Increased Urinary 6-sulfatoxymelatonin Excretion in Women with Non-classical Steroid 21-hydroxylase Deficiency

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Abstract

OBJECTIVES: To determine melatonin production in hyperandrogenic women.

MATERIAL AND METHODS: Seventeen women with late onset adrenal hyperplasia due to 21-hydroxylase deficiency (LOCAH) and 15 control women were studied in early follicular phase of the menstrual cycle. Fasting serum levels of luteinizing hormone (LH), follicle-stimulating hormone (FSH), estradiol (E_2), testosterone, dihydroepiandrosterone sulfate (DHEA-S), 17-hydroxyprogesterone (17-OHP) as well as the peak 17-OHP response to ACTH (250 µg IV) and 24h urinary 6-sulfatoxymelatonin (aMT6s) were determined in all participants.

RESULTS: All 17 hyperandrogenic women were carrying mutations of the CYP21 gene. Women with LOCAH had significantly higher serum testosterone, DHEA-S, 17-OHP and ACTH stimulated 17-OHP values compared with controls. Their aMT6s values ($44.6\pm20.3 \mu g/24h$) were significantly higher than the values in control women (31.5 ± 20.3) (p<0.03). The urinary aMT6s values were positively correlated with testosterone (p<0.04), DHEA-S (p<0.02) and peak 17-OHP (p<0.04).

CONCLUSIONS: Women with LOCAH have increased melatonin production. There is a relationship between adrenal androgens and melatonin in these women.

Introduction

The significance of melatonin in human reproduction is not established at present. However, a large body of information suggests that melatonin and the reproductive hormones are inter-related. This concept is based on observations of elevated melatonin levels in hypogonadal patients due to GnRH deficiency [1-4], decreased melatonin concentrations in primary hypogonadism [5] and in precocious puberty [6]. Testosterone or E_2 replacement therapy given to hypogonadal patients, normalized their melatonin levels [2, 7]. The measurement of urinary 6-sulfatoxymelatonin (aMT6s), the major metabolite of melatonin, was shown to correlate with plasma hormone levels and is considered to be a good indicator of pineal melatonin secretion [8–10].

Non-classical adrenal hyperplasia due to 21-hydroxylase deficiency (LOCAH) is the most frequent autosomal-recessive genetic disorder in man. LOCAH is a mild disorder of cortisol biosynthesis characterized clinically in syptomatic females by manifestations of androgen excess as hirsutism, acne, menstrual disorders or unexplained sterility [11, 12]. In most patients, serum 170HP levels are elevated after stimulation with ACTH [13]. On occasion, blood DHEA-S levels are increased after ACTH [11]. Levels of 3α -androstanedione glucoronide, an androgen metabolite, are elevated in LOCAH patients and highly correlated with levels of androstanedione and testosterone [14].

So far, melatonin secretory profiles in women who are hypoandrogenic as a result of adrenal hyperplasia have not been studied. Based on the literature cited, we hypothesized that hyperandrogenic state would cause lower melatonin production. To examine this hypothesis, we determined 24h urinary aMT6s excretion in hyperandrogenic women with LOCAH and in normal cycling nonhirsute women.

Material and methods

Subjects and Protocol

The study was approved by the Institutional Review Board (Helsinki Committee) and all participants gave their informed consent before the start of the study. The control group for this study consisted of 15 normal women aged 23.7 ± 5.2 years with regular menses and no signs of virilization. None were taking medications including oral contraceptives. The patient population consisted of 17 women aged 21.9 ± 5.0 years, all carrying mutations for the CYP21 gene. Patients were referred for investigation of hirsutim, acne or menstrual disorders. Hirsutism was defined as a score ≥ 8 by the Ferriman-Gallwey index [15]. Oligomenorrhea was defined as menstrual

cycles \geq 35 days [16]. Obesity was assessed by estimating body mass index (BMI) with normal values ranging between 17.0–25.9 kg/m² [17]. Blood samples and urine collection were performed during early follicular phase of the spontaneous menses when present or otherwise in amenorrhea, for the determination of serum LH, FSH, prolactin, E₂, testosterone and DHEA-S. In all participants we determined serum 170HP at base-line and in response to 250 µg IV ACTH at 30 and 60 mins.

