# Luzindole but not 4-phenyl-2propionamidotetralin (4P-PDOT) diminishes the inhibitory effect of melatonin on murine Colon 38 cancer growth *in vitro*

#### Katarzyna WINCZYK<sup>1</sup>, Julita FUSS-CHMIELEWSKA<sup>1</sup>, Hanna LAWNICKA<sup>2</sup>, Marek PAWLIKOWSKI<sup>1</sup> and Michal KARASEK<sup>1</sup>

Department of Neuroendocrinology, Chair of Endocrinology, Medical University of Lodz, Poland.
Department of Immunoendocrinology, Chair of Endocrinology, Medical University of Lodz, Poland.

Correspondence to:	orrespondence to: Katarzyna Winczyk, MD., PhD. Department of Neuroendocrinology, Chair of Endocrinology, Medical University of Lodz, 3 Sterling Street, 91-25 Lodz, Poland. PHONE/FAX: +48426365427; E-MAIL: katarzyna.winczyk@umed.lodz.pl	
Submitted: 2009-02-2	Accepted: 2009-03-14	Published online: 2009-11-11
Key words:melatonin; melatonin receptor; luzindole; 4P-PDOT; proliferation; colon cancer		

Neuroendocrinol Lett 2009; 30(5):657-662 PMID: 20035258 NEL300509A04 © 2009 Neuroendocrinology Letters • www.nel.edu

Abstract **OBJECTIVE:** Our earlier studies have shown that MLT exerts the inhibitory effect on murine cancer via membrane and nuclear receptors. We have found that the antagonist of MT<sub>1</sub> receptors does not diminish the antiproliferative effect of MLT on Colon 38 cells, and the contribution of MT<sub>2</sub> receptors has been suggested to be responsible. Therefore, in the present study we have examined the influence of the 4-phenyl-2-propionamidotetralin (4P-PDOT), which is a selective antagonist of MT<sub>2</sub> membrane receptor, and luzindole – an antagonist of both membrane receptors, on an oncostatic action of MLT. MATERIALS AND METHODS: The murine cancer cell line Colon 38 was used in the experiments. In 48 hrs cell culture the effects of MLT, 4P-PDOT and luzindole administered alone and MLT applied jointly with either 4P-PDOT or luzindole were examined. The growth of cancer cells was assessed using the modified colorimetric Mosmann method. **RESULTS:** Melatonin at both examined concentrations (10<sup>-7</sup>, 10<sup>-9</sup> M) significantly decreased the viability of cancer cells. The selective antagonist of MT2 membrane receptor, namely 4P-PDOT and luzindole applied separately did not have an effect on the growth of Colon 38 cells. The addition of 4P-PDOT to MLT did not change the inhibitory effect of MLT, whereas luzindole given together with MLT diminished, but failed to block totally, the oncostatic properties of MLT. **CONCLUSIONS:** The obtained data and our previous studies conducted on Colon 38 cancer indicate that membrane melatonin receptors are not indispensable to the oncostatic action of melatonin and thus other pathways such as nuclear signaling and receptor-independent mechanism may be also involved.

## INTRODUCTION

The influence of melatonin (MLT) on the growth of various experimental cancers, including colon cancer, has been investigated intensively over recent years (Pawlikowski et al. 2002). It has been shown that MLT decreases the incidence, number and size of 1,2-dimethylhydrazine - induced colon tumors in rodents by suppressing tumor growth and invasiveness (Anisimov et al. 1997; 2000). The inhibitory effect of MLT on animal and human colon cancer cells in culture has been also reported (Farriol et al. 2000; García-Navarro et al. 2007). The majority of the investigations were related to the antiproliferative action of melatonin. Our previous study conducted on murine Colon 38 cancer showed that MLT inhibits cell proliferation in vivo and in vitro conditions (Melen-Mucha et al. 1998; Winczyk et al. 2002). Melatonin exerts the oncostatic effect also via induction of apoptosis. We have found that MLT enhanced the cells apoptosis in murine colonic cancer (Melen-Mucha et al. 1998; Winczyk et al. 2001). The proapoptotic effect of the pineal hormone was confirmed by other study (García-Santos et al. 2006; Martín-Renedo et al. 2008). Although data supporting the inhibitory action of MLT on the growth of human cancer is documented well enough in the literature, the molecular and cellular mechanisms by which this hormone can exert the oncostatic effect still remain unclear. Melatonin acts via the modulation of endocrine and immune systems and the said hormone may also directly influence cancer cells through specific binding sites. The best characterized binding sites of MLT are G protein-coupled membrane receptors, named MT<sub>1</sub> and MT<sub>2</sub> (Dubocovich et al. 1998). In mammals membrane receptors participate in the regulation of circadian and seasonal rhythms. The binding sites of melatonin were found also in gastrointenstinal tract, where this hormone plays an important role (Lee and Pang, 1993; Bubenik, 2008). As a small lipophilic molecule, melatonin easily crosses cellular membranes and may exert its biological effects through cytoplasmic and nuclear signaling. The nuclear orphan receptors named RZR/ROR were proposed as a nuclear binding sites for melatonin (Calberg and Wiesenberg, 1995; Wiesenber, 1998). The oncostatic effects of the pineal hormone seem to depend on membrane and nuclear receptors. It has been shown that melatonin-induced suppression of hepatoma growth is mediated via G-protein connected membrane receptors (Blask et al. 1999). The involvement of membrane receptors in antiproliferative effects of melatonin have been described in the following cell lines of human carcinoma: Jar and JEG-3 choriocarcinoma, MCF-7 breast cancer, Ishikawa endometrial cancer, LNcaP and Du-145 prostate cancers (Shiu et al. 1999; 2000; Ram et al. 2002; Kanishi et al. 2000; Xi et al. 2001; Marelli et al. 2000). The possibility of melatonin action via nuclear biding sites were examined in the human breast cancer, androgen-dependent and androgen-independent prostate cancers, the cells of which possess the RZR/ROR receptors (Dai et al. 2001; Moretti et al. 2000; 2001a). We have documented that both MT<sub>1</sub> and MT<sub>2</sub> membrane receptors as well as RZR/ RORa receptors are expressed in Colon 38 cancer cells (Karasek et al. 2002). Our earlier study showed that melatonin exerts an inhibitory effect on murine cancer via membrane and nuclear receptors (Winczyk et al. 2002; Karasek et al. 1998). Moreover, we have found that antagonist of MT<sub>1</sub> receptors does not diminish the antiproliferative effect of the pineal hormone on Colon 38 cells, and the contribution of MT<sub>2</sub> receptors has been suggested. Therefore, in the present study we examined the influence of the 4-phenyl-2-propionamidotetralin (4P-PDOT) – a selective antagonist of  $MT_2$  membrane receptor - as well as luzindole - an antagonist of MT<sub>1</sub> and MT<sub>2</sub> membrane receptors – on an oncostatic action of melatonin.

