Effects of different antioxidants on lipid peroxidation in brain homogenates induced by thyrotoxicosis in rats

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Abstract **OBJECTIVE:** It is known that thyrotoxicosis induces lipid peroxidation (LPO). In contrast, propylthiouracil (PTU), a thyrostatic drug is a well-known antioxidant. Also melatonin has been shown to protect against free radical-induced neuronal destruction. At the same time, it is generally accepted that the brain is the most vulnerable tissue to oxidative stress.

METHODS: The goal of the study was to examine the components of LPO, i.e., conjugated dienes (CD), malondialdehyde (MDA) and Schiff's bases (SB), in the brain of male Wistar rats. Two experiments were performed, with two control groups created for each experiment: Group 1 - intact animals and Group 2 - animals injected with 0.9% NaCl. In Experiment I, the animals received L-thyroxine $(L-T_4)$ in a dose of 100 μ g/kg BW, i.p., daily, for two weeks (Groups 3–5). After one week of L-T₄ treatment, the following agents were added during a subsequent week: Group 4 – PTU in drinking water (45 mg/kg BW/day); Group 5 - melatonin (5 mg/kg BW, daily). In Experiment II, lasting 7 days, the animals were divided into the following groups: Group 1 – intact animals; Group 2 – animals injected with 0.9% NaCl; Group 3 – PTU in drinking water (45 mg/kg BW/day); Group 4 - melatonin (5 mg/kg BW, daily). **RESULTS:** In Experiment I, we observed a significantly higher SB level in saline treated animals and a significant increase in both CD and SB levels in rats treated with L-T₄, compared to those in non treated control. CD levels were also elevated in rats treated with L-T₄, compared to values in the saline only treated animals. Melatonin and PTU reduced CD levels and melatonin diminished SB levels, as com-

pared to those in L-T₄-treated rats. In Experiment II, we observed significantly higher CD, SB and MDA levels in saline treated rats, when compared to respective values in non treated control. Melatonin decreased CD levels, when compared to CD levels in both the non treated and saline injected controls. Additionally, melatonin reduced SB levels relative to change in the brains of saline treated rats. Furthermore, PTU decreased CD levels in brain homogenates compared to non-treated animals. **CONCLUSIONS:** (1) Thyrotoxicosis stimulates LPO in the rat brain; (2) All the examined antioxidants decrease LPO in L-T₄-administered animals; (3) All the examined antioxidants reduce the basal LPO; 4) Stress, when induced by handling, intensifies oxidative processes in the organism.

Introduction

The central nervous system is highly susceptible to damage by oxidative stress induced by a variety of biological agents [1, 2]. Because of its high metabolic activity, the brain utilizes large amounts of molecular oxygen, followed by high free radical generation; it is also contains very large amounts of polyunsaturated fatty acids which are easily oxidized [2]. Cerebrospinal fluid is highly enriched with iron and ascorbic acid which can generate the highly toxic hydroxyl radical (•OH) [1]. On the other hand, the brain is poorly equipped with an endogenous antioxidative defense system [3]. This organ exhibits a higher degree of tissue peroxidation than does either the liver or the lung [4].

It is well established that thyroid hormones accelerate the basal metabolic rate and, particularly, oxidative metabolism, as evidenced by the induction of certain mitochondrial enzymes in target tissues. Thyroid hormone administration is associated with a higher electron flow through the mitochondrial electron-transport systems, as indicated by an enhanced production of superoxide radicals ($O_2^{\bullet-}$). In the course of thyrotoxicosis, the oxidative stress and lipid peroxidation (LPO) are generated, which are not compensated by changes in the endogenous antioxidative system [5].

Propylthiouracil (PTU) is widely used in the therapy of hyperthyroidism. There is also some evidence that thyrostatic agents *per se* may reduce the oxidative stress [6]. It has been shown that PTU can directly scavenge hydrogen peroxide (H2O2) and •OH; but it is, however, less reactive towards $O_2^{\bullet-}$. Moreover, it protects lipids from peroxidation [6]. PTU has been demonstrated to reduce LPO in patients with hyperthyroidism [5, 7].

