

Neuroendocrine and cytokines-induced responses to minutes, hours, and days of mental stress

Khalid Z. Matalka

Faculty of Pharmacy and Medical Technology, University of Petra, Amman, JORDAN.

Correspondence to: Khalid Z. Matalka, Ph.D.
Faculty of Pharmacy and Medical Technology,
University of Petra,
P. O. Box 961343, Amman, JORDAN,
EMAIL: kzm@uop.edu.jo or kzm@go.com.jo
FAX: +962 6-571-5570, TEL: +962 6-571-5546

Submitted: April 24, 2003

Accepted: June 2, 2003

Key words: **mental stress; examination; cytokines; HPA axis and SAM system**

Neuroendocrinol Lett 2003; 24(5):283-292 pii: NEL240503R01 Copyright © Neuroendocrinology Letters www.nel.edu

Abstract

OBJECTIVES: Previously, a large number of studies reported that psychological stress and psychiatric illness reduces immune responsiveness. However, it turned out that stress reduces immune responsiveness is an oversimplified statement because the interactions between central nervous system, endocrine system and the immune system are undoubtedly complex. Therefore, this study aims in reviewing mental stress models (e.g. brief and written examination stress as subacute and acute type of stressor) that have been utilized to understand the effect of stress on the neuroendocrine and immune systems.

METHODS: The published findings from human mental stress models on catecholamines, cortisol, prolactin levels and on T helper (Th) 1 and 2-induced cytokines are presented and discussed with respect to the *in vitro* and *in vivo* effects of glucocorticoids, catecholamines, and prolactin on the induction of cytokines.

RESULTS: This review shows evidence that short-time (minutes) or preparation to a written examination, in those students who are stressed, induces the production of proinflammatory cytokines which may be related to Th1 response. However, longer mental stress (days) causes dysregulation in the immune function by shifting the cytokine response to Th2 response.

CONCLUSIONS: The outcome from neuroendocrine and immune function prior to, following and after mental stress depends on multiple variables most importantly on the amount of stress, exposure time, coping behavior and adjustment of the individual. A few minutes of stress may improve immune performance but longer times of mental stress have detrimental effects that may lead to loss of immune integrity. Furthermore, studies on stress and common health problems are necessary to increase our knowledge and understanding of the mechanisms responsible for producing neuroendocrine-induced immune changes in health and common diseases.

1. Introduction

Stress is the reaction of the body to stimuli that disturb its homeostasis. These stimuli could be physical, chemical, or psychological which commonly stimulate the hypothalamic-pituitary-adrenal (HPA) axis, leading to increasing levels of serum glucocorticoids (and other hormones), and peripheral sympathetic/adrenomedullary (SAM) system followed by release of catecholamines [1]. Glucocorticoids and catecholamines and other hormones (e.g. prolactin) were shown to have detrimental effects on the immune system via their expressed receptors on the immune cells. Most of these detrimental affects are on cytokines produced from immune cells.

The immune response is composed of cell-mediated and humoral components. The balance between cell mediated and humoral responses is basically maintained by the release of cytokines from the T helper (Th) lymphocytes. Th lymphocytes are divided into two subpopulations, Th1 and Th2 cells, based on their ability to produce specific pattern of cytokines [2,3]. Th1 cells induce cell-mediated immunity via their release of cytokines such as interleukin (IL)-2 and interferon (IFN)- γ while Th2 cells induce humoral immunity via their release of cytokines such as IL-4, IL-5, and IL-10. However, naive T helper cells (Th0) serve as precursors to either Th1 or Th2 cells depending on the signal of activation. Cytokine such as IL-12, produced by activated monocytes/macrophages or other antigen presenting cells, is a major inducer of Th1 cell and its cytokines. Monocytes/macrophages-derived IL-12 and tumor necrosis factor- α (TNF- α) with natural killer (NK) cells and Th1-derived IFN- γ , stimulate the function of T cytotoxic cells, NK cells, and activated macrophages. The cytokines such as, IL-12, IFN- γ and TNF- α , are considered major inflammatory cytokines because they stimulate the synthesis of nitric oxide and other inflammatory mediators that derive chronic delayed hypersensitivity reactions [2,3]. While IL-12 and IFN- γ can inhibit Th2 response, Th2 cytokines such as, IL-10 and IL-4, inhibit Th1 activity and macrophage activation. In addition, they stimulate differentiation of B cells to antibody-producing cells (especially class switching to IgE) and stimulate the growth and activation of eosinophils and mast cells, but inhibit macrophage activation. Therefore, Th1 and Th2 responses are mutually inhibitory [4]. Additionally, T cytotoxic cells can secrete Th1 as well as Th2 cytokines and were referred as Tc1 and Tc2.

Previously, a large number of studies reported that psychological stress and psychiatric illness reduces immune responsiveness. However, it turned out that stress reduces immune responsiveness is an oversimplified statement. Many studies have shown the impact of different types of stressors on CNS, endocrine and the immune system. For instance, surgery [5], depression [6,7], bereavement [8], exercise [9], marital conflict [10–12] and academic stress [13–23] were some of the stress models used. However, each of the above stress model differs basically in duration and intensity, thus

different responses from the CNS, endocrine, and the immune systems would develop. Also, within the same type of stressor used, time of testing or challenge, subjects and their health status, coping-behavior, age, sex, lymphoid compartment examined and seasonal variations all could play a major influence on the outcome measured and therefore the data in the literature seem conflicting.

In this review, the effects of different models of mental and academic stress (as subacute to acute type of stress) on the levels of catecholamines, cortisol, prolactin, Th1 and Th2-induced cytokines are presented and discussed. The discussion is performed with respect to the *in vitro* and *in vivo* effects of glucocorticoids, catecholamines, and prolactin on the induction of cytokines. This review shows evidence that short-time (minutes) or preparation to a written examination, in those subjects who are stressed, induces the production of proinflammatory cytokines and maybe related to Th1 response. The brief mental stress or preparation to exam (depending on the type of exam and duration of preparation) is more likely to induce a challenge (i.e. potential for growth [24,25]) to the individual and causes a mild and transient SAM and HPA activation, which is correlated with self-confidence and anxiety. This mild and transient increase in catecholamines, glucocorticoids and prolactin were found to induce the production of proinflammatory cytokines probably via the induction of transcription nuclear factor (NF- κ B). However, longer mental stress (days) causes dysregulation in the immune function through shifting towards Th2 mediated response. During repeated examinations, for instance, the stressed individuals are exposed to repeated stressors without recovery period and the threat feeling or losing is eventually higher and it is more likely to induce a moderate and consistent HPA activation, which follows SAM activation, and thus consistently glucocorticoids are increased [24,25]. The latter form is more likely to induce an immune dysregulation by shifting the cytokine response into Th2 rather than Th1 response. However, this dysregulation might be limited if other hormonal levels like prolactin are increased. Moreover, the aim is to provide a resource for helping in the formulation of strong rationales for the design of future stress studies which will help understanding the mechanisms responsible for producing neuroendocrine-induced immune changes in health and different disease conditions.