The diagnosis of non-classical steroid 21-hydroxylase deficiency was made on the basis of genetic analysis and a serum 17OHP level after ACTH more than 2SD above the normal mean [18].

Melatonin production was assessed as the 24h urinary 6-sulfatoxymelatonin (aMT6s) excretion. All studies were performed between January and May of the same year.

Genetic Analysis of the CYP21 Gene

DNA was isolated from peripheral blood leucocytes. For mutation analysis of CYP21, we employed PCR amplification of CYP21 specific fragments and restriction enzymes analysis [19]. The following mutations in CYP21 were analysed: P30L, I2 splice, I172L, exon 6(E6) cluster, V281L, Q318X, and the homozygous state of the exon 3 del 8 bp (E3 del8) [20].

Hormone Measurements

Blood samples were centrifuged, immediately separated and stored at -20°C until assayed. The concentrations of serum LH, FSH, E₂, testosterone, cortisol, DHEA-S and 17OHP were determined by commercial kits. Serum LH and FSH were determined by the immunometric technique (Biodata Diagnostics, Rome, Italy). Normal levels are: LH - 10-18 mIU/ml and FSH - 4-13 mIU/ml (follicular phase of the menstrual cycle). Serum cortisol, DHEA-S and 17-hydroxyprogesterone were determined by radioimmunoassay methods (Diagnostic Products Corporation, Los Angeles, CA). The normal levels of these hormones are as follows: cortisol $-5-25 \,\mu g/dl$, DHEA-S - 35–430 ng/ml and 170HP - less than 5.0 pg/ml. Serum total testosterone and 17β estradiol were determined by competitive immunoassay using the Immulite analyzer (Diagnostic Products Corporation. Los Angeles, CA). The normal levels are: testosterone – 0.2–0.8 ng/ml and E_2 – 7–178 pg/ml during the follicular phase of the menstrual cycle. Melatonin was measured as urinary 6-sulfatoxymelatonin (aMT6s). Urine was collected over a 24h period, the volume of each collection was recorded and aliquotes were frozen at -20°C until assayed. Creatinine concentration was determined in each sample to verify complete 24h collection with values of 18mg/kg/24h taken for normal females [21].

The concentrations of aMT6s in urine samples were determined by an enzyme-immunoassay method (ELISA) as previously described [22]. The melatonin-sulfate ELISA kits were provided by Immunobiological Laboratories, Hamburg, Germany. The assay sensitivity was 0.3 ng/ml. The intra-assay and interassay coefficients of variation (CV) were 10% and 16%, respectively.

The assay sensitivity was 0.3 ng/ml. The cross-reactivity of the anti-melatonin sulfate antiserum is 0.0002% for melatonin, 0.001% for 6-hydroxymelatonin, 0.0005% for N-acetyl-L-hydroxytryptophan and less than 0.0001% for N-acetyl-L-tryptophan.

Statistical Analysis

We examined urinary aMT6s values expressed as total μ g per 24h, serum testosterone, DHEA-S, 17OHP (basal and peak) as well as LH, FSH and E₂ levels in LOCAH patients and controls. Data are given as mean ±SD. Differences between groups were tested by Wilcoxon 2 sample test. Spearman rank correlation were computed for the entire population and for each group separately.

Results

The clinical and endocrine characteristics of the study subjects are shown in Table 1. All 17 patients with LOCAH had hirsutism, four had acne (23.5%), three had oligomenorrhea (17.6%) and two patients were amenorrheic (11.6%). Genetic analysis revealed

that nine women carried the Q318X mutation and eight women carried the V281L mutation. Serum testosterone, basal and ACTH stimulated 17OHP levels were significantly higher in LOCAH patients than in controls. Their cortisol levels at baseline and in response to ACTH stimulation were not different from values in controls. Serum DHEA-S levels in LOCAH patients (236±90 ng/ml) were higher than the values in controls (196±84 ng/ml) although not statistically significantly different.

