effects

of movement tempo (cadence) on adaptive processes in terms of strength, power, muscle hypertrophy or endocrine responses (Wilk et al. 2018a; Headley et al. 2011; Hatfield et al. 2006; Sakamoto and Sinclair 2006; Hunter et al. 2003; Keeler et al. 2001; Westcott et al. 2001). Repetition speed is the only variable which has not been widely explored scientifically with respect to adaptation and response of the endocrine system. In most studies, the tempo of performing strength exercises is volitional, according to the natural movement rhythm. Studies have found that the lower the movement speed the more intensive decline in the generated muscle force (Hutchins 1993, Westcott et al. 2001, Kraemer et al. 2002). Wilk et al. (2018b) showed that the movement tempo in strength training impacts training volume, both in terms of repetitions and total time under tension (TUT). The optimal volume and intensity of training loads in resistance exercises that may most effectively stimulate the anabolic hormones while diminishing the secretion of catabolic ones has not been determined. This may be due to numerous factors such as movement speed for an exercise, age, sex, training experience, type of muscular contractions used which complicate this issue. Additional factors include type of equipment, diet, supplementation and how these factors interact with genetic endowment (Wilk et al. 2018c). Exceeding the optimal training volume causes the anabolic hormone peak to occur during training, and continuation of exercise results in an excess concentration of catabolic hormones (Virus & Virus, 2004; Uchida et al. 2009; West et al. 2012).

The aim of this study was to determine the effect of variable volume in squat exercise with constant intensity and constant tempo on post-exercise concentrations of selected anabolic and catabolic hormones and growth factors (GH, T, IGF-1 and C). An additional objective was to determine the range of training volume, which elicited the greatest anabolic hormone secretion while limiting the increase in C.

## MATERIALS AND METHODS

### *Experimental Approach to the Problem*

All testing was performed in the Strength and Power Laboratory at the Jerzy Kukuczka Academy of Physical Education in Katowice. The experiment was performed following a randomized cross sectional design, where each participant performed a familiarization session with a 1-RM test and three different testing protocols a week apart. During the experimental sessions, subjects performed barbell squats at low volume - 3 sets (LV<sub>3</sub>); medium volume - 6 sets (MV<sub>6</sub>); high volume- 12 sets (HV<sub>12</sub>). In each set 3 repetitions were done using 90% 1RM and a 2/0/3/0 tempo. Subjects were required to refrain from resistance training 72 hours prior to each experimental session, were familiarized with the exercise protocol and were informed about the benefits

and risks of the research before expressing their consent for participation in the experiment.

### Subjects

Participants for this study were 28 experienced powerlifters who competed at the national and international level. The age of the subjects was  $27.8 \pm 2.9$  years, with  $6.64 \pm 1.29$  years of training experience, average body mass of  $85.3 \pm 3.3$  kg and body height of  $165.8 \pm 10.3$  cm. The participants were allowed to withdraw from the experiment at any moment and were free of injuries. The study protocol was approved by the Bioethics Committee for Scientific Research, at the Academy of Physical Education in Katowice, Poland, according to the ethical standards of the Declaration of Helsinki, 2013. Participants were instructed to maintain their normal dietary habits over the entire study period and did not use any dietary supplements or stimulants for the duration of the study.

### Procedures

#### Familiarization session and one repetition maximum test

The participants arrived at the laboratory at the same time of day (in the morning between 09:00 and 11:00) and cycled on an ergometer for 5 minutes at an intensity that resulted in a heart rate of around 130 bpm, then performed a general lower body warm-up. Next, the participants completed 15, 10, 5 and 3 barbell squat repetitions using 20%, 40%, 60%, 80% of their estimated 1RM using a 2/0/2/0 cadence. Knee wraps were allowed and three spotters were present at all times during the testing protocol. The participants then executed single repetitions using a volitional cadence with 5 min of rest between successful trials. The load for each subsequent attempt was increased by 5 kg, and the process was repeated until failure.