2. Effect of Glucocorticoids, Catecholamines and Prolactin on Cytokines-Induced Levels

It is well known that glucocorticoids and their analogs down regulate the immune system via binding glucocorticoid receptors on lymphocytes and monocytes/macrophages [26]. This down regulation consisted mainly of inhibiting mitogen or antigen-induced levels of IL-1 [27], IL-12 from monocytes/macrophages [28,29], and IL-2, IFN- γ from T lymphocytes [30,31]. In addition, glucocorticoids down regulate the expres-

sion of IL-12 receptors on Th1 and NK cells [28]. However, glucocorticoids showed different effects on IL-4 and IL-10 induction levels. In phytohemagglutinin (PHA)+ lipopolysaccharide (LPS)-stimulated whole blood or PHA-stimulated peripheral blood mononuclear cells (PBMC), glucocorticoids inhibited the production of IL-10 induction less than IFN- γ causing a sharp decrease in IFN- γ /IL-10 ratio [32,33,34]. However, when the same cells were further stimulated with IL-2, IL-4 and IL-10 levels increased while IFN- γ levels decreased [34]. Furthermore, the addition of glucocorticoid-treated monocytes/macrophages to antigen-primed CD4+ T cells was associated with increased IL-4 production levels [35,36]. In addition, glucocorticoids *in vitro* have no effect on the production of IL-10 from LPS-stimulated monocytes [29]. In endotoxemia and multiple sclerosis patients treated with glucocorticoids there was increasing plasma levels of IL-10 [37,38]. These results indicate that glucocorticoids inhibit directly the production of IL-12 from monocytes/macrophages and thereby inhibiting production from IFN- γ from Th1/NK cells, but have less effect on IL-10 productions. The IL-10 productions might increase, however, if concomitant with another stimulatory signal to T cells. In this case, the levels of Th1 and Th2 cytokines are decreased and increased respectively, and thus down regulation of Th1 persists. The stimulatory signals could be cytokines, such as IL-2, or other hormones e.g. estradiol which results in increase in IL-10 levels [33,34]. On the other hand and following brief *in vitro* exposure to glucocorticoids, mitogen-stimulated PBMC increased IFN- γ and IFN- γ /IL-10 ratio [34]. In addition, brief exposure to cortisol (2 h) in rats increased delayed type hypersensitivity reaction (DTH) [39] and found to be mediated by local increase in IFN- γ production [40].

Catecholamines, the end products of SAM system also regulate immune system response through specific α (α 1 and α 2) and β (β 1 and β 2) receptors, which are categorized, based on their different sensitivity to certain agonists. β 2 receptors, for example, have been identified only on Th1 cells, but not on Th2 cells [41]. *In vitro*, catecholamines or β and β 2 adrenoceptor agonists induced a decrease in IL-12 and IFN- γ and an increase in IL-10, IL-4 and IL-5 levels [29,42,43]. These effects were prevented by β -adrenoceptor antagonist and IL-12. In one of these studies [43], salbutamol, a β 2 adrenoceptor agonist, inhibited IL-12 production but not IL-1 α and β , IL-6, or IL-10 in IFN- γ -primed LPS-stimulated monocytes or whole blood cultures. In addition, β 2 adrenoceptor agonists seemed to inhibit neonatal T cells to differentiate to Th1 cells but promote Th2 differentiation [43]. Moreover, administration of β 2-adrenoceptor agonists in healthy volunteers elevated plasma IL-6 [44], suppressed the production of IL-12 from IFN- γ -primed LPS-stimulated whole blood [43] and a massive release of catecholamines following acute brain trauma increased IL-10 levels [45]. Furthermore, the α adrenoceptors are also involved in cytokines alteration. *In vitro* α 2 adrenoceptor agonists decreased IFN- γ and IFN- γ /IL-10 ratio in PHA+LPS stimulated whole

blood cultures [46]. However, administration of α 2 versus β -adrenoceptor agonists in LPS-treated mice gave contrasting patterns on IL-10 plasma levels. β -adrenoceptor agonists or α 2 adrenoceptor antagonists increased IL-10 plasma whereas α 2 adrenoceptor agonists or β -adrenoceptor antagonists decreased IL-10 plasma levels [47]. These results could be explained by the fact that LPS does not induce the production of IFN- γ , and the only source of IL-10 in LPS-treated whole blood or PBMC is monocytes [31,32]. Therefore α and β -adrenoceptors expressed on monocytes/macrophages play different roles on IL-10 induction [47]. However, brief exposure to epinephrine (2 h) in rats increased DTH [39] and found to be mediated by local increase in IFN- γ production [40].

Prolactin is another hormone, which has an important role in immune regulation [48,49]. A great deal of evidence suggests that lymphocytes and immunocompetent cells from thymus, spleen and peripheral lymphocytes contain prolactin mRNA and these cells release a bioactive prolactin which is similar to pituitary prolactin [49–51]. Furthermore, it has been shown that lymphocytes contain dopamine receptors, such as D4 and D5, which may be involved in regulation of prolactin release from lymphocytes [52,53]. It has been shown that lymphocyte proliferation and macrophage activation is reduced by either antibodies against prolactin or suppression of prolactin release from the pituitary [48,54]. However, the proliferation of lymphocytes and IL-2 production from hyperprolactinemia patients was decreased [56]. *In vitro*, prolactin enhanced IFN- γ activity from PBMCs and NK cells [55], and increased IL-12 and IFN- γ productions from PHA+LPS-stimulated whole blood [32]. The latter was not seen in LPS-stimulated whole blood, however, it demonstrated an increase in IL-10 levels, which indicates that prolactin effects on cytokines induction is stimulus specific [32].

3. Mental Stress Models

In this review, the stress models are categorized as a brief mental stressor, one written exam and multiple exams and the time of testing the stressed individuals. For a brief mental stress (i.e. minutes of mental stress), a speaking stressor or laboratory (Stroop test) or solving a difficult puzzle stress tests were used. For a written examination (i.e. hours of mental stress), some researchers used the time 24h prior to one exam to test the stressed individual, others used 0.5–1h (just) before examination, immediately after examination or 24–48 h following an exam. The last two models described in the literature were during examinations period and 48-h post examinations period (i.e. days of mental stress).

All the studies presented here were performed on college students (or otherwise stated). The parameters tested were compared between blood samples drawn at the “stressed” time (stressed samples) to pre- and/or post levels (baseline samples). For written examination, the pre-stress samples were drawn at the

beginning of a semester or 2–4 weeks before examination. The post-stress samples were drawn 10–30 days following examination.

3.1 Effect of Mental Stress on Cortisol Levels

Minutes after beginning a laboratory stressor test or oral exam, adrenocorticotrophic hormone (ACTH) and cortisol levels rose significantly when compared to levels just before examination. However these values went back to baseline after the exam was finished [57,58]. It has to be mentioned that the increase in cortisol was delayed few minutes after increase in ACTH, which indicates a descending activation of HPA. Furthermore, twenty-four hours prior to examination, cortisol levels were either not changed (also in students who have high stress scores) [59] or significantly higher [60]. In this respect, the release of cortisol depends on the time of testing and magnitude level of the stressor [61]. Also, there are students' variations regarding exam preparation and time of anxiety, therefore, in some cases drawing blood 24 h prior to exam may be early to see an endocrine effect because the elevation of cortisol is transient and might be missed in non-kinetic studies. This was clearly observed when blood was drawn immediately before examination where cortisol levels significantly increased [61–63] and became even higher immediately after the exam [63]. In addition, urine excretion of cortisol was increased during and immediately after a 6-h exam and was more in males than females [64,65]. During examinations period, the average ACTH levels from male students were significantly increased (38%, 1 pmol/L) during the day of the fall time but not the spring-time [66]. This increase, however, was not sufficient to increase cortisol levels. The adrenal sensitivity to ACTH in dogs was found that a 2 pmol/L change in ACTH increased cortisol level by 55 nmol/L [67] and ACTH stimulation (1 mg) in humans significantly increased cortisol levels (~30 nmol/L) after one hour of injection [68]. In Malarkey et al [66] study, however, when students were categorized according to their stress scores, the average cortisol level significantly increased (~28 nmol/L) during examination periods which was accompanied with ~1.4 pmol/L increase in ACTH. This study suggests that a strong stimulation and or the less coping behavior of the responder is of critical importance in determining the endocrine profile to stress. Furthermore, during examinations period, also a significant increase of cortisol levels was observed in female students [22]. These results indicate that minutes, hours or days of mental stress stimulate HPA axes and resulted in increasing cortisol level. This increase in cortisol levels, however, was not seen days after ending examinations period [21,69]. The latter suggest that cortisol levels subsides when the stressor ends (half life of cortisol 1–1.5 h) or such studies are in need of collecting urine for period of time (6–12 h) to detect differences in cortisol values.