Melatonin, the main secretory product of the pineal gland, is an efficient free radical scavenger and antioxidant. It directly neutralizes •OH, O_2 •⁻, hypochlorus acid, nitric oxide, peroxynitrite anion, H_2O_2 , and singlet oxygen (8–10). The efficacy of melatonin in scavenging the peroxyl radical is controversial [11]. It has well been documented that melatonin is capable of inhibiting LPO [12,13]. Also, melatonin has been reported to stimulate activities of several antioxidative enzymes, for example, superoxide dismutase (SOD), glutathione peroxidase (GSH-Px), glutathione reductase (GSH-Rd); also it is known to inhibit nitric oxide synthase, a prooxidative enzyme [14–16]. At the same time, melatonin can synergistically interact with other antioxidants [17]. Additionally, this indoleamine stabilizes cell membranes [18].

The goals of this study were :

- to evaluate the levels of the three LPO parameters, i.e, conjugated dienes (CD), malondialdehyde (MDA) and Schiff's bases (SB) in the brain of male Wistar rats treated with L-thyroxine (L-T₄);
- 2) to examine the effectiveness of both PTU and melatonin in inhibiting oxidative stress;
- 3) to evaluate the action of antioxidants on basal LPO levels.

Materials and Methods

In the study, 70 male Wistar rats, weighing 150–175 g each, were used. The animals were fed with standard diet, having a free access to food and water. The rats were kept in controlled lighting conditions (12 h light; 12 h darkness).

Two experiments were performed (Fig. 1). The following control groups were created in each experiment: Group 1 – intact animals and Group 2 – animals injected with 0.9% NaCl. In Experiment I, the animals were intraperitoneally (i.p.) injected with L-thyroxine $(L-T_4)$ in a dose of 100 µg/kg BW, daily for two weeks (Groups 3-5). After one week of $L-T_4$ treatment, the rats received the following substances for subsequent 7 days: Group 4 - PTU in drinking water (45 mg/kg BW/day); Group 5 - melatonin (5 mg/kg BW, i.p.; at 16.00 p.m.). In Experiment II, which lasted 7 days, the animals were divided into the following groups: Groups 1 - intact animals, Group 2 – animals injected with 0.9% NaCl, Group 3 – PTU in drinking water (45 mg/kg BW/day), Group 4 – melatonin (5 mg/kg BW, daily). At the end of the study, the animals were decapitated, the brains were removed and stored at -80°C.

In the present study, the following products of LPO were examined in the homogenates of the brain: MDA concentrations [19], the levels of CD and SB [20]. The results are expressed as MDA, CD, or SB per gram of wet tissue. A one-way analysis of variance (ANOVA) and the Newman-Keuls' test were applied to evaluate statistical significance. All the calculations were performed with the use of the Statistica `99 computer software.

Results

In Experiment I, we observed a significantly higher SB levels in saline treated animals, compared to those in non treated controls (Fig. 2). Moreover, we showed significantly higher CD (Fig. 3) and SB (Fig. 2) levels in rats treated with L-T₄, compared to those in non treated controls; CD level was also elevated, compared to values in saline injected animals (Fig. 3). Melatonin and PTU reduced CD levels (Fig. 3) and melatonin diminished SB levels (Fig. 2), as compared to those in L-T₄-treated rats.

In Experiment II, we noticed significantly higher CD (Fig. 4) and SB levels (Fig. 5) and MDA concentrations (Fig. 6) in saline treated rats, when compared to respective values in non treated animals. Melatonin decreased CD levels, when compared to CD levels in both the saline trated animals and non treated controls (Fig. 4). Melatonin reduced also SB levels, when compared to those in saline injected controls (Fig. 5). On the other side, concentrations of MDA after melatonin were higher, when compared to those in untreated controls (Fig. 6). Propylthiouracil decreased CD levels in brain homogenates compared to those in respective values in non treated animals (Fig. 4).

Discussion

In the reported study, we have established that thyrotoxicosis induces LPO in the brain, what is consistent with other previously published data. Chehade et al. [21] demonstrated an increased MDA concentration in the brain of rats injected with triiodothyronine (T_3), while no changes in SB level were noticed. Many authors



Figure 1. Scheme of Experiment I and Experiment II: L-thyroxin (L-T₄); Melatonin (MEL); propylothiouracil (PTU).

emphasize excessive LPO in other organs of thyrotoxic rats after T_3 administration [22]. In another investigation, thyrotoxicosis, induced by L- T_4 , was accompanied by excessive LPO [23]. Moreover, numerous publications have reported an intensification of oxidative metabolism and, as a consequence, an acceleration of LPO in hyper-



Figure 2. Levels of Schiff's bases (SB) in the brains of rats in the five treated groups. Data represent means ± SEM. Level of significance:

- vs. untreated controls, p<0,005;</p>

- vs. untreated controls, p<0,05;</p>
 - vs. L-T₄-treated group, p<0,05;</p>



Figure 3. Levels of conjugated dienes (CD) in the brains of rats in the five treated groups.





