

Effect of estradiol 17- β on LH subunits and prolactin mRNAs expression in the pituitary of old female rats

Ewa Wolinska-Witort,^{1,2} Marek Snochowski,² Alina Gajewska,² Yannick Lerrant,³ Raymond Counis³ & Boguslawa Baranowska¹

1. Department of Neuroendocrinology, Medical Centre of Postgraduate Education, Fieldorfa 40, 04-158 Warsaw, Poland.
2. The Kielanowski Institute of Animal Physiology and Nutrition, 05-110 Jablonna, Poland.
3. Endocrinologie Cellulaire et Moleculaire de la Reproduction, Universite Pierre et Marie Curie, ESA CNRS 7080, Paris, France.

Correspondence to: Dr Ewa Wolinska-Witort, Neuroendocrinology Department, Medical Centre of Postgraduate Education, Fieldorfa 40, 04-158 Warsaw, Poland.
FAX: +48 22 610-31-59
E-MAIL: zncmkp@polbox.com

Submitted: October 11, 2000
Accepted: October 23, 2000

Key words: **LH subunits mRNA expression; LH release; PRL mRNA expression; PRL release; pituitary; old rats.**

Neuroendocrinology Letters 2000; 21:431-436 pii: NEL210600A01 Copyright © Neuroendocrinology Letters 2000

Abstract

OBJECTIVES: The aim of the study was to examine susceptibility of the pituitary gland to estrogenic impulse in old, noncycling rats by measurement of steady state level of mRNAs encoding LH subunits α and β and mRNA for PRL.

METHODS: 22-month-old rats were ovariectomized and after one week they were subcutaneously implanted with silastic tubing filled with oil or with estradiol 17- β . Pituitary α , LH β and PRL mRNAs content and serum LH and PRL concentration was determined.

RESULTS: The effect of E₂ treatment was manifested by the significant increase in the weight of the uterus and pituitary gland as well as by elevation of total pituitary RNA (109%, 60% and 78%, respectively; p<0.001). No significant changes (p>0.05) in serum LH concentration were observed, while levels of mRNAs encoding α and LH- β subunits were lowered by 54% (p<0.05) and 96% (p<0.01), respectively, in the rats subjected to E₂ stimuli. No direct correlation between synthesis and release of LH in E₂ treated old rats was observed. The blood PRL concentration and the pituitary level of PRL mRNA increased up to 2,000% and 1,300%, respectively (p<0.001). Spontaneous pituitary adenoma was observed in about 30% of the rats, irrespective of treatment.

CONCLUSIONS: These data show that in old rats estrogenic stimulus can effectively diminish both pituitary LH subunits mRNAs as well as stimulate pituitary PRL mRNA level indicating that the E₂-dependent processes involved in the regulation of corresponding genes are still functional.

Introduction

The gonadal steroid, estradiol 17- β (E_2) plays an important role in the regulation of the biosynthesis and release of gonadotropines as well as prolactin. It has previously been shown that treatment *in vivo* of ovariectomized young mature rats with E_2 resulted in a decreased transcriptional activity of genes encoding the α and LH β subunits [1] and stimulated PRL gene transcription [2]. The regulatory effect of E_2 is mediated into the target cell via estrogen receptor, a ligand-activated transcriptional factor. The estrogen-receptor complex binds to estrogen response elements in target genes and modifies their transcription. The amount of available estrogen receptor seems to decline with age in both pituitary and hypothalamic structures. Our previous study [3] has shown that the amount of the estrogen receptor was diminished by 40–50% in the anterior hypothalamus and the pituitary in 18-month-old rats compared to young female rats (3–6 months old). It is attractive to speculate that the reduced estrogen receptor concentration in the structures of the hypothalamo-pituitary axis in the old female is associated with a change in the estrogen responsiveness in these structures.