#### Experimental sessions

The participants arrived at the laboratory in the morning (09:00 to 11:00 am). After completing the same warm-up as in the familiarization session, they performed 3 sets (LV<sub>3</sub>), 6 sets (MV<sub>6</sub>) or 12 sets (HV<sub>12</sub>) of the squat with 90% 1RM (Table 1) using 2/0/3/0 metronome guided cadence (Korg MA-30, Korg, Melville, New York, USA). The time between experimental sessions of training was one week. The participants were verbally encouraged throughout all testing sessions. All repetitions were performed without intentionally pausing at the transition between the eccentric and concentric phases.

#### Blood sampling and analysis

During the experiment, 10 ml venous blood samples were collected from the antecubital vein to determine pre- and post-exercise concentrations of T, GH, IGF-1, and C at rest, immediately after the cessation of the last set of squats, and after 60 minutes of recovery. Commercially available radioimmunoassay evaluations

**Table 1.** The testing protocols applied during the experiment

	Low volume (LV <sub>3</sub> )	Medium volume (MV <sub>6</sub> )	High volume (HV <sub>12</sub> )
Load (%1RM)	90%1RM	90%1RM	90%1RM
Tempo	2/0/3/0	2/0/3/0	2/0/3/0
Set / repetition	3 / 3	6 / 3	12 / 3
Rest interval between sets	5 min	5 min	5 min

were performed for the evaluation of T (DSL-4000), GH (DSL-1900), IGF-1 (DSL-2800), and cortisol (DSL-2100). The ICC for the biochemical analysis varied from 0,88 to 0,99 for the 4 conducted test.

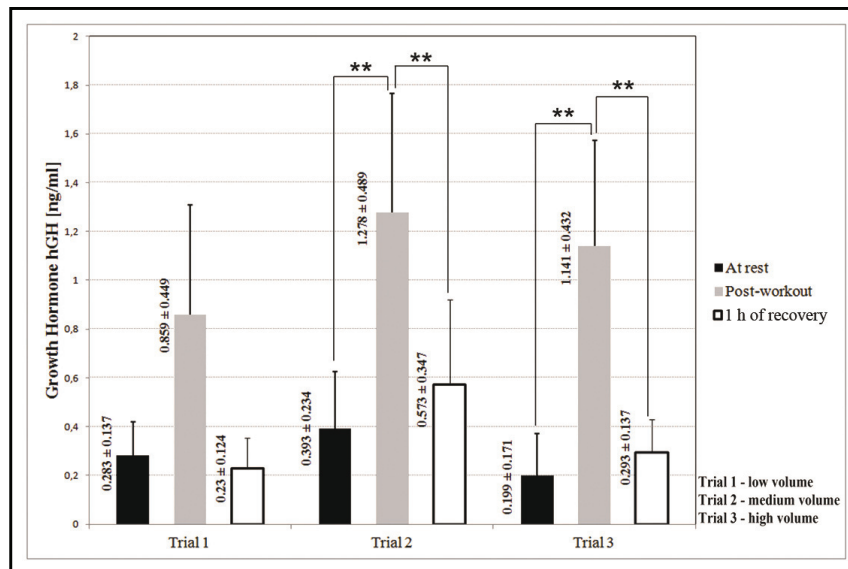
### Statistical Analyses

Means, standard deviations, confidence levels and standard errors were calculated for all measured variables, and all variables were tested for normality by the quantile-quantile test. To identify significant group by time interactions, t test for independent trials was used for each dependent variable. When a significant interaction occurred post hoc test by Rodger's method was performed for detecting differences among groups (pair wise comparisons). Rodger's method belongs to the most powerful post-hoc tests for detecting differences among groups. This test protects against loss of statistical power as the degrees of freedom increase. The statistical significance was set at  $p \leq 0.05$ .