Results of urinary aMT6s excretion are shown in Table 2. Patients had significantly higher total aMT6s values ($44.6\pm20.3\,\mu$ g/24h vs. 31.5 ± 7.5 in controls, p<0.03). When aMT6s were expressed per body weight, per BMI or as the rate of excretion, values in patients were statistically not significantly different from the values in controls.

Spearman correlation analysis revealed that aMT6s values were positively correlated with DHEA-S levels (r=0.41; p=0.02) and with peak 17OHP levels (r=0.36; p=0.04) but not with testosterone levels (r=0.04; p=NS). A significant positive correlation between aMT6s (expressed as rate of excretion) and testosterone was observed in controls (r=0.53; p=0.04) or when expressed per body weight (r=0.51; p=0.05). In LOCAH patients, these correlations were of borderline significance (r=-0.42; p=0.09, r=-0.44; p=0.07, respectively). Plot of BMI vs. aMT6s (Fig. 1) revealed a weak correlation between these two variables. Regression analysis indicated no linear relationship between aMT6s and BMI in either group.

Table 1. Clinical and Endocrine Characteristics in Hyperandrogenic Women with Late-Onset Adrenal Hyperplasia due to 21-Hydroxylase Deficiency (Data are the mean±SD)

Cortisol (µg/dl) Basal Peak	(pg/ml) Peak		(mIU/ml)	(mIU/ml)	(m m /ml)						Group
13.3+3.9 30.3+6				((pg/ml)	(ng/ml)	(ng/ml)	Score*	(kg/m²)	(years)	·
1010_019 0019_01	15.7±3.8	3.6±0.7	5.6±1.8	5.3±2.3	57.6±25.9	236±89	0.5±0.2	13.6±3.7	26.4±4.9	21.9±5.0	Patients (n=17)
14.1±5.6 29.5±5.	3.5±0.9	1.3±0.7	6.5±1.4	4.6±2.2	50.9±21.3	196±84	0.3±0.1	<8	24.0±5.2	23.7±5.2	Controls (n=15)
N.S. N.S.	0.0001	0.02	N.S.	N.S.	N.S.	N.S.	0.03	0.0001	0.05	N.S.	P value
		110_01/	010_111		5005_2105	100_01	0.03	-			Controls (n=15)

Table 2. Urinary 6-Sulfatoxymelatonin (aMT6s) Excretion in Hyperandrogenic Women with Late-Onset Adrenal Hyperplasia Due to 21-Hydroxylase Deficiency. (Data are the Mean±SD)

Group	AMT6s-totalAMT6s per BMIAM(μg/24h)(ng/kg/m²)		AMT6s – Rate of excretion (ng/h/kg)	AMT6s – Per Body weight (ng/kg/24h)	
Patients (n=17)	44.6±20.3	1757±905	28.1±14.9	675±357	
Controls (n=15)	31.5±7.5	1375±429	22.4±7.8	537±186	
P value	0.03	N.S.	N.S.	N.S.	

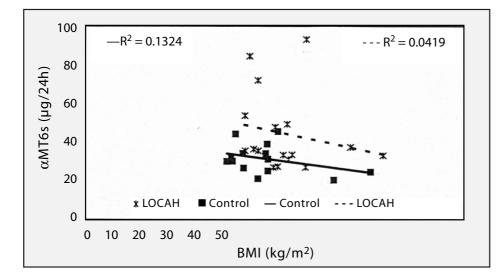


Fig. 1. Urinary aMT6s excretion in hyperandrogenic women with late-onset adrenal hyperplasia (LOCAH) and controls revealed a weak correlation with BMI. Regression analysis indicates no linear relationship between melatonin and BMI in patients and controls.