The aim of our study was to assess the susceptibility of the pituitary to estrogenic impulse in old, noncycling rats. The effect of estradiol-17 β on the steady-state level of mRNAs coding LH subunits and mRNA for PRL in the pituitary gland as well as LH and PRL concentrations in the peripheral blood of 22-month-old ovariectomized rats were examined.

Materials and methods

Animals

Female Wistar rats (350–360 g) were kept under controlled light (LD 10:14) and temperature (22°C) with free access to pelleted food and tap water *ad libitum*. At the age of 22 months all animals were ovariectomized, and one week later they were randomly assigned into two groups. Fifteen rats were subcutaneously implanted with silastic tubing (inner diameter 0.158 cm outer diameter 0.318 cm; Dow-Corning Corporation, Midland MI, USA) filled with sunflower oil. The other fourteen rats received subcutaneous implants containing 375 μ g of estradiol 17- β (E_2 ; Sigma) in oil. Eight days after implantation, all the animals were decapitated and tissues (pituitary and uterus) were excised and weighed. The anterior pituitary was deep frozen in liquid nitrogen, and stored at -80°C until RNA preparation. Trunk blood was collected and plasma samples were stored at -20°C . In 5 out of 15 ovariectomized rats and 4 out of 14 ani-

mals treated with E_2 , pituitary adenomas were found. Pituitaries that had visible enlarged adenomas were not used for RNA preparation. All experimental procedures were approved by the Ethics Committee at the Kielanowski Institute of Animal Physiology and Nutrition.

Radioimmunoassay

Rat serum LH and PRL were measured by using RIA kits provided by Dr. A.F. Parlow and NIDDK. Values were expressed in terms of rat LH (RP-3) and PRL (RP-3). The intra-assay coefficient of variation was below 5% for both hormones.

RNA preparation and hybridization

The total RNA was prepared from anterior pituitaries according to a scale-adapted CsCl-guanidinium method [4]. Northern blot analysis of LH subunits was performed as previously described [5] with β -actin mRNA, probed with 1150 bp mouse cDNA used as internal reference to correct for the amount of mRNA loading. The rat prolactin cDNA probe was a gift from Prof. J. Martial and Dr. J.N. Laverriere [6]. All probes were labelled with an Amersham Megaprime labeling kit (Amersham, Arlington Heights, IL) The specific band densities were revealed by autoradiography (X-OMAT-AR films, Kodak) at -80°C and quantified by densitometry using a Hoefer GS 300 densitometer.

Statistical analysis

The results are expressed as mean \pm SEM. Statistical evaluation was made using a nonparametric Kruskal-Wallis test [7].

Results

Hormone serum concentrations and LH subunits and PRL mRNAs pituitary levels

LH concentration was not significantly altered after E_2 treatment; however, the levels of pituitary mRNAs encoding α - and LH- β subunits were significantly depressed by 54% and 96%, respectively (Fig. 1). PRL serum concentration increased by 2,000% in the animals subjected to E_2 treatment whereas pituitary mRNA content for PRL increased by 1,300% (Fig. 2).

Effect of E_2 on pituitary gland and uterus weight and total pituitary RNA content

Selected tissue parameters in the animals with macroscopically normal pituitary gland are shown in Table 1. Exogenous E_2 increased uterus and pituitary gland weight as well as pituitary RNA concentration by 109%, 60% and 78% respectively, as compared to ovariectomized animals.