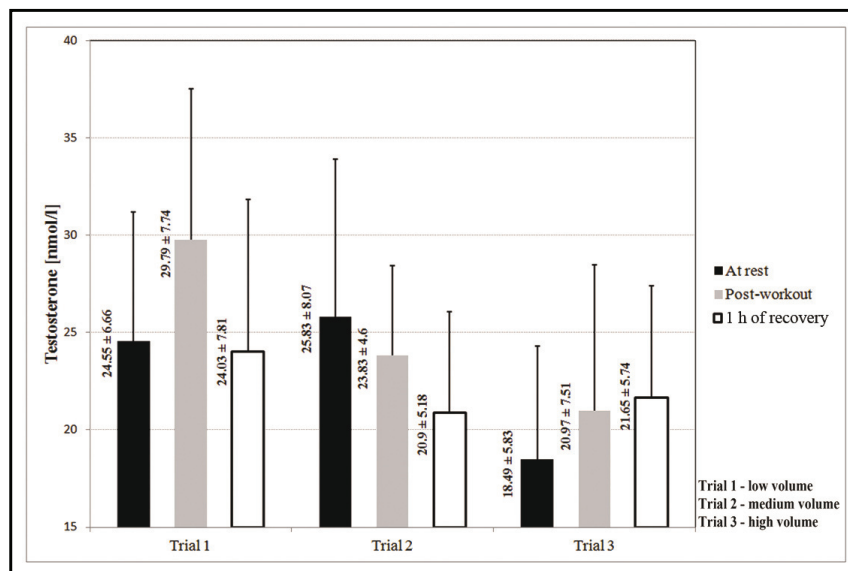
## RESULTS

All variables were normally distributed as determined by the quantile-quantile test results ( $p > 0.05$ ). Among 28 experienced powerlifters ( $N = 28$ ), there were statistically significant differences of GH concentration in MV<sub>6</sub>, between the mean post-workout values ( $1.278 \pm 4.89$  ng/ml) and the mean rest value ( $0.393 \pm 0.234$  ng/ml)  $t(7) = 5.87$ ,  $p \leq .01$ , as well as between the value obtained one hour after exercise ( $0.573 \pm 0.347$  ng/ml)  $t(7) = 5.01$ ,  $p \leq .01$ . Statistically significant difference also occurred in GH during HV<sub>12</sub>, between the mean post-workout value ( $1.141 \pm 0.432$  ng/ml) and the mean rest value ( $0.199 \pm 0.171$  ng/ml)  $t(7) = 6.15$ ,  $p \leq .01$ , as well as the value obtained one hour after exercise ( $0.293 \pm 0.137$  ng/ml)  $t(7) = 6.07$ ,  $p \leq .01$ . Therefore, we reject the null hypothesis that there is no difference in concentration of growth hormone at rest and post-workout in MV<sub>6</sub> and HV<sub>12</sub> training. For LV<sub>3</sub> training, GH concentrations were not significantly different between mean values at rest, post-exercise and after one hour of recovery when LV<sub>3</sub> was applied. Significant differences were found just for GH with MV<sub>6</sub>, and HV<sub>12</sub> (Figure 1).

With regard to testosterone, no significant differences in concentration were observed as a result of training volume at any time point. Therefore, we fail to



**Fig. 1.** The average concentration of the growth hormone at various levels of training volume. \*\* $p < 0.01$ .



**Fig. 2.** The average concentration of the testosterone at various levels of training volume.

reject the null hypothesis that there is no difference in concentration of testosterone between training volume at each time point (Figure 2).

For IGF-1, there were no significant differences in HV<sub>12</sub> between the mean rest concentration and the post-exercise value. In the moderate volume trial, significant differences were found between the mean rest value ( $657.29 \pm 205.36$  ng/ml) and the value obtained one hour after exercise ( $534.77 \pm 102.3$  ng/ml)  $t(7) = 3.10$ ,  $p \leq .01$ . Statistically significant differences also occurred in LV<sub>3</sub>s, between post-exercise value ( $476.43 \pm 197.82$  ng/ml) and the mean rest value ( $573.42 \pm 169.76$  ng/ml)  $t(7) = 3.69$ ,  $p \leq .01$ . Therefore, we reject the null hypothesis that there is no difference in concentration of the IGF-1 between values at rest and post-exercise in LV<sub>3</sub>s, as well as between rest and after one hour of recovery in MV<sub>6</sub>s (Figure 3).