Discussion

In the current study we have demonstrated that total aMT6s values were significantly higher in hyperandrogenic women with late-onset adrenal hyperplasia compared with controls. Total aMT6s values positively and significantly correlated with DHEA-S and peak 170HP levels. Also, a positive correlation was observed between testosterone and aMT6s expressed as rate of excretion. Our findings first demonstrate that LOCAH patients have increased melatonin secretion which is associated with their androgen levels.

How the reproductive hormones modulate melatonin secretion in humans is still largely unknown. Several candidates are possible: androgens (either directly or through conversion to estradiol), gonadotropins, adrenal steroids, mainly cortisol or a combination of all. In children with congenital adrenal hyperplasia who had low cortisol levels, plasma melatonin levels were normal. Replacement therapy with glucocorticoids did not modify their melatonin levels [23]. In patients with Cushing's syndrome, melatonin secretion was intact [24]. In our study, all patients had intact basal and ACTH stimulated cortisol levels, suggesting that cortisol does not participate in the modulation of melatonin secretion.

A possible relationship between melatonin and GnRH or gonadotropins was suggested by the findings of increased melatonin secretion in hypogonadal patients [3,4], decreased melatonin levels in conditions of gonadotroipins excess as precocious puberty [6] and Klinefelter's syndrome [5], the demonstration of melatonin receptors in the hypothalamic suprachiastmatic nuclei [25] and by the enhancing effect of melatonin on the gonadotropins response to GnRH in normal women [26].

However, the lack of correlation between serum gonadotropin levels and aMT6s in the present study and in a previous report [27] suggest that the possibility of gonadotropins or GnRH modulate melatonin secretion is unlikely.

In favor of estrogens affecting melatonin secretion in our patients are the findings of elevated melatonin levels in estrogen deficient women with endometriosis receiving GnRH analogue therapy and the diminution of melatonin after E_2 replacement in women with GnRH deficiency [2]. Yet the efficacy of E_2 in modulating melatonin secretion was not confirmed by others [28], nor was suppression of E_2 levels during GnRH agonist therapy given to patients with precocious puberty, associated with normalization of melatonin levels [6]. In the current study, E_2 serum levels were normal in LOCAH patients and were not correlated with aMT6s values suggesting that estrogens probably do not participate in the modulation of melatonin secretion.

A more plausible explanation is that androgens modulate melatonin secretion in our patients. This hypothesis is supported by the findings of significant positive correlations between aMT6s and serum androgen levels in the current study, increased aMT6s values in hyperandrogenic women with polycystic ovary syndrome which was also correlated with their testosterone levels [29], the normalization of melatonin levels during testosterone treatment given to hypogonadal men [3,7] and by the demonstration of androgen and estrogen receptors in rat and human pineal glands [30].

On the other hand, in men with leuprolide-induced hypogonadism, testosterone treatment did not change melatonin levels [31]. Similarly, in primary hypogonadism or in adult-onset hypogonadism, testosterone replacement therapy did not alter plasma melatonin levels [22, 32]. Caglayan et al [27] have recently demonstrated that in patients with Klinefelter's syndrome, plasma melatonin levels tended to be higher than levels in normal controls, whereas those of aMT6s were lower. Testosterone replacement was associated with a fall in plasma melatonin levels and urinary aMT6s values increased. The authors suggested that alterations in melatonin metabolism rather than any effect on net sympathetic activity were responsible for their findings. These data suggest that the pathophysiological role of melatonin in human reproduction is far from being clarified. It is clear though that there is no simple classic feed-back regulation between the pineal and the gonads.

In conclusion, women with adrenal hyperandrogenism due to 21-hydroxylase deficiency have increased melatonin production which is associated with their increased androgen secretion.

