

Daytime pineal gland activation in rats with colon tumors induced by 1,2-dimethylhydrazine.

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Abstract

OBJECTIVES: Intact rats and rats with 1,2-dimethylhydrazine induced tumors of large intestine were used in experiments. Previously, blood melatonin concentration in these tumor-bearing rats was shown to increase at night, but not in the daytime. **METHODS:** The extracellular microelectrode registration of rat daytime pineal gland activity was performed. **RESULTS:** The existence of different types of pinealocytes in the pineal gland was confirmed. Tumor-bearing rats, in comparison to intact, demonstrated higher spike frequency due to cells switching from regular to pattern (4-6 times gain) activity and appearance of "fast" cells (>5Hz frequency). **CONSLUSIONS:** The literature about pinealocytes points to the correlation between electrical and secretory processes in pinealocytes; thus we suppose the groups of interacting cells, detected in tumor-bearing rats, to reflect cascade cells activation while pineal gland secretion increases. The results indicate, that in the daytime pinealocytes are secreting substances (not melatonin) in dependence with hormonal background.

Introduction

As photoneuroendocrine transducer the pineal gland modulates many of the physiological systems activity according to circadian rhythms. It is a part of the system controlling adaptive reactions of organism on various conditions of environment [27, 28]. Pineal indolamine and peptide hormones influence immune functions [5, 7, 18]. Melatonin, in particular, increases immune memory while T-dependent antigen immunizes and stimulates the antibody production. Pinealectomy or constant light regime, which suppresses pineal activity, promotes tumor processes, whereas melatonin injections resulted in the decrease of carcinogenesis [2, 3, 4, 6]. Many investigators attach too much importance to melatonin, ignoring the other pineal hormones and pineal function in the daytime. But except melatonin, the pineal gland produces the multitude of peptides. These peptides have a wide range of activity. In particular, they are able to suppress RNA synthesis in tumor cells [22] and to selectively modulate DNA transcription [21].

Microelectrode registration of action potentials, which are correlated with exocytosis in neurosecretory cells [9, 10, 20], allows us to estimate exocytosis intensity in pinealocytes. Active nighttime pineal melatonin secretion and electrical activity are usually supposed to be unquestionably connected. But, in the daytime pineal melatonin production is known to be low; thus the daytime electrical activity of the pineal gland does not reflect its secretion. Melatonin is a highly lipophilic substance and can pass the cell membrane freely; thus its release is probably not connected with the excretion of secretory vesicles, and, consequently, not connected with the cellular membrane electrical processes. It can be confirmed with the following observations. Elimination of action potential-like spikes with exposure to Ca^{2+} -binding substances like verapamil or nifedipine or removal of extracellular calcium did not substantially affect melatonin production [19]. But, the pineal peptide and serotonin secretion seem to be excreted with an ordinary exocytosis, which in neurosecretory cells is usually accompanied with membrane depolarization [9, 10, 20]. Furthermore, we previously have noticed the correlation between daytime pinealocyte secretion (an accumulation of secretory vesicles with peptide content) and their electrical activity [12].

The aim of this work was to corroborate the hypothesis about involving the pineal gland in inhibition of tumor progression. Secretory activity of pineal cells was estimated according to their electrical activity in intact animals and in tumor-bearing rats.

Methods

Wistar rats, bred in the N. N. Petrov Research Institute of Oncology (St. Petersburg) were used in experiments. The study was performed according to FELASA guidelines (Category C). Two to three month aged rats were 5 times weekly subcutaneously injected with 1.2-dimethylhydrazine dihydrochloride (DMH, Sigma, USA) in dose (calculated to base) of 21mg/kg or 0.5ml 0.9% NaCl for intact. Solutions were made *ex tempore* and neutralized with sodium bicarbonate (pH= 7.0). Four to six months later such DMH dose inevitably caused adenocarcinoma in the large intestine. Rats were kept in 12:12 day/night rhythm, fed with standard laboratory fodder and had a free access to water.