For C, only the MV<sub>6</sub> trial yielded significant differences between the concentrations at rest ( $673.76 \pm 251.32$  nmol/l) and after one hour of recovery ( $320.28 \pm 114.17$  nmol/l)  $t(7) = 6.89$ ,  $p \leq .01$ , as well as post-exercise ( $479.54 \pm 218.24$  nmol/l)  $t(7) = 4.17$ ,  $p \leq .01$ . Therefore, we reject the null hypothesis that there is no difference in concentration of the cortisol between training volume at every time point (Figure 4).

## DISCUSSION

The main finding of this study is that the different training volume (LV<sub>3</sub>, MV<sub>6</sub>, HV<sub>12</sub>) with constant movement tempo in resistance exercise doesn't impact on post-exercise concentrations of T, only MV<sub>6</sub> and HV<sub>12</sub> can elicit anabolic GH post-exercise response, and only HV<sub>6</sub> can elicit post-exercise IGF-1 response

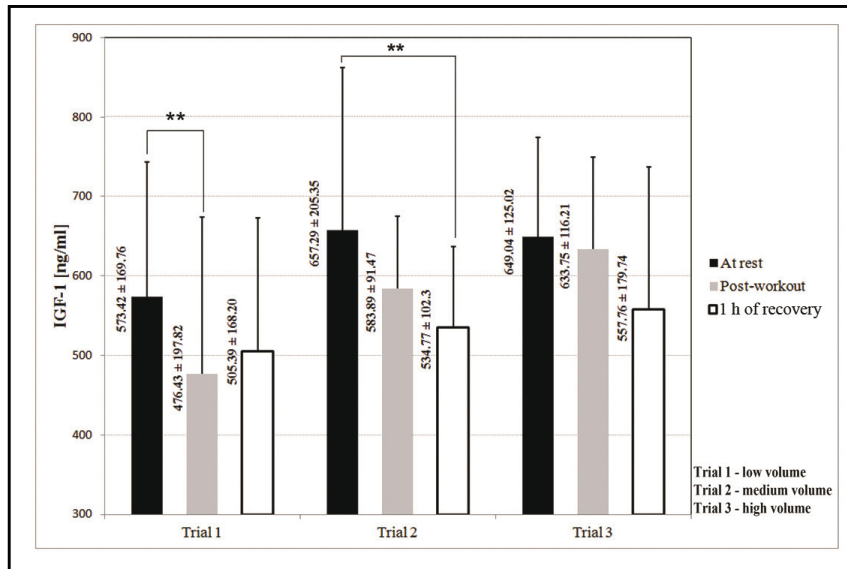


Fig. 3. The average concentration of the IGF-1 at various levels of training volume. \*\*p<0.01.