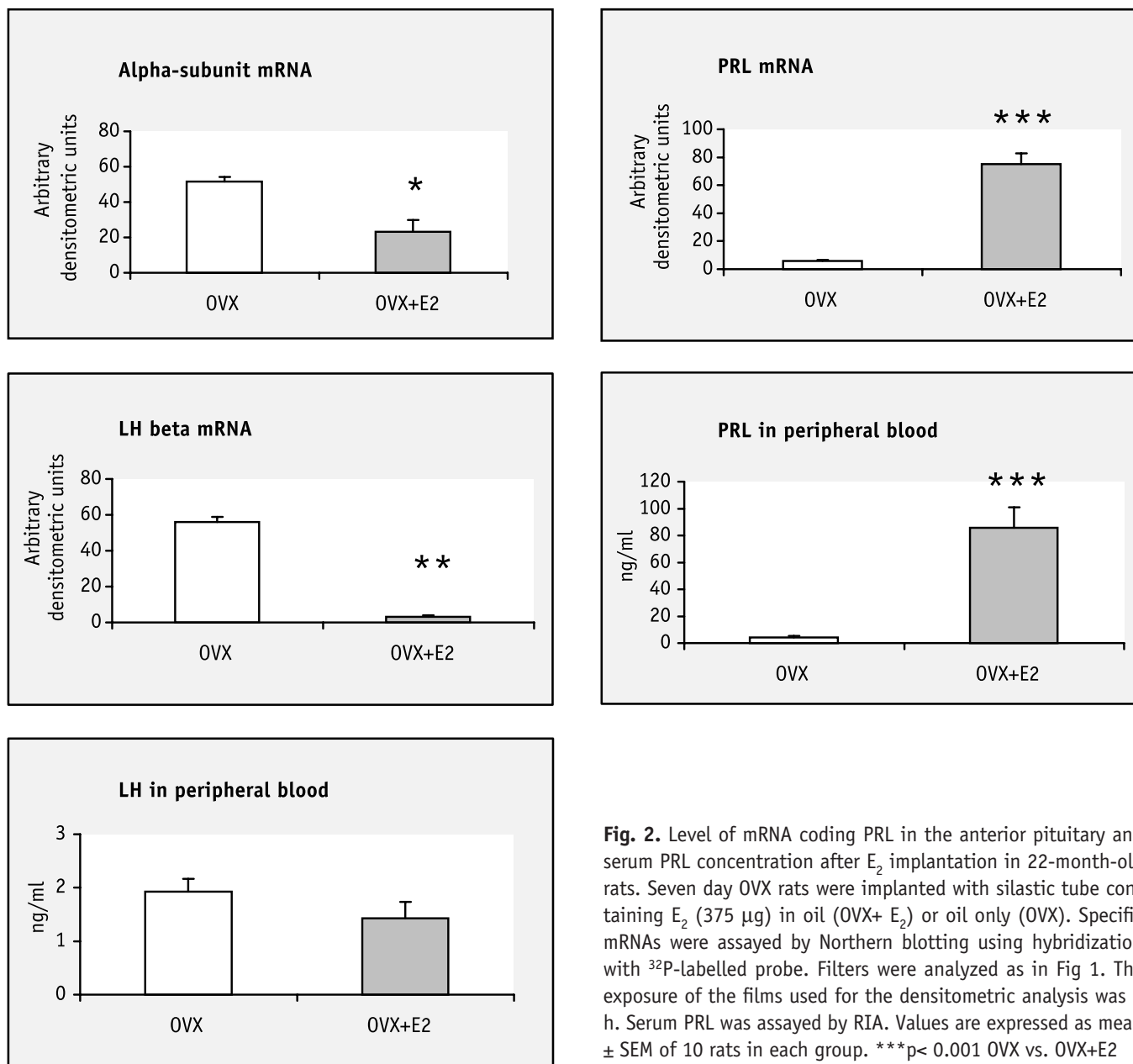


Fig. 2. Level of mRNA coding PRL in the anterior pituitary and serum PRL concentration after E₂ implantation in 22-month-old rats. Seven day OVX rats were implanted with silastic tube containing E₂ (375 μ g) in oil (OVX+ E₂) or oil only (OVX). Specific mRNAs were assayed by Northern blotting using hybridization with ³²P-labelled probe. Filters were analyzed as in Fig 1. The exposure of the films used for the densitometric analysis was 1 h. Serum PRL was assayed by RIA. Values are expressed as mean \pm SEM of 10 rats in each group. ***p< 0.001 OVX vs. OVX+E2