Four months after the last DMH injection, 5 injected and 5 intact rats were used for electrophysiological investigation. Experiments were performed in daytime from 13:00 to 15:00, because pineal melatonin production is known to be lower in the daytime. We suppose that it would be easier to detect an activation of peptide secretion on such background. Urethane-anesthetized rats (1.2 g/kg body weight, i.p.) were fixed in stereotaxic frame in screened and grounded camera. Dorsal surface of the brain was exposed by craniotomy with a special milling cutter. Saggital sinus was ligated and cut. Glass microelectrodes with a tip diameter of 10–30 μm (resistance 4–6M Ω) filled with 3M NaCl were visually positioned at dorsal surface of the pineal gland. Action potentials were extracellularly recorded from the superficial part of the pineal gland and stored on FM-tape for off-line computer analysis. Parameters of single cells and intercellular interactions were investigated (>160 cells studied).

Extracellular microelectrode registration allows to simultaneously record discharging of 1–10 cells. We have developed the software for analyzing such records and for extracting the “voices” of separate cells. For this purpose an amplitude, length of depolarization and repolarization and polarity were measured for each spike; regular and pattern forming groups of spikes were sought for. The search of interacting cells was performed. The basic principles of this analysis [8] are widely used by many authors for investigation of neuronal structures.

After electrophysiological investigation the intestine was removed and prepared to measure tumor square. Correlation between medium frequency of pineal cells discharging and tumor square was calculated.

Results and discussion

“Slow,” with spike frequency $< 2\text{Hz}$, and “fast,” with spike frequency $> 4\text{Hz}$, type of pineal cells activity were revealed in this study. “Slow” types of activity were presented by regular, irregular and pattern spikes (Fig. 1). Electrophysiological parameters of pinealocyte activity in intact animals and in tumor induced rats are presented in Table 1. Intact rats usually demonstrated only “slow” types of activity (frequency $0.86 \pm 0.49\text{Hz}$, regularly and irregularly discharging cells).

Data obtained allow us to suppose comparatively low pineal peptide secretory activity in intact rats. Previously the same conclusion about daytime low pineal secretion of intact animals was made during ultrastructural cell study with electron-microscopy, where we have observed insignificant number of vesicles of different nature in pinealocytes of intact as compared to osmotic stress-subjected rats where a great deal of peptide-containing vesicles were present in cytoplasm and in cell processes [13].

In colon tumor-bearing animals, pineal cells with pattern activity were revealed. This type of activity was absent in intact rats. In tumor-bearing rats

“slow” cells increased spike frequency 4–6 times due to switching to pattern discharging, “fast” type of activity also appeared ($5.71 \pm 0.73\text{Hz}$) (Fig. 2).

Pattern activity was supposed to appear regularly in discharging cells during their secretory activation. This assumption was confirmed with the following observations:

1. reversible switching between regular and pattern discharging in one cell;
2. variable number of spikes in a pattern;
3. frequency of patterns and frequency of regularly discharging cells are close and belong to “slow” type of activity;
4. longitude of single spike coincides in “pattern” and in “regular” cells.

The results of our electrophysiological study of intact and colon tumor-bearing animals are concordant with the existence of several cellular types in the pineal gland [26]. It is interesting, because for a long period only two type of pineal cells—“light” and “dark”—were supposed to be presented in the gland. Recently several researchers [11] have identified more types of pineal cells, namely light, dark,

Table 1. Comparison of different type pineal cells in intact and tumor-bearing rats

The type of activity	Intact rats	Tumor-bearing rats
“Slow type” (frequency of spikes $< 2\text{Hz}$)	regular single spikes ($0.86 \pm 0.49\text{Hz}$) and irregular spikes	regular pattern activity (1.43 ± 0.35 patterns/sec; 4–6 spikes in a pattern) and regular single spikes
“Fast type” (frequency of spikes $> 4\text{Hz}$)	usually absent	regular spikes ($5.71 \pm 0.73\text{Hz}$), sometimes short patterns (2–3 spikes in a pattern).