3.2 Effect of Mental stress on Catecholamines Levels

Minutes after beginning a laboratory stressor test or oral presentation, levels of catecholamines (epinephrine and norepinephrine) increased significantly when compared to levels just before examination [58,70,71]. However these values went back to baseline after the exam was finished [58,70,71]. Furthermore, during an examination plasma and urine catecholamine levels were also increased [64,65,72,73]. In addition, Powlak et al [71] demonstrated an increase in β 2-adrenoceptors on PBMC following a 10-min stress. These studies also showed that epinephrine excretion was higher in males more than females [64,65,72]. Overall, these results showed that minutes or hours of mental stress are able to stimulate SAM and thus catecholamine levels are increased. Feeling of success and confidence, however, were more common in males than females and high discomfort was correlated with poor performance in males but with good performance in females [64,65,72]. These results suggest that coping behavior adopted by males and females during stressful situations are different. Males seemed to be more dependent on SAM as well as HPA activation when confronted with challenging situations. However, females tend to be more sensitive to sympathetic stimulation than men [64].

3.3 Effect of Mental stress on Prolactin Levels

Prolactin plasma levels were also studied following mental stress. Just before examination, male students showed a significant increase in prolactin levels but no change was seen in female students when compared to baseline levels [14]. However, immediately after an exam and in males only prolactin levels were significantly less than baseline levels. In addition, Meyerhoff et al. [57] have shown that minutes after beginning an oral examination, prolactin levels were increased in young males when compared to values just before examination. This increase, however, went back to baseline levels after the exam was finished. During and after examinations period, prolactin levels in males did not change when compared to baseline levels [69,74], however, in female students prolactin levels showed a tendency to increase [22]. These results indicate two things: first, males and females behaves differently in prolactin secretion upon response to stress which could be due to differences in prolactin regulatory secretions though tuberoinfundibular dopaminergic system such as dopamine, norepinephrine, serotonin, endorphins, estrogen and prolactin itself [75]. Second, the differences in stress duration (one exam versus examinations period) may enable other hormones or peptides, like estradiol, to cause significant change in prolactin level especially in female students [75]. More studies regarding the effect of mental stress (short versus long times of mental stress) on prolactin in both sexes are mandatory to establish a link between student behavior during such stress and prolactin levels.

3.4 Effect of Mental Stress on the Induced-Cytokines Level

The effect of mental stress on cytokines production from mitogen-stimulated blood cells either using PBMCs or whole blood was studied. Twenty-four hours prior to examination, Maes et. al. [76] reported that mitogen-stimulated whole blood at 24 h prior to examination significantly produced higher levels of IFN- γ , TNF- α , IL-6, and IL-10 than baseline. However, when students were divided according to stress perception and anxiety, subjects with higher stress perception and anxiety showed even higher production of IFN- γ , TNF- α and IL-6. However, mitogen-stimulated blood from students with low anxiety scores produced significantly higher levels of IL-10, IL-5 and IL-4 than from students with high anxiety. In other words, the results of Maes et al [76] suggest that students who are responding to stress and anxiety show a proinflammatory response and those students who are capable to cope with such stress or less anxious show a Th2 response. In a similar model, Guidi et. al. [60], reported a significant reduction in lymphocyte proliferation and IL-2 production and an increase in cortisol levels. Just before examination, however, it was found that production of IL-1 α , IL-10 and IL-6 from mitogen stimulated whole blood was increased, IFN- γ level was decreased, and no change in TNF- α production [77]. A similar effect immediately after examination was seen when stimulated monocytes produced significantly higher levels of IL-1 β when compared to baseline samples but stimulated PBMCs produced significantly less IFN- γ . These changes in IL-1 α and IFN- γ returned to baseline values within 10 days [78]. Twenty four hours after an exam, Uchakin *et al.* [79] reported that the percentage of IL-2 producing cells (CD4+ and CD8+) and CD8+ IFN- γ cells and IL-2 production levels was significantly lower than in non-stressed samples but no change in IFN- γ and IL-10 productions were observed.

During examinations period, IFN- γ mRNA, IFN- γ [16,17], IFN- γ (only from PBMC but not whole blood) and IL-2 [80] productions were less in stressed samples, IL-4 and IL-5 did not change [80], but higher IL-6 levels were observed when compared to baseline values [80]. From adolescence students (~16 y), examinations period reduced IL-4 and IL-5 productions in healthy but not in asthmatic subjects [81]. Furthermore, Marshall *et al.* [21] studied synthesis and release of IFN γ and IL-10 from stimulated PBMCs 48 h post examination periods and found a significant increase (87%) in IL-10 production but insignificant decrease in IFN- γ and was correlated positively to number of hassles. Subjects who reported more hassles and greater subjective adjustment to hassles at pre-exam had higher IL-10 levels and lower IFN- γ /IL-10 ratio at both pre exam and 48 h post examinations period [21]. The above results may indicate that the coping behavior or health status (e.g. asthmatic) of the individual at baseline has a major effect on immune changes observed during the stress period [21,76,81]. Individuals who were at baseline (pre-stress) shifting toward Th2 response (high hassle group or asthmatic) will be less responsive to stress

than those individuals who have a predominant Th1 response (low hassle group or healthy individuals). In addition, students who are less anxious before examination produce more of a TH2 response [76].

These results suggest that cytokines production pattern is altered prior to, after one exam, during and post examinations period. Proinflammatory cytokines levels such as IL-1 β and IL-6 are increased in almost all the mental stress conditions [76–78,80]. IL-1 β and IL-6 are produced from immune and non-immune cells and in the studies mentioned here, their increasing levels are mainly from mitogen-stimulated monocytes. For TNF- α , the inconsistent results could be due to TNF genes polymorphism between individuals. However, the pattern of Th1 versus Th2-induced cytokines seemed to be different depending on when the blood sample was drawn, age of the students examined and in the students who reacted with such type of stress (high perceived stress scores, anxiety or hassles). The discussion below will provide an explanation of how transient activation of SAM and/or HPA resulted from brief stress will lead to increase in proinflammatory cytokines, which may be related to Th1 response. On the other hand, days of mental stress results in consistent activation of SAM and HPA which will lead to Th2 shift.