Fig. 1. Levels of α -, LH β subunit mRNAs in the anterior pituitary and serum LH concentration after E₂ implantation in 22-month-old rats. Ovariectomized rats were implanted with 375 μ g of E₂ (OVX+ E₂) or oil only (OVX). Specific mRNAs were assayed by Northern blotting using hybridization with ³²P-labelled probes. Absorbance of each band detected from the autoradiogram of the RNA blots was determined by scanning densitometry and expressed as arbitrary densitometry units (ADU). Exposure of films used for densitometric analysis was 24 h for α subunit and 48h for LH β . Serum LH concentration was assayed by RIA. Values are expressed as mean \pm SEM of 10 rats in each group. *p<0.05; **p< 0.01 OVX vs. OVX+E2

Table 1. The effect of estradiol treatment on the selected parameters of reproductive tissues in old rats with normal pituitary gland.

TISSUE PARAMETERS	TYPE OF TREATMENT	
	OVX (n=10)	OVX + E ₂ (n=10)
Uterus weight [mg]	510 ± 48	1067 ± 42 ***
Pituitary weight [mg]	12.1 ± 0.4	19.4 ± 0.7***
Total RNA [µg/mg of pituitary]	1.81 ± 0.06	3.22 ± 0.15***

Values are expressed as mean ± SEM; *** p < 0.001 OVX vs. OVX+E₂

Table 2. Tissue weight and the pituitary hormone plasma concentration in old rats with pituitary adenomas.

TISSUE WEIGHT [mg]	TYPE OF TREATMENT	
	OVX (n=5)	OVX + E ₂ (n=4)
Uterus	534 ± 70	817 ± 50**
Pituitary gland	18.7 ± 3.43	39.3 ± 6.55**

CONCENTRATION OF HORMONES IN BLOOD [ng/ml]		
LH	1.15 ± 0.17	< 0.96
PRL	154 ± 71	351 ± 97*

Values are expressed as mean ± SEM; * p < 0.05; **p < 0.01 OVX vs. OVX+E₂

Tissue and hormonal status of old female rats with pituitary adenomas

Frequency of pituitary adenomas appearance was similar (33% in ovariectomized group and 29% in E₂ treated rats). E₂ supplementation increased uterus and pituitary weight by 50% and 110% respectively, as compared to control. In rats with pituitary adenomas mean serum PRL concentration was higher while LH concentration was lower in comparison to their level in rats with normal pituitary glands. The mean concentration of PRL was higher by 128% in rats treated with E₂ (Table 2).

Discussion

In our previous work [1] it was shown that estradiol administered to young ovariectomized rats negatively regulated expression of mRNA gonadotropin subunits *in vivo*. The present study shows that E₂ supplementation to old animals also caused significant reduction of pituitary levels of α -subunit (54%) and LH β (96%) subunits mRNA. It is well established that in the ovariectomized animal model the major phenomenon reflected by increased gonadotropin gene expression and gonadotropin release is the suppression of the inhibitory effect of steroids

on GnRH secretion (synthesis and release), which results in hyperstimulation by the neurohormone of pituitary gonadotrophs and, consequently, hyperexpression of gonadotropin genes [8, 9]. The ability of estradiol and other steroids to modulate gonadotropin gene expression has been investigated predominantly on young animal models. In this model steroids were demonstrated to operate either or both directly at the pituitary and indirectly via negative actions at the hypothalamus to depress GnRH secretion [10, 11]. The greater changes after E₂ treatment observed in LH β mRNA content as compared to those of α -subunit mRNA suggest that estrogen directly or indirectly is a major regulator of LH β subunit synthesis. Several down and upstream DNA binding transcription factors regulate LH β gene expression [12] forming tripartite GnRH response element in the rat LH β gene promoter. Data based on the expression of the equine LH β promoter in α T3-1 cells show that GnRH induces transcriptional factors with opposite actions: firstly, Egr-1, acting as a stimulator in conjunction with SF-1 and secondly, Nab1, acting as an inhibitor of Egr-1 [13]. Whether steroids could regulate the expression of gonadotropin genes directly, at the pituitary level, has been intensively investigated [11]. Specific 15-base palindromic LH β gene region was identified as estradiol responsive element (ERE)