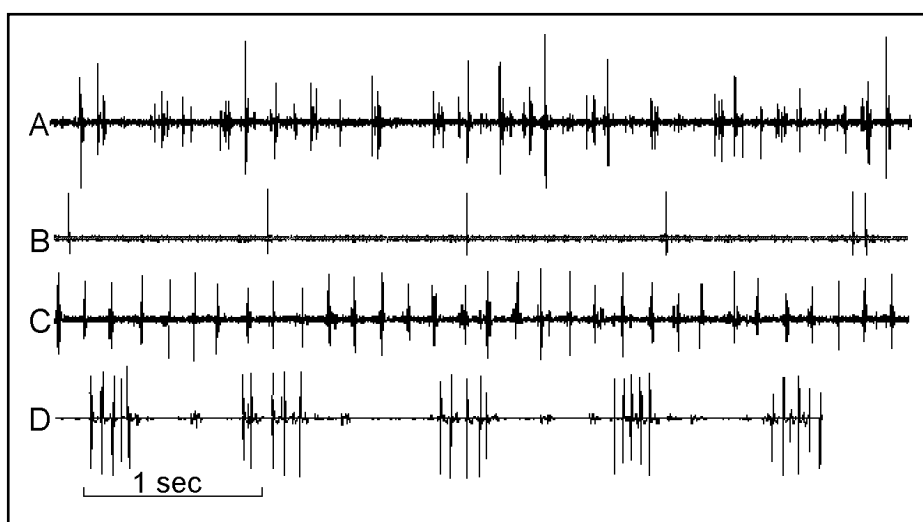


Fig. 1. The examples of spontaneously active pineal cells: A—multicellular activity; B—“slow” type cell in intact rat (regular spikes); C—“fast” type cell in colon tumor-bearing rat (regular spikes); D—“slow” type patterns in colon tumor-bearing rat cell.

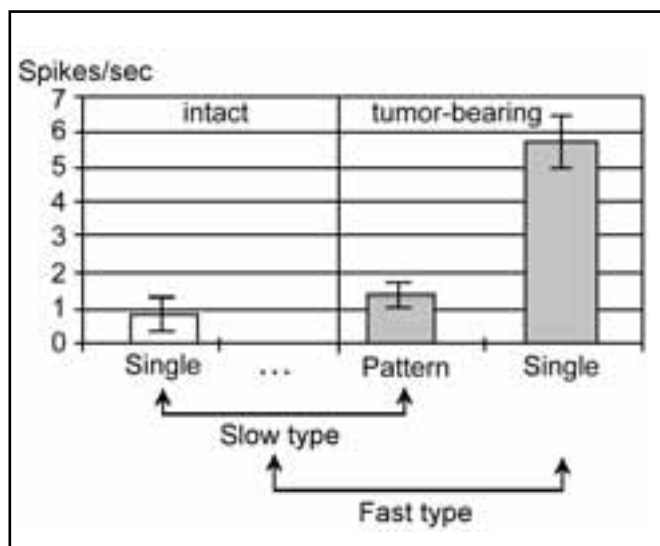


Fig. 2. Spontaneously active pineal cells frequency comparison in intact and colon tumor-bearing rats. "Slow" type regular discharging cells of intact rats are compared with "slow" type pattern activity cells of colon tumor-bearing rats. It allows us to show that summary frequency increased 4–6 times because of switching of cells from "regular" to "pattern" type of activity (from 1 spike to 4–6 spikes in a pattern).