Figure 4. Levels of conjugated dienes (CD) in the brains of rats in the four treated groups.

Data represent means \pm SEM. Level of significance: - vs. untreated controls, p<0,05; vs. coline treated group, p<0,0005;

vs. saline treated group, p<0,0005;</p>

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Figure 5. Levels of Schiff's bases (SB) in the brains of rats in the four treated groups.

Data represent means ± SEM. Level of significance: Data represent means ± SEM. Level of significance: P<0,0005; P<0,0005; P<0,0005;

thyroid men and their normalization after therapy [5, 7, 24]. Also, other authors have shown increased LPO both, in hyper- and hypothyroidism [25].

The effects of PTU on oxidative processes in the brain during thyrotoxicosis have not been carefully examined to date. We observed in the study that PTU reduced CD levels, both in hyperthyroid rats and under basal conditions. However, it has been reported in numerous publications that PTU therapy inhibits LPO and normalizes the activity of antioxidative enzymes in blood of hyperthyroid patients [5, 7]. At the same time, it has not yet been possible to ascertain whether PTU directly scavenges free radicals [6] or whether its effect is a consequence of euthyreoidism restoration [5]. Recent data from animal studies have demonstrated that PTU protects from acetaminophen and thioacetamide hepatotoxicity in rats; this thyrostatic drug loweres MDA concentration, as well as diminishes liver necrosis and inflammation [26,27]. The authors believe that the advantageous action of PTU is exerted at the metabolic level, depending on induced hypothyroid state.

Recently, melatonin has been shown to be highly effective in abating oxidative damage in the central nervous system [2, 28]. It has been reported that this indolamine easily crosses through the blood-brain barrier and is rapidly taken up by the brain. Skinner and Malpaux [29] have recently found that melatonin concentrations in the cerebro-spinal fluid of the third ventricle is 20-fold higher than its nocturnal plasma levels. In our study, we observed that melatonin reduced LPO in rats injected with L-T₄. There has been no other published work, concerning the role of melatonin in oxidative processes in the course of thyrotoxicosis but numerous authors have described a neuroprotective action of this indoloamine.



Figure 6. Concentrations of malondialdehyde (MDA) in the brains of rats in the four treated groups.

Data represent means ± SEM. Level of significance:

- vs. untreated controls, p<0,05;</p>

- vs. saline treated group, p<0,05</p>

Experiments under both in vivo [30] and in vitro [31] conditions have shown that melatonin effectively inhibits LPO in the brain, induced by kainic acid, a neuroexitotogic agent. In our experiments, melatonin reduced the basal LPO in the brain. The results are in agreement with those published in other papers, showing that melatonin diminishes MDA and 4-hydroxyalkenals (4-HDA) in the brain during ischemia-reperfusion, another high oxidative stress condition [32, 33]. Additionally, many authors have noticed that this indoleamine limits the basal LPO in rat [15, 31] and mouse brains [34]. Accordingly, Tan et al. [35] have found that melatonin reduces the autooxidation of lipids in homogenates of monkey brain cortex.

Unexpectedly, in the present study, a difference in LPO was noted between the two control groups, i.e., untreated and saline injected. Our results suggest that the stress, associated with handling and injections, intensifies oxidative processes in the organism and, as a consequence, exaggerates LPO. In numerous publications, the authors have reported that surgical stress intensifies free radical production [36]. Well-known stressful procedures, such as immobilization or excessive physical exercise, may also result in LPO acceleration [37].

In summary, our results demonstrate that thyrotoxicosis, as well as stress, induced by handling, stimulate LPO in rat brains and that the examined antioxidants (PTU, melatonin) exert protective effects, i.e., decrease LPO in the rat brain under both basal conditions and in animals treated with L-T₄. These results are consistent with a large number of studies, documenting the ability of melatonin to limit peroxidation of lipids under various conditions [38–40].