3.5 Minutes Versus Days of Mental Stress

Many important issues have emerged while writing this review. First, does brief mental stress (minutes) or 24 h prior to one major exam induce a challenge to the individual and more likely be a preparation of the stressed or the challenged individual to cope with the stressor? This is suggested because minutes of stress or preparation to exam caused transient increase in NK cells [73, 82–85], increase in CD8+, CD2+CD26+, CD2+HLADR+ cells [62] increase in IFN- γ , TNF- α , IL-6, IL-1 and IL-10 production from stimulated immune cells [76,77], and increase in S-IgA [63,86,87] (Table I). Similarly, a brief restrained stress (2h) in animal skin hypersensitivity (DTH) model enhanced skin immune function by increasing drainage of T lymphocytes from lymph nodes and increased local IFN- γ , which was abrogated by adrenalectomy [39,40]. In the latter condition, a similar pattern was seen when rats were exposed to low levels of cortisol or epinephrine. In addition, brief exposure of PBMC to glucocorticoids caused an initial increase in IFN- γ and IFN- γ /IL-10 ratio [34]. A very recent work has shown that a 15-min of laboratory stressor test increased ACTH, cortisol, epinephrine and norepinephrine [58]. This was paralleled with an increase in redox-sensitive NF-kB induction from PBMC. This increase in NF-kB activity went back to normal within 60 minutes as well as the hormones. Subjects who did not show any stress-dependent increase in stress hormones did not induce NF-kB binding activity indicating that the latter response depends on the response to psychological stress [58]. Thus, timing and duration of stress may significantly affect the nature, enhancing or suppressing, of stress influence

Table I. The immune parameters outcome following short mental stressors (minutes) or during preparation to an exam:

Immune parameter	Outcome	References
Antibodies:		
S-IgA (Saliva)	Increased	63, 86, 87
Immune Cells:		
Total leukocytes	Increased	59, 83
Total lymphocytes	Increased	59
Monocytes	Increased	59
NK cells	Increased	71, 82, 83, 84, 85
CD8 +	Increased	59, 83, 84, 85
CD4/CD8 ratio	Decreased	59, 83, 84, 85
CD2CD26+	Increased	59
CD2+	Increased	59
CD2+HLADR+	Increased	59
Transcription Factors		
NF-kB	Increased	58
Cytokines:		
IFN- γ	Increased	76
IL-6	Increased	76
TNF- α	Increased	76
IL-10	No change*	76
IL-4	No change*	76
IL-5	No change*	76

* See text for more details

on the immune system. The low concentration and or exposure time of stress hormones (catecholamines and cortisol) may enhance immune function by informing the immune system about the impending challenges such as infection [88]. Further studies are needed to prove if brief mental stress protects individuals from infections, e.g. respiratory.

Second, if catecholamines, cortisol and prolactin increasing levels during different times of mental examinations are concomitant then the effect on leukocytes, leukocyte subsets, Th1 and Th2 mediated responses would be different. Each of the mentioned hormones has different effects on leukocytes distribution, lymphocyte responses upon challenge and Th1 and Th2 cytokine release. In addition, catecholamines may enhance the intensity of the cortisol signal by increasing either cortisol receptor activity [89] or the transfer of occupied glucocorticoid receptor or transcription factors to the nucleus [90,91], and cortisol might either induce an effect on β 2 adrenergic receptor expression on the immune cells surface to increase the number of receptors available for stimulation [92]. Prolactin, on the other hand, reverses suppression of IFN- γ and IL-12 production induced by cortisol [32]. Therefore, the ratio of catecholamine/cortisol, catecholamines/prolactin, cortisol/prolactin would be an important factor to measure in mental stress.

Third, the stressed individual during days of examinations period is more likely to feel more threatened and the potential of loss is higher [24,25]. Furthermore, stress from examinations period seemed to induce a Th2 mediated response upon immune cells challenge (Table II). It may be proposed that these immune effects reported in these studies are attributable to increased

Table II. The immune parameters during or after days of mental examination.

Immune parameter	Outcome	References
Antibodies:		
S-IgA (Saliva)	Decreased	94, 95, 96, 97, 98, 99
EBV IgG	Increased	15, 18, 19, 23, 100
EBV IgG (Saliva)	Increased	101
EBV IgA (Saliva)	Increased	101
Immune Cells:		
Total leukocytes	No Change	22
Total lymphocytes	Decreased	22, 62
Monocytes	No Change	22, 62
NK cells	Decreased	94
Cytokines:		
IFN- γ	Decreased	16, 17, 21, 77, 78
IL-2	Decreased	80
IL-6	Increased	77, 78, 80, 81
IL-10	Increased	21
IL-1 β	Increased	78
TNF- α	No Change	77
IL-4	No Change	80, 81
IL-5	No Change	80, 81
IL-5 (sputum cells)	Increased	109

and/or prolonged exposure to endogenous cortisol and catecholamines. As it has been shown that prolonged stress (days) shifts the cytokine response towards Th2 mediated response and thus cell-mediated immunity is dysregulated. This dysregulation increased susceptibility to viral infection as it was found that the rate of respiratory infection (e.g. common cold), increased in a dose-response manner with increases in the degree of psychological stress in individuals who had been intentionally exposed to rhinoviruses [93]. During examinations period, NK cell activity was reduced [94] and S-IgA secretions and concentrations were lowered [95–99]. This reduction of S-IgA remained for 6–14 days following examination period [97,98]. The above studies correlated lower S-IgA secretion rate and concentration to high stress [95,96] and strong power motives especially with prolonged increase in salivary norepinephrine [86]. In addition, students with social support or characterized to maintain warm personal relationship had consistently higher S-IgA at baseline (pre stress) and also at stress periods than those with less social support indicating that social support might enhances health outcomes irrespective to stressful experiences [95,96].

Furthermore, latent herpesvirus reactivation also indicates dysregulation of Th1 mediated response. Several studies showed that specific IgG concentrations against EBV viral capsid antigen (VCA) were increased during examinations period [15,18,19,23,100]. Similarly, a recent study has shown a significant increase in salivary IgG and IgA against EBV during examination period [101]. In addition, the effect of mental stress on specific IgG concentrations against CMV and HSV1 and HHV-6, was also studied [15,99,23] but failed to

show a significant increase in viral-specific antibody concentration. The increase in IgG specific antibodies against latent herpes viruses indicates latent viral reactivation. Even though this might not be a complete viral reactivation [18], it is still seen more in EBV and not other latent herpesviruses. It has to be mentioned that *in vitro* studies were almost always successful in showing reactivation of herpesviruses following exposure with pharmacological doses of cortisol [102–106], catecholamines [107], and both cortisol and catecholamines [108], but many *in vivo* studies, such as stress studies, failed to show such reactivation. It is evident that where the virus becomes latent (cell type), hormonal influence (e.g. ↑cortisol, ↑catecholamines, ↓prolactin), cytokines (e.g. ↓IFN- γ and ↑IL-6), mechanism of replication, immune control and other factors play an important role in latent viral reactivation. For example, even though not statistically significant, the percent change in EBV VCA IgG levels showed a negative correlation ($r = -0.457$) with the percent increase in prolactin levels during examination stress [23]. The latter means that stress-induced prolactin might help in controlling viral reactivation. Further studies are needed to understand the mechanisms of stress on latent viral reactivation.