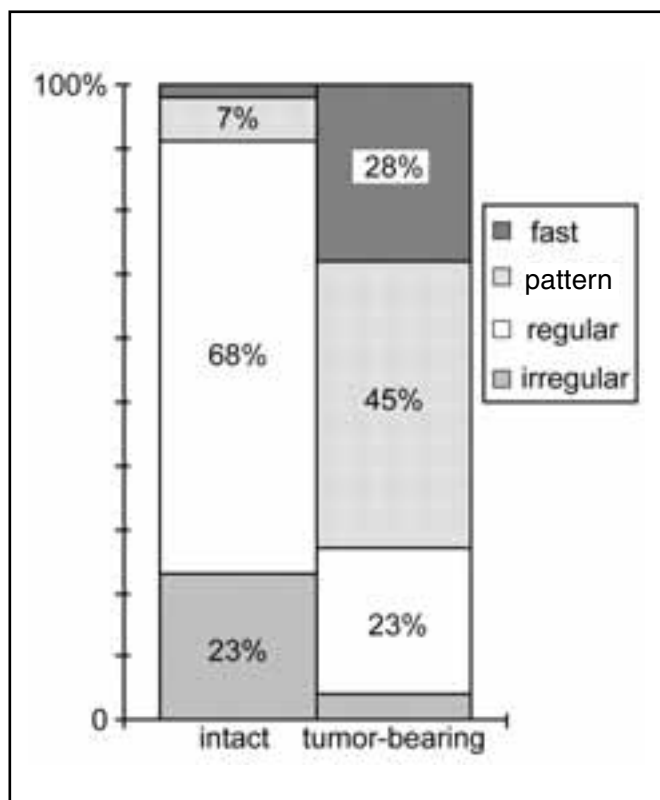


Fig. 3. The ratio of different cell types in intact and colon tumor-bearing rats

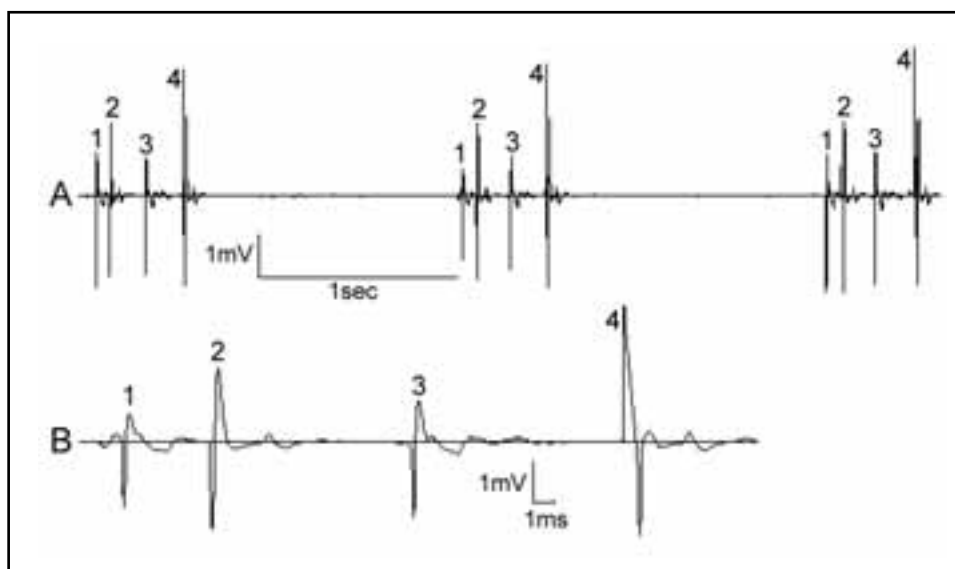
intermediate and granular in foxes. Besides, the cells were decided to belong to a certain type according to their current functional state, because cell optical density depended on time. Pinealocytes with regular, irregular and pattern activity were electrophysiologically identified by different authors [23, 25, 26, 30]. They have established that spike longitude in different cell types vary from 1 ms to about 200 ms. Diversity in types of cellular activity and in spike parameters helped us to distinguish separate cells in multicellular recordings.

The ratio of cells with different types of activity were distinguished in colon tumor-bearing and in intact rats (Fig. 3). The main part of cells in intact rats (91%) was presented by "regularly" and "irregularly" active cells, thus explaining the low summary frequency of spikes. In tumor-bearing rats the number of "pattern" and "fast" cells increased and consequently increased the summary frequency. A three-fold decrease of "regular" cell numbers and a simultaneous 550% increase of "pattern" cell numbers also points to switching of cells to pattern type of activity. This fact and the decrease of "irregular" cell numbers and increase of "fast" cell numbers reflect intensive pineal secretory processes in rats with tumors in the large intestine.

In previous experiments [4] DMH-tumor induced rats demonstrated higher serum melatonin concentration only at night, but not in the daytime, as compared to intact animals. Colon tumor progression may decrease the number of melatonin secreting enterochromaffine cells [15], which decrease reduces the protection of organism against tumor. Pineal hormones (both melatonin and peptides) are known to have antitumor properties. The lack of melatonin leads to compensatory increase of pineal melatonin (nighttime) and peptide (daytime) production. On the basis of known correlation between electric and secretory activity of neurosecretory cells [9, 10, 20], we have supposed the higher pineal electric activity in the daytime (not at night) to reflect intensive non-melatonin secretion. We think that daytime activation of pineal electric activity in DMH-treated rats reflects secretion of peptides, because in our previous study [12, 13] the higher frequency of pinealocyte depolarization was accompanied by an accumulation and exocytosis of peptide/protein containing vesicles in osmotic stress-subjected rats. The ratio of different cellular types, medium frequency of spikes and other peculiarities of electrical activity in tumor-bearing rats looked similar to the same parameters in 48-hour water and food deprived rats [12].