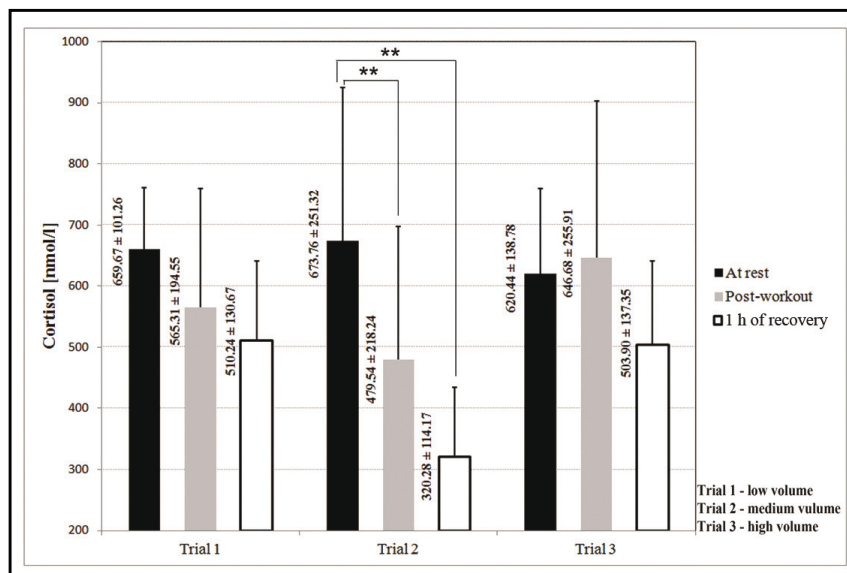


Fig. 4. The average concentration of the cortisol at various levels of training volume. \*\*p<0.01.

with decreased post-exercise C level. Therefore, the MV<sub>6</sub> resulted in most efficient hormonal post-exercise response in terms of post exercise recovery.

Despite our use of variable-volume of exercise (LV<sub>3</sub>, MV<sub>6</sub>, HV<sub>12</sub>) in the present study, no significant differences in post-exercise plasma testosterone levels were observed, contrary to previous findings (Crewther *et al.* 2008). While Kraemer and Ratamess (2005) suggested that strong increases in serum testosterone levels occur in participants with a relatively high baseline levels of this hormone, this was not confirmed by the results of the present study. The absence of significant differences between resting and post-exercise values may have resulted from high initial resting testosterone levels in these young participants (mean age 24), extensive training experience (mean of 6 years), and/or time of day of sampling (9 am). It is possible

that circadian rhythm changes masked exercise-related changes of in testosterone (Sedliak *et al.* 2007; Cook and Crewther, 2012). It is also possible that the lack of significant differences in post-exercise testosterone levels could have been due to the length of rest periods between sets (5 minutes), since Kraemer *et al.* (1991) suggested that the length of rest periods between sets determines the effective impact of strength training on the elevation of testosterone levels and should not exceed two minutes. Kraemer *et al.* (1991) also suggested that exercise-induced changes in testosterone may be influenced by the type of exercise performed. While an increase in plasma testosterone levels was observed when powerlifting exercises targeting several muscle groups were performed at the same time (Kraemer *et al.* 1991), the present study did not find

this association with just one exercise being used (back barbell squat), even at varying exercise volumes.

Strength training which results in a significant increase in C levels generally involves a higher number of repetitions than three and significantly shorter rest intervals between sets than were performed in the present study (Smilios *et al.* 2003). Our findings suggest that a five-minute rest period between sets, despite a large number of sets performed at 90% of the 1RM load, does not stimulate glycolysis significantly, and thus does not result in a significant increase in C levels. Research showed that when the resting concentration of C is high, no post-exercise elevation was found (Beaven *et al.* 2008), and in some cases a post-exercise decrease in concentrations of this hormone was observed, compared to resting levels. This study demonstrated a significant difference in cortisol concentrations only in the MV<sub>6</sub> where exercise-induced C levels were significantly lower than baseline values prior to exercise.

The present study demonstrated that the volume of 12 sets (HV<sub>12</sub>) did not result in a greater GH secretion in comparison to the LV<sub>3</sub> and MV<sub>6</sub>. Thus, we suggest that the unfavorable increase in C levels associated with a higher volume indicates that the MV<sub>6</sub> could be more beneficial with respect to post-exercise endocrine adaptation. Previous research suggested that when the optimal volume is exceeded, the GH peak occurs already during the training session (Schwarz *et al.* 1996; West *et al.*, 2012). It is possible that this was the case during the maximal volume protocol (HV<sub>12</sub>), yet no measurements were performed between particular sets to confirm this hypothesis.