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REFERENCES

- 1 Brzezinski A, Lynch HJ, Seibel NM, Deng MH, Wurtman RJ. The circadian rhythm of plasma melatonin during the normal menstrual cycle and in amenorrheic women. J Clin Endocrinol Metab 1988; **66**:891–895.
- 2 Okatani Y, Sagara Y. Amplification of nocturnal melatonin secretion in women with functional secondary anemorrhea: relation to endogenous oestrogen concentration. Clin Endocrinol 1994; 41:766–770.
- 3 Luboshitzky R, Lavi S, Thuma I, Lavie P. Testosterone treatment alters melatonin concentrations in male patients with gonadotropin-releasing hormone deficiency. J Clin Endocrinol Metab 1996; 81:770-774.
- 4 Kadva A, Djahanbakach O, Monson J, Di WL, Silman R. Elevated nocturnal melatonin is a consequence of gonadotropin-releasing hormone deficiency in women with hypothalamic amenorrhea. J Clin Endocrinol Metab 1998; **83**:3653–3662.
- 5 Luboshitzky R, Wagner O, Lavi S, Herer P, Lavie P. Decreased nocturnal melatonin secretion in patients with Klinefelter's syndrome. Clin Endocrinol 1996; 45:749–754.
- 6 Waldhauser F, Boepple PA, Schempter M, Mansfield MJ, Crowley WF. Serum melatonin in central precocious puberty is lower than age-matched pre- pubertal children. J Clin Endocrinol Metab 1991; 73:793–796.
- 7 Rajmil O, Puig-Domingo M, Tortosa F, Viader M, Petterson AG, Schwarzstein, D. et al. Melatonin concentration before and during testosterone replacement in primary hypogonadic men. Europ J Endocrinol 1997; 137:48–52.
- 8 Bojkowski CJ, Arendt J. Factors influencing 6-sulphatoxymelatonin – a major melatonin metabolite in normal human subjects. Clin Endocrinol 1990; **33**:435–44.
- 9 Mathews CD, Guerin MV, Wang X. Human plasma melatonin and urinary 6-sulphatoxymelatonin: studies in natural animal photoperiod and in extended darkness. Clin Endocrinol 1991; **35**:21–27.
- 10 Kovacs J, Brodner W, Kirchlechner V, Arif T, Waldhauser F. Measurement of urinary melatonin: A useful tool for monitoring serum melatonin after its administration. J Clin Endocrinol Metab 2000; 85:666–670.
- 11 Eldar-Geva T, Hurwitz A, Vecsei P, Palti Z, Milwidsky R, Rösler A. Secondary biosynthetic defects in women with late-onset congenital adrenal hyperplasia. N Engl J Med 1990; **323**:855–863.
- 12 White PC, Speiser PW. Congenital adrenal hyperplasia due to 21-hydroxylase deficiency. Endocrinol Rev 2000; **21**:245–291.