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REFERENCES

- 1 Halliwell B. Reactive oxygen species and the central nervous system. J Neurochem 1992; **59**:1609–1623.
- 2 Reiter RJ, Tan D-X, Qi W, Manchester LC, Karbownik M, Calvo JR. Pharmacology and physiology of melatonin in the reduction of oxidative stress in vivo. Biol Signals Recept 2000; 9:160–171.
- 3 Perez-Campo R, Lòpez-Torres M, Rojas C, Cadenas S, Baria G. A comparative study of free radicals in vertebrates I. Antioxidant enzymes. Comp Biochem Physiol 1993; **105B**: 749–755.
- 4 Lòpez-Torres M, Pérez-Campo R, Cadenas S, Rojas C, Baria G. A comparative study of free radicals in vertebrates II. Non-enzymatic antioxidants and oxidative stress. Comp Biochem Physiol 1993; **105B**: 757–763.
- 5 Bianchi G, Solaroli E, Zaccheroni V, Grossi G, Bargossi AM, Melchionda N, Marchesini. Oxidative stress and anti-oxidant metabolites in patient with hyperthyroidism: effect of treatment. Horm Metab Res 1999; **31**: 620–624.
- 6 Hicks M, Wong LS, Day RO. Antioxidant activity of propylthiouracil. Biochem Pharmacol 1992; **43**: 439–444.
- 7 Seven A, Tasan E, Hatemi H, Burcak G. The impact of propylthiouracil therapy on lipid peroxidation and antioxidant status parameters in hyperthyroid patients. Acta Med Okayama 1999; **53**: 27–30.
- 8 Tan D-X, Manchester LC, Reiter RJ, Plummer BF, Hardies LJ, Winteraub ST, et al. A novel melatonin metabolite, cyclic 3-hydroxymelatonin]: a biomarker of in vivo hydroxyl radical generation. Biochem Biophys Res Commun 1998; **253**:614–620.
- 9 Allegra M, Reiter RJ, Tan DX, Gentile C, Tesoriere L, Livrea MA. The chemistry of melatonin's interaction with reactive species. J Pineal Res 2003; **34**:1–10.
- 10 Tan D-X, Manchester LC, Reiter RJ, Qi W, Karbownik M, Calvo JR. Significance of melatonin in antioxidative defense system: reactions and products. Biol Signal Recept 2000; 9: 137–159.
- 11 Antunes F, Barclay RC, Ingold KU, King M, Norris JQ, Scaiano JC, et al. On the antioxidant activity of melatonin. Free Radic Biol Med 1999; 26:117–128.
- 12 Sewerynek E, Melchiorri D, Chen L-D, Reiter RJ. Melatonin reduces both basal and bacterial lipopolysaccharide-induced lipid peroxidation in vitro. Free Rad Biol Med 1995; **19**: 903–909.
- 13 Sewerynek E, Reiter RJ, Melchiorri D, Ortiz GG, Lewinski A. Oxidative damage in the liver induced by ischemia-reperfusion: Protection by melatonin. Hepato-Gastroenterology 1996; 43:898– 905.
- 14 Reiter RJ, Tan DX, Osuna C, Gitto E. Actions of melatonin in the reduction of oxidative stress. J Biochem Sci 2000; **7**:444–458.
- 15 Pablos MI, Reiter RJ, Ortiz GG, Guerrero JM, Agapito MT, Chuang JI, et al. Rhythms of glutathione peroxidase and glutathione reductase in brain of chick and their inhibition by light. Neurochem Int 1998; **32**:69–75.
- 16 Pozo D, Reiter RJ, Calvo JP, Guerrero JM. Physiological concentrations of melatonin inhibits nitric oxide synthase in rat cerebellum. Life Sci 1994; 55:PL-455-PL-460.
- 17 Poeggeler B, Reiter RJ, Hardeland R, Sewerynek E, Melchiorri D, Barlow-Walden LR. Melatonin, a mediator of electron transfer and repair reactions, acts synergistically with the chain breaking antioxidants ascorbate, troxol and glutathione. Neuroendocrinol Lett 1995; **17**:87–92.
- 18 Garcia JJ, Reiter RJ, Guerrero JM, Escames G, Yu BP, Oh CS, et al. Melatonin prevents changes in microsomal membrane fluidity during induced lipid peroxidation. FEBS Lett 1997; 408:297–300.
- 19 Rice-Evans CA, Diplock AT,. Symons MCR. Techniques in Free Radical Research. Elsevier, Amsterdam. 1991.
- 20 Buege JA, Aust SD. Microsomal lipid peroxidation. Methods Enzymol 1978; **52**:302–310.
- 21 Chehade J, Kim J, Pinnas JL, Mooradian AD. Age-related changes in the thyroid hormone effects on malondialdehyde-modified proteins in the rat heart. Proc Soc Exp Biol Med 1999; 222:59–64.