In allergic and asthma responses, the inflammatory response is mediated by Th2 cytokines, particularly, IL-4 and IL-5. Recent studies suggest that catecholamines directly affect Th1 and not Th2 cells cytokine production and function through β_2 adrenergic receptor that are expressed on Th1 cells [41]. Catecholamines and β_2 adrenergic receptor agonists inhibited IFN- γ production from Th1 cells and increased IL-10, IL-4 and IL-5 production from Th2 cells [42,43]. Also, cortisol suppresses Th1 cytokines production and may even induce an increase in Th2 cytokines depending on stimulatory signals to T cells [31–36]. Therefore, if mental stress (examinations period) can constantly elevates catecholamine and cortisol levels and these hormonal concentrations can decrease IFN- γ , increase IL-4 production (in low stressed individuals) and induce B cell responsiveness to produce IgE, then does long-term effect of stress increase vulnerability to allergies? It has been shown that examination period did not exhibit an exam-related drop in lung function in healthy or patients with asthma [94]. However, when sputum samples were collected during hours of examination stress, eosinophils and eosinophils-derived neurotoxin and IL-5 levels increased significantly [109]. Thus, examination stress may act as cofactor to increase the airway inflammation and a local Th2 mediated response especially in those individual who become exposed to a virulent respiratory pathogen at the time of stress, or in individuals with poor managed asthmatic or under chronic type of stress. This opens a challenge for future studies to resolve such common abnormality under also common psychological challenge.

4. Conclusions

The above review describes the modulations in the immune system following minutes, hours and days of mental stress, which is considered as subacute to acute type of stressor. A chronic type of stressors, as caregiving of persons with dementia, also caused a shift from Th1 to Th2 response by demonstrating an increase in IL-10+ CD4 and CD8 T cells [110]. This shift was also negatively related to age i.e. the pattern of change was higher with younger than older caregivers, which indicates an age related influence on Th1 to Th2 shift. In addition, parents of cancer patients were found to be resistant to glucocorticoids to suppress *in vitro* production of IL-6, one of the proinflammatory cytokines [111].

In subacute or acute mental stress, the following points are concluded: 1) minutes of mental stress induces the production of proinflammatory cytokines (a Th1 like response) via a mild and transient increase in catecholamines and cortisol, 2) a strong stimulation, coping behavior and adjustment of the responder is of critical importance in determining the endocrine profile to stress, 3) males seemed to behave differently than females during or following mental stress and thus SAM as well as HPA activation to induce catecholamines, cortisol or prolactin, respectively, are different when confronted with challenging situations especially with long term stress. 4) coping behavior (high versus low anxiety or hassle and health status) of the individual at baseline (prior to stress) have a major effect on immune changes observed during the stress period, 5) hours of mental stress may cause shift in Th1/Th2 ratio however, this depends on multiple variables such as the amount of successful coping and adjustments to the stress, 6) the dysregulation to Th2 mediated response is more evident in days or longer of stress.

The effects of stress on neuroendocrine and immunity are undoubtedly complex. Evidence from mental stress models suggests that such stress can induce changes in the SAM, endocrine and immune systems. However, whether these changes are only unidirectional (SAM-Immune or HPA-Immune), or bi-directional is still far from clear. Further studies in mental stress should provide a rational on how individual state of mind/SAM/HPA/immune system interacts. Furthermore, studies on stress and common health problems are necessary to increase our knowledge and understanding of the mechanisms responsible for producing neuroendocrine-induced immune changes in health and common diseases.

Acknowledgements

This work is supported by a grant # 5/3/2000 from the Deanship of Research at University of Petra-Amman, Jordan.