Fig. 4. The group of 4 interacting pineal cells in colon tumor-bearing rat:

A—constant intervals 1–2, 2–3, 3–4 and 4–1 between spikes,
B—the different spikes amplitude, form and polarity. Unequal intervals 1–2, 2–3, 3–4.



The groups of interacting cells were noticed in colon tumor-bearing rats (Fig. 4A). It looked like pattern activity, but cells could be distinguished by spike forms, and cells were discharging in strict consequence with different delays between spikes (Fig. 4B). Such intercellular interactions, probably, realize an activation and synchronic discharging of groups of cells due to existence of tight [14] and gap junctions [29], which probably transfuse signals between the pinealocytes.

An increase of pineal electrical activity during tumor process in the large intestine may be also due to an activation of a sympathetic system. Such activation takes place, for example, in stress [17, 24, 28]. So, we suppose the higher electrical activity of pinealocytes, which in the daytime reflects non-melatonin excretion, and higher blood melatonin concentration in the night, detected by us, to be adaptive reaction against tumor forming, because both melatonin [1, 2, 3, 6] and pineal peptides [3, 16, 22] are known to have anticarcinogenic properties.

In our experiments tumors in the large intestine were revealed in all DMH-treated rats. The square of tumors was $56 \pm 32 \text{ mm}^2$, but there was no correlation between the square and the frequency of spikes of spontaneously pineal active cells.

Thus, the results of our investigation demonstrated that pinealocytes of tumor-bearing rats are secretorily active not only at night, but also in the daytime, as compared to intact rats. Tumor presence or humoral disorders while 48-hour water and food deprivation have a similar incentive effect on pineal secretory processes even in the daytime.

REFERENCES

- Anisimov VN, Reiter RJ. Funkzii epifiza pri rake a starenii [(Pineal function during cancer and ageing) (In Russian with English abstract)] *Vopr oncol* 1990; **36**:259–268.
- Anisimov VN, Khavinson VKh, Morozov VG. Cancirogenesis and ageing IV. Effect of low-molecular-weight factors of thymus, pineal gland and anterior hypothalamus on immunity, tumor incidence and life span of C3H/5n mice. *Mech Ageing Dev* 1982; **19**:245–258.
- Anisimov VN, Khavinson VKh, Morozov VG. Twenty years of study of pineal preparation: epithalamin in experimental gerontology and oncology, *Ann NY Acad Sci* 1994; **719**:483–493.
- Anisimov VN, Popovich IG, Zabezhinski MA. Melatonin and colon carcinogenesis I. Inhibitory effect of melatonin on development of intestinal tumors induced by 1,2-dimethylhydrazine in rats. *Carcinogenesis* 1997; **18**:1549–1553.
- Arend TJ. Melatonin and the mammalian pineal gland. London: Chapman & Hall; 1995.
- Blask DE. Melatonin in oncology. In: Yu, HS, Reiter RJ, editors. Melatonin biosynthesis, physiological effects, and clinical applications. Boca Raton: CRC Press; 1993. p.447–475.
- Blask DE, Wilson ST, Lemus-Wilson AM. The oncostatic and oncomodulatory role of the pineal gland and melatonin. *Adv Pineal Res* 1994; **7**:235–241.
- Bures J, Krekule I, Brozek G. Primenenie EVM v neirofiziologicheskikh issledovaniyah [(Computers in electrophysiological investigations) (In Russian)] Leningrad: Nauka; 1984.
- Douglas WW. Mechanism of release of neurohypophysial hormones: stimulus-secretion coupling. In: Handbook of physiology Sect.7. Washington: Amer Physiol Soc; 1974; **1**:191–224.
- Glebov RN. Endocitoz i Exocitoz [(Endocytosis and exocytosis) (In Russian)] Moscow: "Vyschaya Shkola"; 1987.
- Kolesnikova LA. Epifiz sravnitel'no dikih i domesticirovannyh lisic: Morfofunkcional'nye izmeneniya v techenie sutok [(The pineal gland of comparatively wild and domesticated foxes: morpho-functional circadian changes) (In Russian with English abstract)] *Sechenov Physiol J* 1995; **82**:91–97.