necessary and sufficient to confer *in vitro* E₂ mediated transcriptional response [14]. No such ERE has been identified on α -subunit gene; however, despite the absence of high affinity binding site for the estrogen receptor it has been shown that E₂ inhibits transcription of the human glycoprotein hormone α subunit gene [15]. Moreover, tissue specific expression of the glycoprotein hormone α subunit gene in pituitary gonadotrope has been shown to rely on gonadotrope-specific element (GSE) which binds the orphan nuclear receptor steroidogenic factor-1 [16]. So far there are no detailed studies on the old animal model which explore the possible effect of aging on interactions of differential transcriptional factors with specific genes coding gonadotropin subunits mRNAs. Reduced ability of GnRH to stimulate LH release from pituitary glands of old female and male rats both *in vivo* [17] and *in vitro* [18] have been demonstrated which may reflect decreasing with age sensitivity of GnRH receptors for its stimulation and in consequence markedly affect LH biosynthesis. Strong inhibitory effect of estradiol on LH subunits mRNA pituitary level as well as its powerful hypertrophic effect exerted on pituitary and uterus tissue weight observed in old ovariectomized rats suggest that gene and tissue estradiol sensitivity is not abolished in old animals. Nevertheless, in old female mice age-related changes in estradiol receptor dynamics in hypothalamus, pituitary and uterus have been reported which result in suboptimal reaction of target tissues to estradiol [19, 20]. Inhibitory effect of estradiol on LH subunits mRNA expression was not accompanied by its parallel influence on mean plasma levels of circulating LH. Such result may reflect a decline with aging of the negative feedback loop exerted by E₂ [21] as well as diminished ability of old pituitary to release gonadotropin. Moreover, it also suggests that the effect observed at the transcription level (less mRNA for LH α subunit) may not adequately "translate" to parallel changes observed at the level of mature, biologically active hormone (less concentration of circulating LH). The steady-state level of cytoplasmic mRNA is determined by the balance between synthesis, processing, transport and degradation processes. Also stability of some mRNA species can be changed in response to hormonal induction and this process can play an important role in the control of gene expression and translation efficiency.

Changes in PRL gene expression and synthesis were opposite to those of LH subunits in response to E₂ treatment. It has been shown that E₂ induced increased expression of the PRL gene. The estrogen-receptor complex binds to an estrogen response element in the PRL gene and modifies its transcrip-

tion. An ERE sequence has been identified in the distal promoter region of the PRL gene [22]. The estrogen responsiveness of the rat PRL gene expression requires not only the presence of the estrogen receptor but also the tissue-specific transcription factor, Pit-1 protein [23]. Increased level of pituitary PRL mRNA is accompanied by high concentration of serum PRL. In young animals PRL may rise 10- to 20-fold after *in vivo* E₂ administration [24]. In our study a similar effect of E₂ on the transcription process of the PRL gene and the secretion PRL in old rats was observed. In many cases the main source of elevated level of PRL in aging rats are adenomas developing in their pituitaries [25, 26]. It is well established that PRL exerts suppressive effect on LH release. Increasing activity of hypothalamic dopaminergic neurons affected frequency and amplitude of LH pulses [27] as well as reduced mean level of circulating gonadotropines and pituitary GnRH receptor numbers [28]. Also stimulation of hypothalamic endorphinergic neurons observed in hyperprolactinemic rats resulted in inhibition of GnRH release [29]. A direct effect of PRL on the pituitary gland cannot be excluded [30] as PRL inhibited LH release from gonadotrope cells *in vitro* and inhibited their sensitivity to exogenous GnRH stimulation.