- 22 Venditti P, Balestrieri M, Di-Meo S, De-Leo T. Effect of thyroid state on lipid peroxidation, antioxidant defenses, and susceptibility to oxidative stress in rat tissues. J Endocrinol 1997; **155**: 151–157.
- 23 Shinohara R, Mano T, Nagasaka A, Hayashi R, Uchimura K, Nakano I, et al. Lipid peroxidation levels in rat cardiac muscle are affected by age and thyroid status. J Endocrinol 2000; **164**:97– 102.
- 24 Sewerynek E, Wiktorska JA, Nowak D, Lewinski A. Methimazole protection against oxidative stress induced by hyperthyroidism in Graves' disease. Endocrine Regul 2000; 34:82–89.
- 25 Costantini F, Pierdomenico SD, De-Cesare D, De-Remigis P, Bucciarelli T, Bittolo-Bon G, et al. Effect of thyroid function on LDL oxidation. Arterioscler Thromb Vasc Biol 1998; 18:732–737.
- 26 Bruck R, Frenkel D, Shirin H, Aeed H, Matas Z, Papa M, et al. Hypothyroidism protect rat liver from acetaminophen hepatotoxicity. Dig Dis Sci 1999; **44**:1228–1235.
- 27 Bruck R, Oren R, Shirin H, Aeed H, Papa M, Matas Z, et al. Hypothyroidism minimalizes liver damage and improves survival in rats with thioacetamide induced fulminant hepatic failure. Hepatology 1998; 27:1013–1020.
- 28 Reiter RJ, Cabrera JC, Sainz RM, Mayo JC, Manchester LC, Tan D-X. Melatonin as a pharmacological agent against neuronal loss in experimental models of Huntington's disease, Alzheimer's disease and Parkinsonism. Ann N Y Acad Sci 1999; 890:471–484.
- 29 Skinner DC, Malpaux B. High melatonin concentration in third ventricular cerebrospinal fluid are not due to Galen vein blood recirculating through the choroid plexus. Endocrinology 1999; **140**: 4399–4405.
- 30 Tan D-X, Manchester LC, Reiter RJ, Qi W, Kim SJ, El-Sokkary GH. Melatonin protects hippocampal neurons in vivo against kainic acid-induced damage in mice. J Neurosci Res 1998; 54:382–389.
- 31 Melchiorri D, Reiter RJ, Sewerynek E, Chen L-D, Nistico G. Melatonin reduces kainate-induced lipid peroxidation in homogenates of different brain regions. FASEB J 1995; 9:1205–1210.
- 32 Li X-J, Zhang L-M, Gu J, Zhang A-Z, Sun F-Y. Melatonin decreases production of hydroxyl radical during cerebral ischemiareperfusion. Acta Pharmacol Sinica 1997; 18:393–396.
- 33 Cheung RT. The utility of melatonin in reducing cerebral damage resulting from ischemia and reperfusion. J Pineal Res 2003; 34:153–60.
- 34 Yamamoto H, Tang HW. Effect of melatonin, piperonyl butoxide, or cobalt chloride on L-cysteine-induced lipid peroxidation in homogenate from whole brain of mice. Toxicol Lett 1996; **89**: 51–56.
- 35 Tan D-X, Manchester LC, Reiter RJ, Cabrera J, Burkhardt S, Phillip T, et al. Melatonin suppresses autooxidation and hydrogen peroxide-induced lipid peroxidation in monkey brain homogenate. Neuroendocrinol Lett 2000; 21:361–365.
- 36 Anup R, Aparna V, Pulimood A, Balasubramanian KA. Surgical stress and the small intestine: role of oxygen free radicals. Surgery 1999; **125**:560–569.
- 37 Hara M, ligo M, Ohtani-Kaneko R, Nakamura N, Suzuki T, Reiter RJ, et al. Administration of melatonin and related indoles prevents exercise-induced cellular oxidative changes in rats. Biol Signals 1997; 6:90–100.
- 38 Ohta Y, Kongo M, Kishikawa T. Preventive effect of melatonin on the progression of alpha-naphthylisothiocyanate-induced acute liver injury in rats. J Pineal Res 2003; 34:185–93.
- 39 Sener G, Sehirli AO, Paskaloglu K, Dulger GA, Alican I. Melatonin treatment protects against ischemia/reperfusion-induced functional and biochemical changes in rat urinary bladder. J Pineal Res 2003; 34:226–30.
- 40 Esparza JL, Gomez M, Romeu M, Mulero M, Sanchez DJ, Mallol J, et al. Aluminum-induced pro-oxidant effects in rats: protective role of exogenous melatonin. J Pineal Res 2003; **35**:32–9.