Analyses of changes in IGF-1 concentrations during our different strength training protocols demonstrated decreases in the LV<sub>3</sub> and MV<sub>6</sub> trials, and despite comparable training variables, the results we obtained were contradictory to those reported in earlier research (Kraemer *et al.* 1991). Some research results indicate that when anabolic processes in the body are predominant, strength training stimulates the exercise-induced elevation of IGF-1 concentrations, as demonstrated by previous research Forbes *et al.* (1989). The results of the present research, may be indicative of the predominance of catabolic environment in the subjects (with exception of decreased C level in MV<sub>6</sub>), which could partly explain the absence of exercise-induced increase in IGF-1 levels. It is known that the metabolic state of the body and the level of target cell sensitivity to the released IGF-1 is the essential stimulating mechanism for changes in IGF-1 concentrations (Ambrosio *et al.* 1996). In this study, a significant elevation of IGF-1 concentrations was not observed, and even decreased significantly after the low volume training protocol (Figure 3). A significant decrease also occurred between the concentrations measured prior to exercise and after one hour of recovery, in the moderate training volume trial. Resting IGF-1 levels were high in this

study, what confirms a previous hypotheses that the exercise-induced elevation of IGF-1 levels are more likely to be observed when baseline concentrations are low (Kraemer *et al.* 1991). The high resting concentration of IGF-1 may be associated with the effects of nocturnal GH secretion (Ohlsson *et al.* 2009). The measurement of resting IGF-1 concentration was performed at approximately 9 am. Research showed that the duration of IGF-1 secretion due to the influence of GH can be approximately 12 hours long (Kraemer & Ratamess, 2005), which may partly explain the high resting IGF-1 concentrations.

A negative correlation between the concentrations of cortisol and testosterone has been reported (Brownlee *et al.* 2005). The study by Brownlee *et al.* (2005) also show a positive correlation between concentrations of cortisol and free testosterone. In addition to the effects of testosterone, the important post-training role of GH or IGF-1 should be taken into account. Anabolic hormones were identified as having a significant impact on muscle tissue remodelling (Virus & Virus, 2004). Hansen *et al.* (2001) suggested that the adaptation is dependent on the exercise-induced concentrations of anabolic hormones, therefore optimizing the volume may be crucial for maximal training effects. Research suggests that the first hour of recovery is critical for the endocrine response. After this period, the concentration of hormones and growth factors generally returns to resting levels (Tremblay *et al.* 2005; West *et al.* 2014) which was confirmed by the present study but not in IGF-1 and cortisol. The results of global research also indicate that there are significant differences in individual hormonal responses to specific types of exercise, (McGuigan *et al.* 2004; Beaven *et al.* 2008).

## PRACTICAL APPLICATION

This study demonstrated that in terms of anabolic hormone response, the most effective volume is close to 6 sets. It has been established that performing 12 sets resulted in an increase of cortisol concentrations, while 6 sets led to a significant decrease in exercise-induced cortisol levels compared to baseline. We believe it is reasonable to suggest the volume of training which should not be exceeded, since our data did not demonstrate any favorable changes in hormone response with higher volume training in experienced powerlifters.

## CONCLUSIONS

This research indicates that among the many variables of strength exercise, training volume and total training load are the key factors stimulating the secretion of various hormones, both anabolic and catabolic. The conducted study demonstrated that in terms of endocrine response, the optimal volume of high intensity strength exercise is about 6 sets in experienced powerlifters.