- New HT, White PC, Pang S, Dupont B, Speiser PW. The adrenal hyperplasias. In: The metabolic basis of inherited disease, Seriver CR, Beaudet AL, Sly WS & Valle D, eds. pp 1881–1917, 6th ed. Vol 2, New York: McGraw-Hill, 1989.
- 14 Lopes LA, Catzeflis C, Cicotti I, Rey C, Sizonenko PC. Plasma 3 alpha-androstanediol glucoronide in normal children and in congenital adrenal hyperplasia due to 21-hydroxylase deficiency. Horm Res 1997; **48**:35–40.
- 15 Ferriman D, Gallwey JD. Clinical assessment of body hair growth in women. J Clin Endocrinol Metab 1961; **21**:1440–1447.
- 16 Paoletti AM, Cagnacci A, Orru M, Ajossa A, Guerriolo S, Melis GB. Treatment with flutamide improves hyperinsulinemia in women with idiopathic hirsutism. Fertil Steril 1999; **72**:448–453.
- 17 Ehrmann DA, Barnes RB, Rosenfeld RL. Polycystic ovary syndrome is a form of functional ovarian hyperandrogenism due to dysregulation of androgen secretion. Endocr Rev 1999; 16:322–353.
- New MI, Lorenzen R, Lerner AJ, Kohn B, Oberfield SE, Pollack MS, Dupont B, Stoner E, Levy DJ, Pang S, Levine LS. Genotyping steroid 21-hydroxylase deficiency: hormonal data. J Clin Endocrinol Metab 1983; 57:320–326.
- 19 Oriola J, Plesa I, Machuca I, Pavia C, Rivera-Fillat F. Rapid screening method for detecting mutations in the 21 hydroxylase gene. Clin Chem1997; 43:557–561.
- 20 Wedell A. Molecular genetics of congenital adrenal hyperplasia (21-hydroxylase deficiency): implications for diagnosis, prognosis and treatment. Acta Paediat 1998; 87:159–164.
- 21 Bistran BR, Blackburn GL, Sherman M. Therapeutic index of nutritional depletion in hospitalized patients. Surg Gynec Obst 1975; 141:512–518.
- 22 Luboshitzky R, Shen-Orr Z, Ishai A, Lavie P. Melatonin hypersecretion in male patients with adult-onset idiopathic hypogonadotropic hypogonadism. Exper Clin Endocrinol Diab 2000; 108:142–145.
- 23 Waldhauser F, Frisch H, Krautgasser GA, Sparotti A, Schober E, Bieglmayer C. Serum melatonin is not affected by glucocorticoid replacement in congenital adrenal hyperplasia. Acta Endocrinol 1986; 111:355–359.
- 24 Terzolo M, Piovesan A, Ali A, Codegone A, Pia P, Reimondo G, et al. Circadian profile of serum melatonin in patients with Cushing's syndrome or acromealy. J Endocrinol Invest 1995; **18**:17–24.
- 25 Weaver DR, Stehle JH, Stopa EG, Reppert SM. Melatonin receptors in human hypothalamus and pituitary implications for circadian and reproductive responses to melatonin. J Clin Endocrinol Metab 1993; 76:295–301.
- 26 Cagnacci A, Paoletti AM, Soldan R, Orru M, Maschio E, Mellis GB. Melatonin inhances the luteinizing hormone in the follicular, but not in the luteal menstrual phase. J Clin Endocrinol Metab 1995; 80:1095–1099.
- 27 Caglayan S, Ozata M, Ozisik G, Turan M, Bolu E, Oktenli C, Arslan N, Erbil K, Gul D, Ozdemir IC.. Plasma melatonin concentration before and during testosterone replacement in Klinefelter's syndrome: Relation to hepatic indolamine metabolism and sympathoadrenal activity. J Clin Endocrinol Metab 2001; 86:738–743.
- 28 Delfs TM, Baarks S, Fock C, Schumacher M, Olcese J, Zimmerman RC. Sex steroids do not alter melatonin secretion in humans. Hum Reprod 1994; 9:49–54.
- 29 Luboshitzky R, Qupti G, Ishai A, Shen-Orr Z, Futerman B, Linn S. Increased 6-sulfatoxymelatonin in women with polycystic ovary syndrome. Fertil Steril 2001; **76**:506–510.
- 30 Luboshitzky R, Dharan M, Goldman D, Herer P, His Y, Lavie P. Seasonal variations of gonadotropins and gonadal steroids receptors in the human pineal gland. Brain Res. Bull. 1997; **44:**665–670.
- 31 Leibenluft E, Schmidt PJ, Turner EH, Danaceau MA, Ashman SB, Wehr TA, Rubinow DR. Effects of leuprolide-induced hypogonadism and testosterone replacement on sleep, melatonin and prolactin secretion in men. J Clin Endocrinol 1997; **82**:3203–3207.
- 32 Ozata M, Bulkur M, Bingal N, Beyhan Z, Corakci A, Bolu F. et al. Daytime plasma melatonin levels in male hypogonadism. J Clin Endocrinol Metab 1996; **81**:1877–1881.