REFERENCES

- 1 Khansari DN, Murgo AJ, Faith RE. Effects of stress on the immune system. *Immunol Today* 1990; **11**:170-175.
- 2 Abbas AK, Murphy KM, Sher A. A functional diversity of helper T lymphocytes. *Nature* 1996; **383**:787-793.
- 3 Mosmann TR, Sad S. The expanding universe of T-cell subsets: Th1, Th2 and more. *Immunol Today* 1996; **17**:138-146.
- 4 Elenkov IJ, Chrousos GP. Stress hormones, Th1/Th2 patterns, pro/anti-inflammatory cytokines and susceptibility to disease. *Trends Endocrinol Metab* 1999; **10**:359-368.
- 5 Schulte HM, Bamberger CM, Elsen H, Herrmann G, Bamberger AM, Barth J. Systemic interleukin-1 α and interleukin-2 secretion in response to acute stress and to corticotropin-releasing hormone in humans. *Eur J Clin Invest* 1994; **24**:773-777.
- 6 Smith RS. The macrophage theory of depression. *Med Hypothesis* 1991; **35**:298-305.
- 7 Maes M. Major depression and activation of the inflammatory response system. In *Cytokines, Stress, and Depression* (Dantzer et al., ed), pp 25-46, Kluwer Academic/Plenum Publishers, NY, 1999.
- 8 Schleifer SJ, Keller SE, Camerino M, Thornton JC, Stein, M. Suppression of lymphocyte stimulation following bereavement. *J Am Med Assoc* 1983; **250**:374-377.
- 9 Hoffman-Goetz L, Pedersen BK. Exercise and the immune system: a model of the stress response? *Immunol Today* 1994; **15**:382-387.
- 10 Kiecolt-Glaser JK, Fischer LD, Ogrocki P, Stout JC, Speicher CE, Glaser R. Marital quality, marital disruption, and immune function. *Psychosom Med* 1987; **49**:13-34.
- 11 Kiecolt-Glaser JK, Malarkey WB, Chee M, Newton T, Cacioppo J, Mao HY, Glaser, R. Negative behavior during marital conflict is associated with immunological down-regulation. *Psychosom Med* 1993; **55**:395-409.
- 12 Malarkey WB, Kiecolt-Glaser JK, Pearl D, Glaser R. Hostile behavior during marital conflict alters pituitary and adrenal hormones. *Psychosom Med* 1994; **56**:41-51.
- 13 Kiecolt-Glaser JK, Garner W, Speicher C, Penn GM, Holliday J, Glaser R. Psychosocial modifiers of immunocompetence in medical students. *Psychosomatic Med* 1984; **46**:7-14.
- 14 Johansson G, Laakso ML, Peder M, Karonrn SL. Initially high plasma prolactin levels are depressed by prolonged psychological stress in males. *Int J Psychophysiol* 1990; **9**:195-199.
- 15 Glaser R, Kiecolt-Glaser JK, Speicher CE, Holliday JE. Stress, loneliness, and changes in herpesvirus latency. *J Behav Med* 1985; **8**:249-260.
- 16 Glaser R, Rice J, Speicher CE, Stout JC, Kiecolt-Glaser JK. Stress depresses interferon production by leukocytes concomitant with a decrease in natural killer cell activity. *Behav Neurosci* 1986; **100**:675-678.
- 17 Glaser R, Lafuse WP, Bonneau RH, Atkinson C, Kiecolt-Glaser JK. Stress-associated modulation of proto-oncogene expression in human peripheral blood leukocytes. *Behav Neurosci* 1993; **107**:525-529.
- 18 Glaser R, Pearson GP, Bonneau RH., Esterling BA, Atkinson C, Kiecolt-Glaser, J. Stress and the memory T-cell response to the Epstein-Barr virus in healthy medical students. *Health Psychol* 1993; **12**:435-442.
- 19 Glaser R, Pearl DK, Kiecolt-Glaser JK, Malarkey WB. Plasma cortisol levels and reactivation of latent Epstein-Barr virus in response to examination stress. *Psychoneuroendocrinology* 1994; **19**:765-772.
- 20 Maes M, Hendriks D, Van Gastel A, Demedts P, Wauters A, Neel H, Janca A, Scharp'e S. Effects of psychological stress on serum immunoglobulin, complement and acute phase protein concentrations in normal volunteers. *Psychoneuroendocrinology* 1997; **22**:397-409.
- 21 Marshall GD, Agarwal SK, Liody C, Cohen L, Henninger EM, Morris GJ. Cytokine dysregulation associated with exam stress in healthy medical students. *Brain Behav Immun* 1998; **12**:297-307.
- 22 Matalka KZ, Sidki A. Academic stress - influence on leukocyte distribution, cortisol, and prolactin. *Lab Med* 1998; **29**:697-702.
- 23 Matalka KZ, Sidki A, Abdul-Malik S, Thewaini A. Academic Stress-Influence on Epstein Bar virus and cytomegalovirus reactivation, cortisol, and prolactin. *Lab Med* 2000; **31**:163-168.
- 24 Folkman S, Lazarus RS. If it changes it must be a process: Study of motion and coping during three stages of a college examination. *J Personal Soc Psychol* 1985; **48**:150-170.
- 25 Dienstbier RA. Arousal and physiological toughness: implications for mental and physical health. *Psychol Rev* 1989; **96**:84-100.
- 26 Munk A, Guyre PM. Glucocorticoids and immune function. In: *Psychoneuroimmunology*, 2nd ed (ed R. Ader,, D. L. Felton & N. Cohen), pp 447-474. Academic Press: New York, 1991.
- 27 Lew W, Oppenheim JJ, Matsushima K. Analysis of the suppression of IL-alpha and IL beta production in human peripheral blood mononuclear adherent cells by a glucocorticoid hormone. *J Immunol* 1988; **140**:1895-1902.
- 28 Wu CY, Wang K, McDyer JF, Seder RA. Prostaglandin E2 and dexamethasone inhibits IL-12 receptor expression and IL-12 responsiveness. *J Immunol* 1998; **161**:2723-2730.
- 29 Elenkov IJ, Papanicolaou DA, Wilder RL, Chrousos GP. Modulatory effects of glucocorticoids and catecholamines on human interleukin-12 and interleukin-10 productions. *Proc Assoc Am Phys* 1996; **108**:374-381.
- 30 Daynes RA, Araneo BA. Contrasting effects of glucocorticoids on the capacity of T cells to produce the growth factors IL-2 and IL-4. *Eur J Immunol* 1989; **19**:2319-2325.
- 31 Franchimont D, Galon J, Gadina M, Visconti R, Zhou YJ, Aringer M, Frucht, Chrousos GP, O'Shea JJ. Inhibition of Th1 immune response by glucocorticoids: Dexamethasone selectively inhibits IL-12-induced stat4 phosphorylation in T lymphocytes. *J Immunol* 2000; **164**:1768-1774.
- 32 Matalka KZ. Prolactin enhances the production of interferon- γ , interleukin-12, interleukin-10, but not of tumor necrosis factor- α , in a stimulus specific manner. *Cytokine* 2003; **21**:187-194.
- 33 Matalka KZ. The effect of estradiol, but not progesterone, on the production of cytokines in stimulated- whole blood is concentration-dependent. *Neuroendocrinol Lett* 2003; **24**:185-191.
- 34 Agarwal SK, Marshall GD. Glucocorticoid-induced type 1/type 2 cytokine alterations in humans: A model for stress-related immune dysfunction. *J Interferon and Cytokine Res* 1998; **18**:1059-1068.
- 35 Blotta MH, DeKruyff RH, Umetsu DT. Corticosteroids inhibit IL-12 production in human monocytes and enhance their capacity to induce IL-4 synthesis in CD4+ lymphocytes. *J Immunol* 1997; **158**:5589-5595.
- 36 DeKruyff RH, Fang Y, Umetsu DT. Corticosteroids enhance the capacity of macrophages to induce Th2 cytokine synthesis in CD4+ lymphocytes by inhibiting IL-12 production. *J Immunol* 1998; **160**:2231-2237.
- 37 Van der Poll T, Barber AE, Coyle SM, Lowery SF. Hypercortisolemia increases plasma interleukin-10 concentrations during human endotoxemia-a clinical research center study. *J Clin Endocrinol Metab* 1996; **81**:3604-3606.
- 38 Gayo A, Mozo L, Suarez A et al. Glucocorticoids increase interleukin-10 expression in multiple sclerosis patients with acute relapse. *J Neuroimmunol* 1998; **85**:122-130.
- 39 Dhabhar FS, McEwen, BS. Enhancing versus suppressive effects of stress hormones on skin immune function. *Proc Natl Acad Sci USA*. 1999; **96**:1059-1064.
- 40 Dhabhar FS, Satoskar AR., Bluethmann H, David JR, McEwen BS. Stress-induced enhancement of skin immune function: A role of γ interferon. *Proc Natl Acad Sci USA* 2000; **97**:2846-2851.
- 41 Sanders VM, Baker RA, RA, Ramer-Quinn DS, Kasproicz DJ, Fuchs BA, Street NE. Differential expression of the B2-adrenergic receptor by Th1 and Th2 clones. *J Immunol* 1997; **158**:4200-4210.
- 42 Agarwal SK, Marshall GD. Beta-adrenergic modulation of human type-1/type-2 cytokine balance. *J Allergy Clin Immunol* 2000; **105**:91-98.
- 43 Panina-Bordignon P, Mazzeo D, Di Lucia P, D'Ambrosio D, Lang R, Fabbri L, Self C, Sinigaglia F. β 2-agonists prevents Th1 development by selective inhibition of interleukin-12. *J Clin Invest* 1997; **100**:1513-1519.