## ACKNOWLEDGEMENTS

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## REFERENCES

- Ambrosio MR, Valentini A, Trasforini G, Minuto F, Ghigo E, Cella S, Margutti A, Pansini R, & Degli Uberti EC (1996). Function of the GH/IGF-1 axis in healthy middle-aged male runners. *Neuroendocrinology*. **63**(6): 498–503.
- Beaven CM, Gill ND & Cook CJ (2008). Salivary testosterone and cortisol responses in professional rugby players after four resistance exercise protocols. *J Strength Cond Res*. **22**(2): 426–32.
- Brownlee KK, Moore AW, & Hackney AC, (2005). Relationship between circulating cortisol and testosterone: Influence of physical exercise. *J Sports Science and Med*. **4**: 76–83.
- Cook CJ, & Crewther BT (2012). Changes in salivary testosterone concentrations and subsequent voluntary squat performance following the presentation of short video clips. *Horm Behav*. **61**(1): 17–22.
- Crewther BT, Cronin J, Keogh J & Cook C (2008). The salivary testosterone and cortisol response to three loading schemes. *The J Strength Cond Res*. **22**(1): 250–255.
- Crewther BT, Heke T & Keogh JW (2011). The effects of training volume and competition on the salivary cortisol concentrations of Olympic weightlifters. *J Strength Cond Res*. **25**(1): 10–15.
- Forbes GB, Brown MR, & Welle SL (1989). Underwood LE. Hormonal response to overfeeding. *Am J Clin Nutr*. **49**(4): 608–611.
- Hansen S, Kvorning T, Kjaer M, & Sjogaard G (2001). The effect of short-term strength training on human skeletal muscle: the importance of physiologically elevated hormone levels. *Scand J Med Sci Sports*. **11**(6): 347–354.
- Hatfield DL, Kraemer WJ, Spiering BA, Häkkinen K, Volek JS, Shimano T, Spreuwenberg LP, Silvestre R, Vingren JL, Fragala MS, Gomez AL, Fleck SJ, Newton RU, Maresh CM (2006). The impact of velocity of movement on performance factors in resistance exercise. *J Strength Cond Res*. **20**(4): 760–766.
- Headley SA, Henry K, Nindl BC, Thompson BA, Kraemer WJ, Jones MT (2011). Effects of lifting tempo on one repetition maximum and hormonal responses to a bench press protocol. *J Strength Cond Res*. **25**: 406–13.
- Hunter GR, Seelhorst D, Snyder S (2003). Comparison of metabolic and heart rate responses to super slow vs. traditional resistance training. *J Strength Cond Res*. **17**: 76–81.
- Hutchins K (1993). *Super slow: the ultimate exercise protocol*. 2nd ed. Casselberry, FL: Media Support
- Kadi F, Charifi N, Denis C, Lexell J, Andersen JL, Schjerling PO, & Kjaer M (2005). The behaviour of satellite cells in response to exercise: what have we learned from human studies? *Pflügers Archiv*. **451**(2): 319–327.
- Kadi F (2008). Cellular and molecular mechanisms responsible for the action of testosterone on human skeletal muscle. A basis for illegal performance enhancement. *Br J Pharmacol*. **154**: 522–528.
- Keeler LK, Finkelstein FH, Miller W, Fernhall B (2001). Early-Phase Adaptations of Traditional-Speed vs. Superslow Resistance Training on Strength and Aerobic Capacity in Sedentary Individuals. *J Strength Cond Res*. **15**(3): 309–314.
- Kraemer WJ, Gordon SE, Fleck SJ, Marchitelli LJ, Mello R, Dziadosz JE, Friedl E, Harman E, Maresh C, & Fry AC (1991). Endogenous anabolic hormonal and growth factor responses to heavy resistance exercise in males and females. *Int J Sports Med*. **12**(2): 228–235.
- Kraemer WJ, Fry AC, Warren BJ, Stone MH, Fleck SJ, Kearney JT, Conroy BP, Maresh CM, Weseman A, & Triplett NT (1992). Acute hormonal responses in elite junior weightlifters. *Int J Sports Med*. **13**(2): 103–109.