- 12 Kovalenko RI, Sibarov DA, Nozdrachev AD. The influence of unilateral olfactory epithelium stimulation on rat pineal cells during the stress. In: *Advances in Comparative Endocrinology, Proceedings of the 13th International Congress of Comparative Endocrinology*. Japan. 1997. pp 697–700.
- 13 Kovalenko RI, Sibarov DA, Pavlenko IN, Lukianova EL, Nozdrachev AD. Struktura epifiza krys posle inilateral'nyh intranazal'nyh vvedenii oxitocina [(The pineal gland structure in stress while oxytocin unilateral intranasal infusions) (In Russian with English abstract)] *Rus Physiol J* 1997; **83**:87–93.
- 14 Kunwar P. The ultrastructure of mammalian pinealocytes: a systematic investigation. *Micr Res and Tech* 1992; **21**:85–115.
- 15 Kvetnoy I, Sandvik AK, Waldum HL. The diffuse neuroendocrine system and extrapineal melatonin. *J Molecular Endocr* 1997; **18**:1–3.
- 16 Labunetz IF, Butenko GM. The pineal biological substances activity on functional state of thymus end immune system in ageing animals. *Probl Aging and Longevity* 1992; **3**:280–285.
- 17 Lewczuk B, Przybylska-Gornowicz B. Effects of sympathicolytic and sympathicomimetic drugs on pineal ultrastructure in the domestic pig. *J Pineal Res* 1997; **23**:198–208.
- 18 Maestroni GJM, Conti A, Pierpaoli W. Melatonin, stress and the immune system. *Pineal Res Rev* 1989; **7**:203–226.
- 19 McCance I, Parkington HC, Coleman HA. The association between melatonin production and electrophysiology of the guinea pig pineal gland. *J Pineal Res* 1996; **21**:79–90.
- 20 Nordmann JJ. Stimulus—secretion coupling. *Progr Brain Res* 1983; **60**:283–303.
- 21 Petrelli C, Moretti P, Petrelli F, Barra D. Purification of a low molecular weight calf pineal peptide controlling DNA transcription in vitro. *Ital J Biochem* 1992; **41**:170–182.
- 22 Petrelli C, Moretti P, Petrelli F, Bramucci M. A low molecular weight calf pineal peptide with an inhibiting effect on the growth of L1210 and HL60 cells. *Cell Biol Int Rep* 1992; **16**:967–974.
- 23 Parkington HC, McCance I, Coleman HA. Two types of cells with central innervation in pineal gland of guinea pigs. *Am J Physiol* 1987; **252**:369–377.
- 24 Reuss S, Semm P, Vollrath L. Changes in the electrical activity of the rat pineal gland following stimulation of the cervical sympathetic ganglia. *J Auton Nerv Syst* 1985; **12**:281–288.
- 25 Reuss S, Olcese J, Vollrath L. Electrophysiological and endocrinological aspects of aging in the rat pineal gland. *Neuroendocrinology*, 1986; **43**:466–70.
- 26 Reyes-Vazquez C, Prieto-Gomez B, Aldes LD, Dafny N. Rat pineal exhibits two electrophysiological patterns of response to microiontophoretic norepinephrine application. *J Pineal Res* 1986; **3**:213–222.
- 27 Skrinar GS, Bullen BA, Reppert SM, Peachey SE, Turnbull BA, McArthur JW. Melatonin response to exercise training in women. *J Pineal Res* 1989; **7**:185–194.
- 28 Seggie J, Campbell L, Brown GM, Grota LJ. Melatonin and N-acetylserotonin stress responses: effects of type of stimulation and housing conditions. *J Pineal Res* 1985; **2**:39–49.
- 29 Skopichev VG, Kovalenko RI, Seliverstov YuA. Izmenenie ultrastruktury pinealocytov krys v otdalennye sroki posle gipofizektomii [(Changes of rat pineal ultrastructure after long terms of hypophysis removal) (In Russian with English abstract)] *Tsytologia* 1987; **29**:516–520.
- 30 Stehle J, Reuss S, Vollrath L. Electrophysiological characterisation of the pineal gland of golden hamsters. *Exp Brain Res* 1987; **67**:27–32.