- 44 Mohamed-Ali V, Flower L, Sethi J, Hotamisligil G, Gray R, Humphries SE, York DA, Pinkney J. Beta-adrenergic regulation of IL-6 release from adipose tissue: in vivo and in vitro studies. *J Clin Endocrinol Metab* 2001; **86**:5864–5869.
- 45 Woiciechowsky C, Asadullah K, Nestler D, Eberhardt B, Platzer C, Schoning B, Glockner F, Lanksch WR, Volk HD, Docke WD. Sympathetic activation triggers systemic interleukin-10 release in immunodepression induced by brain injury. *Nat Med* 1998; **4**: 808–813.
- 46 Maes M, Lin A, Kenis G, Egedy B, Bosmans E. Negative immunoregulatory effects of noradrenaline through alpha2-adrenoceptor activation. *Neuroendocrinol Lett* 2000; **21**:375–382.
- 47 Szelenyi J, Kiss JP, Puskas E, Selmeczy Z, Szelenyi M, Vizi ES. Opposite role of alpha2 and beta-adrenoceptors in the modulation of interleukin-10 production in endotoxemia mice. *Neuroreport* 2000; **11**:3565–3568.
- 48 Hartmann DP, Holaday JW, Bernton EW. Inhibition of lymphocyte proliferation by antibodies to prolactin. *FASEB J* 1989; **3**: 2194–2202.
- 49 Sabharwal P, Glaser R, Lafuse W, Varma S, Liu Q, Arkins S, Kojman R, Kutz L, Kelly KW, Malarkey WB. Prolactin synthesized and secreted by human peripheral blood mononuclear cells: an autocrine growth factor for lymphoproliferation. *Proc Natl Acad Sci USA*. 1992; **89**:77130–7716.
- 50 Dimattia GE, Gellersen B, Bohnet HG, Friesen HG. A human B lymphoblastoid cell line produces prolactin. *Endocrinology* 1988; **122**:2508–2517.
- 51 Montgomery DW, Shen GK, Ulrich ED, Steiner LL, Parrish PR, Zukoski CF. Human thymocytes express a prolactin-like messenger ribonucleic acid and synthesize bioactive prolactin-like proteins. *Endocrinology* 1992; **131**:3019–3026.
- 52 Bondy B, De Jonge S, Pander S, Primbs J, Ackenheil M. Identification of dopamine D4 receptor mRNA in circulating human lymphocytes using nested polymerase chain reaction. *J Neuroimmunol* 1996; **71**:139–144.
- 53 Santambrogio L, Lipartiti M, Bruni A, Daltoso R. Dopamine receptors on human T- and B-lymphocytes. *J Neuroimmunol* 1993; **45**:113–120.
- 54 Bernton EW, Meltzer MS, Holaday JW. Suppression of macrophage activation and lymphocyte function in Hypoprolactinemic mice. *Science* 1988; **239**:401–404.
- 55 Matera L, Contarini M, Bellone G, Forno B, Bigliano A. (1999). Up-modulation of interferon- γ mediates the enhancement of spontaneous cytotoxicity in prolactin-activated natural killer cells. *Immunology* 1999; **98**:386–392.
- 56 Vildaller A, Llorente L, Larrea F, Mendez JB, Alcocer-Varela J, Alarcon-Segovia. T cell-dysregulation in patients with hyperprolactinemia: effect of bromocriptine treatment. *Clin Immunol Immunopathol* 1986; **38**:337–343.
- 57 Meyerhoff JL, Oleshansky MA, Mougey EH. Psychologic stress increases plasma levels of prolactin, cortisol and POMC-derived peptides in man. *Psychosom Med* 1988; **50**:295–303.
- 58 Bierhaus A, Wolf J, Andrassy M, Rohleder N, Humpert PM, Petrov D, et al. A mechanism converting psychological stress into mononuclear cell activation. *Proc Natl Acad Sci USA* 2003; **100**: 1920–1925.
- 59 Maes M, Bockstaele DR, Van Gastel A, Song C, Schotte C, Neels H, DeMeester I, Scharpe S, Janca A. The effects of psychological stress on leukocyte subset distribution in humans: Evidence of immune reactivation. *Neuropsychobiology* 1999; **39**:1–9.
- 60 Guidi L, Tricerri A, Vangeli M, Frasca D, Errani AR, Di Giovanni A, Antico L, Menini E, Sciamanna V, Magnavita N, Doria G, Bartoloni C. Neuropeptide Y plasma levels and immunological changes during academic stress. *Neuropsychobiology* 1999; **40**:188–195.
- 61 Lacey K, Zaharia MD, Griffiths J, Ravindran AV, Merali Z, Anisman H. A prospective study of neuroendocrine and immune alterations associated with the stress of an oral academic examination among graduate students. *Psychoneuroendocrinology* 2000; **25**: 339–356.
- 62 Sauer J, Polack E, Wikinski S, Holsboer F, Stalla GK, Arzt E. The glucocorticoid sensitivity of lymphocytes changes according to the activity of the hypothalamic-pituitary-adrenocortical system. *Psychoneuroendocrinology* 1995; **20**:269–280.
- 63 Evans P, Bristow M, Hucklebridge F, Clow A, Pang FY. Stress, arousal, cortisol, and secretory immunoglobulin A in students undergoing assessment. *Br J Clin Psychol* 1994; **33**:575–576.
- 64 Collins A, Frankenhaeuser, M. Stress response in male and females engineering students. *J Human Stress* 1978; **4**:43–48.
- 65 Frankenhaeuser M, von Wright MR, Collins A, von Wright J, Sedvall G, Swahn CG. Sex differences in psychoneuroendocrine reactions to examination stress. *Psychosom Med* 1978; **40**: 334–343.
- 66 Semple CG, Gray CE, Borland W, Espie CA, Beastall GH. Endocrine effects of examination stress. *Clin Sci* 1988; **74**:255–259.
- 67 Malarkey WB, Pearl DK, Demers LM, Kiecolt-Glaser J, Glaser R. Influence of academic stress and season on 24-hour man concentrations of ACTH, cortisol, and β -endorphin. *Psychoneuroendocrinology* 1995; **20**:499–508.
- 68 Wood CE, Shinsako J, Keil LC, Dallman MF. (1982). Adrenal sensitivity to adrenocorticotropin in normovolemic and hypovolemic conscious dogs. *Endocrinology* 1982; **110**:1422–1429.
- 69 Bodner G, Ho A, Kreek MJ. Effect of endogenous cortisol levels on natural killer cell activity in healthy humans. *Brain Behav Immun* 1998; **12**:285–296.
- 70 Naliboff BD, Bente D, Solomon GF, Morley JE, Fahey JL, Bloom ET, Makinodan T, Gilmore SL. Immunological changes in young and old adults during brief laboratory stress. *Psychosom Med* 1991; **53**:121–132.
- 71 Pawlak CR, Jacobs, Mikeska E, Ochsmann S, Lombardi MS, Kavelaars A, Heijnen CJ, Schmidt R, Schedlowski M. Patients with systemic lupus erythematosus differ from healthy controls in their immunological response to acute psychological stress. *Brain Behav Immun* 1999; **13**:287–302.
- 72 van Doornen LJ, van Blokland, R. Serum-cholesterol: sex psychological correlates during rest and stress. *J Psychosom Res* 1987; **31**:239–249.
- 73 Danner SA, Endert E, Koster RW, Dunning A J. Biochemical and circulatory parameters during purely mental stress. *Acta Medica Scandinavica* 1981; **209**:35–308.
- 74 Malarkey WB, Hall JC, Pearl DK, Kiecolt-Glaser J, Glaser R. The influence of academic stress and season on 24-hour concentrations of growth hormone and prolactin. *J Clin Endocrinol Metab* 1991; **73**:1089–1092.
- 75 Freeman ME, Kanyicska B, Lerant A, Nagy G. Prolactin: structure, function, and regulation of secretion. *Physiol Rev* 2000; **80**: 1523–1631.
- 76 Maes M, Song C, Lin A, De Jongh R, Van Gastel A, Kenis G, Bosmans E, De Meester I, Benoy I, Neels H, Demedts P, Janca A, Scharpe S, Smith R. The effects of psychological stress on humans: Increased production of pro-inflammatory cytokines and a Th1-like response in stress-induced anxiety. *Cytokine* 1998; **10**: 313–318.
- 77 Paik IH, Toh KY, Lee C, Kim JJ, Lee SJ. Psychological stress may induce increased humoral and decreased cellular immunity. *Behav Med* 2000; **26**, 139–141.
- 78 Dobbins JP, Harth M, McCain GA, Martin RA, Cousin K. Cytokine production and lymphocyte transformation during stress. *Brain Behav Immun* 1991; **5**:339–348.
- 79 Uchakin PN, Tobin B, Cabbage M, Marshall Jr, Sams C. Immune responsiveness following academic stress in first-year medical students. *J Interferon Cytokine Res* 2001; **21**:687–694.
- 80 Kang DH, Fox C. Th1 and Th2 cytokine responses to academic stress. *Res Nurs Health* 2001; **24**:245–257.
- 81 Kang DH, Coe CL, McCarthy DO, Jarjour NN, Kelly EA, Rodriguez RR, Busse, WW. Cytokine profiles of stimulated blood lymphocytes in asthmatics and healthy adolescents across the school year. *J Interferon Cytokine Res* 1997; **17**:481–487.
- 82 Mills PJ, Berry CC, Dimesdale JE, Ziegler MG, Nelesen RA, Kennedy BP. Lymphocyte subset redistribution in response to acute experimental stress: effects of gender, ethnicity, hypertension, and the sympathetic nervous system. *Brain Behav Immun* 1995; **9**:61–69.
- 83 Mills PJ, Dimesdale JE, Nelesen RA, Dillon E. Psychologic characteristics associated with acute stressor-induced leukocyte subset redistribution. *Psychosom Res* 1996; **40**:417–423.