In summary, our results show that in old rats estrogenic stimulus can effectively diminish both pituitary LH subunit mRNAs as well as stimulate pituitary PRL mRNA level indicating that the E₂-dependent processes involved in the regulation of corresponding genes are still functional.

Acknowledgments

The authors wish to express their warmest thanks to Dr. A.F. Parlow and NIDDK, NHPP Programme for providing us with the RIA kit for LH and PRL (supplied to Dr. K.Kochman, PAN, Jablonna).

REFERENCES

- 1 Corbani M, Counis R, Wolinska-Witort E, d'Angelo-Bernard G, Moumni M, Jutisz M. Synergistic effects of progesterone and oestradiol on rat LH subunit mRNA. *J Mol Endocrinol* 1990; **4**:119–125.
- 2 Shull JD, Gorski J. Estrogen regulation of PRL gene transcription in vivo: Paradoxical effects of 17 β -estradiol dose. *Endocrinology* 1989; **124**:279–285.
- 3 Snochowski M, Wolinska-Witort E, Radzikowska M, Kochman K. Concentration of estrogen receptor in selected structures of central nervous system and pituitary in female rats. XII Congress of the Polish Endocrinology Society, Szczecin, 2–5 June 1987, Abstracts p. 143.
- 4 Counis R, Corbani M, Berault A, Theoleyre M, Janssen de Almeida Catanho MT, Jutisz M. Une micromethode permettant de preparer et de traduire l'acide ribonucleique messenger a partir de cellules adenohipophysaires en culture. *CR Acad Sci Paris* 1981; **293**:115–118.
- 5 Lerrant Y, Kottler ML, Bergametti F, Moumni M, Blumberg-Tick J, Counis R. Expression of GnRH receptor gene is altered by GnRH agonist desensitization in a manner similar to that of gonadotropin β -subunit genes in normal and castrated rat pituitary. *Endocrinology* 1995; **136**:2803–2808.
- 6 Cooke NE, Weiner RI, Baxter JD, Martial JA. Structure of cloned DNA complementary to rat prolactin mRNA. *J Biol Chem* 1980; **255**:6502–6506.
- 7 Kruskal WH, Wallis WA. Use of ranks in one-criterion variance analysis. *J Amer Statist As* 1952, **47**:583–621.
- 8 Jutisz M, Starzec A, Corbani M, Counis R. Synthesis of gonadotropins and its regulation in the pituitary. *Hum Reprod* 1988; **3**:485–489.
- 9 Gharib SD, Wierman ME, Shupnik MA, Chin WW. Molecular biology of the pituitary gonadotropins. *Endocr Rev* 1990; **11**:177–199.
- 10 Mercer JE, Clements JA, Funder JW, Clark IJ. Rapid and specific lowering of pituitary FSH mRNA levels by inhibin. *Mol Cell Endocrinol* 1987; **53**:251–254
- 11 Counis R. Gonadotropin biosynthesis, in: J Neill, E Knobil, editors. *Encyclopedia of Reproduction*, vol 2, Acad Press, 1999; 507–520.
- 12 Kaiser UB, Halvorson LM, Chen MT. Sp1, steroidogenic factor 1 (SF-1) and early growth response protein 1 (egr-1) binding sites form a tripartite gonadotropin-releasing hormone response element in the rat luteinizing hormone β gene promoter: an integral role for SF-1. *Mol Endocrinol* 2000; **14**:1235–1245.
- 13 Call GB, Wolfe MW. Gonadotropin-releasing hormone activates the equine luteinizing hormone β -promoter through a protein kinase C/mitogen-activated protein kinase pathway. *Biol Reprod* 1999; **61**:715–723.