- Kraemer WJ, Koziris LP, Ratamess NA, Hakkinen K, Triplett-McBride NT, Fry AC, Gordon SE, Volek JS, French DN, Rubin MR, Gomez AL, Sharman MJ, Michael Lynch J, Izquierdo M, Newton RU, Fleck SJ (2002). Detraining produces minimal changes in physical performance and hormonal variables in recreationally strength-trained men. *J of Strength Cond Res*. **16**: 373–382
- Kraemer WJ, & Ratamess NA (2005). Hormonal responses and adaptations to resistance exercise and training. *Sports Medicine*, **35**: 339–361.
- Kvorning T, Andersen M, Brixen K, & Madsen K (2006). Suppression of endogenous testosterone production attenuates the response to strength training: a randomized, placebo-controlled, and blinded intervention study. *Am J Physiol Endocrinol Metabol*. **291**(6): 1325–1332.
- McGuigan MR, Egan AD & Foster C (2004). Salivary cortisol responses and perceived exertion during high intensity and low intensity bouts of resistance exercise. *J Sports Sci Med*. **3**: 8–15.
- Ohlsson C, Mohan S, Sjorgen K, Tivesten A, Isgaard J, Isaksson O, Jansson J, & Svensson J, (2009). The role of liver-derived insulin-like growth factor-I. *Endocr Rev*. **30**: 494–535.
- Sakamoto A, Sinclair PJ (2006). Effect of movement velocity on the relationship between training load and the number of repetitions of bench press. *J Strength Cond Res*. **20**(3): 523–7.
- Schwarz AJ, Brasel JA, Hintz RL, Mohan S, & Cooper DM (1996). Acute effect of brief low- and high-intensity exercise on circulating insulin-like growth factor (IGF) I, II, and IGF-binding protein-3 and its proteolysis in young healthy men. *J Clin Endocrinol Metabol*. **81**: 3492–3497.
- Sedliak M, Finni T, Cheng S, Kraemer WJ, & Hakkinen K (2007). Effect of time-of-day-specific strength training on serum hormone concentrations and isometric strength in men. *Chronobiol Int*. **24**(6): 1159–1177.
- Smilios I, Piliandis T, Karamouzis M, & Tokmakidis SP (2003). Hormonal responses after various resistance exercise protocols. *Med Sci Sports Exerc*. **35**(4): 644–654.
- Trembley MS, Copeland JL, & Van Helder W (2005) Influence of exercise duration on post-exercise steroid hormone responses in trained males. *Eur J Appl Physiol*. **94**(5–6): 505–513.
- Uchida MC, Crewther BT, Ugrinowitch C, Bacurau RF, Moriscot AS, & Aoki MS (2009). Hormonal responses to different resistance exercise schemes of similar total volume. *J Strength Cond Res*. **23**(7): 2003–2008.
- Westcott WL, Winett RA, Anderson ES, Wojcik JR, Loud RL, Clegggett E, Glover S (2001). Effects of regular and slow speed resistance training on muscle strength. *J Sports Med Phys Fitness*. **41**(2): 154–8.
- West DW, Cotie LM, Mitchell CJ, Churchward-venne TA, MacDonald MJ, & Phillips SM (2012). Resistance exercise order does not determine postexercise delivery of testosterone, growth hormone, and IGF-1 to skeletal muscle. *Appl Physiol Nutr Metab*. **38**(2): 220–226.
- West DJ, Finn CV, Cunningham DJ, Shearer DA, Jones MR, Harrington BJ, Crewther BT, Cook CJ, & Kilduff LP (2014). Neuromuscular function, hormonal, and mood responses to a professional rugby union match. *J Strength Cond Res*. **28**(1): 194–200.
- Wilk M, Stastny P, Golas A, Nawrocka M, Jelen K, Zajac A, Tufano (2018a). Physiological responses to different neuromuscular movement task during eccentric bench press. *Neuro Endocrinol Lett*. **39**(1): 26–32
- Wilk M, Golas A, Stastny P, Nawrocka M, Krzysztofik M, Zajac A (2018b). Does tempo of resistance exercise impact training volume? *J Hum Kinet*. **62**: 241–250
- Wilk M, Michalczyk M, Golas A, Krzysztofik M, Maszczyk A, Zajac A (2018c). Endocrine responses following exhaustive strength exercise with and without the use of protein and protein-carbohydrate supplements. *Biology of Sport*. **35**(4):xx-xx doi. org/10.5114/biolsport2018.75754
- Viru A & Viru M (2004). Cortisol - Essential adaptation hormone in exercise. *Int J Sports Med*. **25**: 461–464.