- 84 Bachen EA, Manuck SB, Marsland AL, Cohen S, Malkoff SB, Muldoon M, Rabin BS. Lymphocyte subset and cellular immune responses to a brief experiment stressor. *Psychosom Med* 1992; **54**:673-679.
- 85 Brosschot JF, Benschop RJ, Godaert GL, de Smet MB, Olff M, Heijnen CJ, Ballieux RE. Effects of experimental psychological stress on distribution and function peripheral blood cells. *Psychosom Med* 1992; **54**:394-406.
- 86 McClelland DC, Ross G, Patel V. The effect of an academic examination on salivary norepinephrine and immunological levels. *J Human Stress* 1985; **11**, 52-59.
- 87 Bosch JA, de Geus EJ, Kelder A, Veerman EC, Hoogstraten J, Amerongen AV. Differential effects of active versus passive coping on secretory immunity. *Psychophysiology* 2001; **38**:836-846.
- 88 Dhabhar FS. Stress-induced augmentation of immune function-The role of stress hormones, leukocytes trafficking, and cytokines. *Brain Behav Immun* 2002; **16**:785-798.
- 89 Groul DJ, Campbell NF, Bourgeois S. (1986). Cyclic AMP-dependent protein kinase promotes glucocorticoid receptor function. *J Biol Chem* 1986; **261**:4909-4914.
- 90 DiBattista JA, Martel-Pelletier J, Cloutier JM, Pelletier JP. Modulation of glucocorticoid receptor expression in human articular chondrocytes by cAMP and prostaglandins. *J Rheumat Suppl* 1991; **27**: 102-105.
- 91 Korn SH, Wouters EFM, Wesseling G, Arends JW, Thunnissen FBJM. Interaction between glucocorticoids and beta-agonists: alpha and beta glucocorticoid-receptor mRNA expression in human bronchial epithelial cells. *Biochem Pharmacol* 1998; **56**: 1561-1569.
- 92 Nakada MT, Stadel JM, Poksay KS, Crooke ST. (1987). Glucocorticoid regulation of beta-adrenergic receptors in 3T3-L1 preadipocytes. *Mol Pharmacol* 1987; **31**:377-384.
- 93 Cohen S, Tyrrell DA, Smith AP. Psychological stress and susceptibility to the common cold. *N Eng J Med* 1991; **325**, 606-612.
- 94 Kang DH, Coe CL, McCarthy DO. Academic examinations significantly impact immune responses, but not lung function, in healthy and well managed asthmatic adolescents. *Brain Behav Immun* 1996; **10**:164-181.
- 95 Jemmott JB 3rd, Borysenko JZ, Borysenko M, McClelland DC, Chapman R, Meyer D, Benson H. Academic stress, power motivation, and decrease in secretion rate of salivary secretory immunoglobulin A. *Lancet* 1983; **8339**:1400-1402.
- 96 Jemmott JB 3rd, Magloire K. Academic stress, social support, and secretory immunoglobulin A. *J Pers Soc Psychol* 1988; **55**: 803-810.
- 97 Deinzer R, Kleineidam C, Stiller-Winkler R, Idel H, Bachg, D. Prolonged reduction of salivary immunoglobulin A (sIgA) after a major academic exam. *Int J Psychophysiol* 2000; **37**:219-232.
- 98 Deinzer R, Schuller N. Dynamics of stress-related decrease of salivary immunoglobulin A (s-IgA): relationship to symptoms of the common cold and studying behavior. *Behav Med* 1988; **23**: 161-169.
- 99 Lowe G, Urquhart J, Greenman J, Lowe G. Academic stress and secretory immunoglobulin A. *Psychology Report* 2000; **87**: 721-722.
- 100 Glaser R, Friedman S, Smyth J, Ader R, Bijur P, Brunell P, Cohen N, Krilov L, Lifrak S, Stone A. The differential impact of training stress and final examination stress on herpesvirus latency at the United States military academy at West Point. *Brain Behav Immun* 1999; **13**:240-251.
- 101 Sarid O, Anson O, Yaari A, Margalith M. Epstein-Barr virus specific salivary antibodies as related to stress caused by examinations. *J Med Virol* 2001; **64**:149-156.
- 102 Joncas J, Boucher J, Boudreault A, Granger-Julien M. Effect of hydrocortisone on cell viability, Epstein-Barr virus genomic expression, and interferon synthesis in human lymphoblastoid cell lines. *Cancer Res* 1973; **33**:2142-2148.
- 103 Tanaka J, Ogura T, Kamiya S, Sato H, Yoshie T, Ogura H, Hatano M. Enhanced replication of human cytomegalovirus in human fibroblasts treated with dexamethasone. *J Gen Virol* 1984; **65**: 1759-1767.
- 104 Tanaka J, Ogura T, Kamiya S, Yoshie T, Yabuki Y, Hatano M. (1984b). Dexamethasone enhances human cytomegalovirus replication in human epithelial cell cultures. *Virology* 1984; **136**:448-452.
- 105 Prachova K, Roubal J. Effects of hydrocortisone on the synthesis of Epstein-Barr virus antigens in P3HR-1 cells. *Acta Virol* 1981; **25**:163-166.
- 106 Lathey JL, Spector SA. Unrestricted replication of human cytomegalovirus in hydrocortisone - treated macrophages. *J Virol* 1991; **65**:6371-6375.
- 107 Prosch S, Wendt CE, Reinke P, Priemer C, Oppert M, Kruger DH, Volk HD, Docke WD. A novel link between stress and human cytomegalovirus (HCMV) infection: Sympathetic hyperactivity stimulates HCMV activation. *Virology* 2000; **272**:357-365.
- 108 Dobbs CM, Vasquez M, Glaser R, Sheridan JF. Mechanisms of stress-induced modulation of viral pathogenesis and immunity. *J Neuroimmunol* 1993; **48**:152-260.
- 109 Liu LY, Coe CL, Swenson CA, Kelly EA, Kita H, Busse WW. School examinations enhance inflammation to antigen challenge. *Am J Respir Crit Care Med* 2002; **165**:1062-1067.
- 110 Glaser R, MacCallum RC, Laskowski BF, Malarkey WB, Sheridan JF, Kiecolt-Glaser JK. Evidence for shift in Th-1 to Th-2 cytokine response associated with chronic stress and aging. *J Geront: Med Sci* 2001; **56A**:M477-M482.
- 111 Miller GE, Cohen S, Ritchey AK. Chronic psychological stress and the regulation of proinflammatory cytokines: a glucocorticoid-resistance model. *Health Psychol* 2002; **21**:536-541.