- 14 Shupnik MA, Weinman CM, Notkles AC, Chin WW. An upstream region of the rat LH β gene binds estrogen receptor and confers estrogen responsiveness. *J Biol Chem* 1989; **264**:80–86.
- 15 Keri RA, Andersen B, Kennedy GC, Hammernik DL, Clay CM, Brace HD, et al. Estradiol inhibits transcription of the human glycoprotein hormone α -subunit gene despite the absence of a high affinity binding site for the estrogen receptor. *Mol Endocrinol* 1991; **5**:725–733.
- 16 Barnhart KM, Mellon PL. The orphan nuclear receptor, steroidogenic factor-1, regulates the glycoprotein hormone α -subunit gene in pituitary gonadotropes. *Mol Endocrinol* 1994; **8**:878–885.
- 17 Huang H, Marshall S, Meites J. Capacity of old versus young female rats to secrete LH, FSH and PRL. *Biol Reprod* 1985; **14**:538–543.
- 18 Tang HJK, Tang FY. LH response to LHRH, dBcAMP and 17 β -estradiol levels in cultures derived from aged rats. *Am J Physiol* 1981; **240**:E510–E516.
- 19 Nelson JF, Bergman MD, Karelus K, Felicio LS. Aging of the hypothalamo-pituitary-ovarian axis: hormonal influences and cellular mechanisms. *J Steroid Biochem* 1987; **24**:699–705.
- 20 Bergman MD, Karelus K, Felicio LS, Nelson JF. Differential effects of aging on estrogen receptor dynamics in hypothalamus, pituitary and uterus of the C57BL/6J mice. *J Steroid Biochem* 1989; **33**:1027–1033.
- 21 Finch CE, Felicio LS, Mobbs CV, Nelson JF. Ovarian and steroidal influences on neuroendocrine aging in female rodents. *Endocr Rev* 1984; **5**:467–497.
- 22 Maurer RA, Notides AC. Identification of an estrogen-responsive element from the 5'-flanking region of the rat prolactin gene. *Mol Cell Biol* 1987; **7**:4247–4254.
- 23 Ying C, Lin DH, Sarkar DK, Chen TT. Interaction between estrogen receptor and Pit-1 protein is influenced by estrogen in pituitary cells. *J Steroid Biochem Mol Biol* 1999; **68**:145–152.
- 24 Murai I, Ben-Jonathan N. Acute stimulation of prolactin release by estradiol: mediation by the posterior pituitary. *Endocrinology* 1990; **126**:3179–3184.
- 25 McComb DJ, Hellman P, Kovacs K, Scott D, Evans WS, Burdison JA, et al. Spontaneous sparsely granulated prolactin producing pituitary adenomas in aging rats. *Neuroendocrinology* 1985; **41**:201–211.
- 26 Chuknyiska RS, Blackman MR, Hymer WC, Roth GC. Age-related alterations in the number and function of pituitary lactotropic cells from intact and ovariectomized rats. *Endocrinology* 1986; **118**:1856–1862.
- 27 Cohen IR, Becker AM, Seimano M, Wise PM. Hyperprolactinemia alters the frequency and amplitude of pulsatile LH secretion in the ovariectomized rat. *Neuroendocrinology* 1986; **42**:328–333.
- 28 Garcia A, Herbon L, Barkan A, Papavasiliou S, Marshall JC. Hyperprolactinemia inhibits GnRH stimulation on the number of pituitary GnRH receptors. *Endocrinology* 1985; **117**:954–959.
- 29 Carter DA, Cooper JS, Inkster SE, Whitehead SA. Evidence for an increased opioid inhibition of LH secretion in hyperprolactinemic ovariectomized rats. *J Endocrinol* 1984; **101**:57–62.
- 30 Cheung CY. Prolactin stimulates LH secretion and pituitary responsiveness to LHRH by a direct action of the anterior pituitary. *Endocrinology* 1983; **113**:632–638.