### MATERIALS AND METHODS

#### <u>Compounds</u>

The following substances were examined in this study: melatonin (N-acetylo-5-metoksytryptamina, Sigma), 4-phenylo-2-propionamidotetralin (4-P-PDOT, Tocris Bioscience) – a selective antagonist of  $MT_2$  membrane receptor and N-acetylo-2-benzylotryptamin (Luzindole, Tocris Bioscience) – the non-selective antagonist of  $MT_1$  and  $MT_2$  membrane receptors. Melatonin and 4P-PDOT were dissolved in 95% ethanol and DMSO was used as dissolvent for luzindole.

### <u>Cell culture</u>

The murine cancer cell line Colon 38, kindly obtained from Ludwik Hirszfeld Institute of Immunology and Experimental Therapy, Polish Academy of Sciences in Wroclaw, was used in the experiment. The Colon 38 is transplantable adenocarcinoma originally induced in the colon of C57BL/6 strain mouse by 1,2-dimethylyhydrazine (Corbett *et al.* 1975). Adaptation of Colon 38 cells to *in vitro* growth was made by Pajtasz-Piasecka and co-workers (2004).

The continuous culture of the cells was maintained in culture flasks (Nunc Easy flask 25 cm<sup>2</sup>, NUNC). The cells were cultured in the present of RPMI 1640 medium (Sigma), supplemented with 25 mM Hepes buffer (Sigma), 100 U/ml penicillin and 100 µg/ml streptomycin solution (Sigma), 4 mM L-glutamine (Sigma), 2g/l sodium bicarbonate (Sigma) and 5% fetal calf serum (FCS, Biochrom) at 37°C temperature in the humidified atmosphere of 95% air and 5% carbon dioxide. Before confluence (once a week) the cells were harvested after a ten minutes incubation at 37°C in the presence of trypsin-EDTA at the concentrations of 0.05% and 0.02% of Hanks-balanced salt solution (Trypsin-EDTA, Sigma), respectively. Afterwards the cells were washed twice in complete medium and after last centrifugation seeded in a culture flask ( $2 \times 10^5$  cells in 5 ml of a fresh medium) for the four subsequent days.

#### <u>Experiment</u>

The cells were subjected to the trypsinization process as described above and suspended in the complete medium at a concentration of  $4 \times 10^5$  cells /ml. Thereafter 50 µl of cell suspension ( $2 \times 10^4$  cells) were placed in the each well of cell culture plates (96 Cell Culture Cluster Dish, Nunclon MicroWell Plates, NUNC) containing 130 µl of complete medium. After 24 hours of preincubation period at 37 °C in the humidified atmosphere of 95% air and 5% carbon dioxide, the 20 µl solution of investigated compounds were added. The cancer cells were cultured for 48 hours in the presence of melatonin at the final concentrations 10<sup>-7</sup> and 10<sup>-9</sup> M, 4P-PDOT and luzindole, both at the final concentrations 10-6-10-9 M applied alone and melatonin given together with 4P-PDOT or luzindole. The equal volume of culture medium (20 µl) and 95% ethanol (solvent for melatonin and 4P-PDOT) or DMSO (solvent for luzindole) at the highest concentration were added to the appropriate wells with control samples. The cell growth in the culture was measured using EZ4Y system (EZ4Y, Easy for You, The 4th Generation Non Radioactive Cell Proliferation & Cytotoxity Assay, Biomedica Gruppe, Austria, Bellco Biomedica Poland). The assay is based on the transformation of tetrazolium salt into colored soluble formazans as a result of the mitochondrial activity of the viable cells. The red soluble formazans, released to the culture medium, were determined by the extinction measurement using the enzyme-linked immunosorbent assay reader. The optical density (OD) of each sample was measured at 450 nm wave length.

### Statistical analysis

The data were statistically analysed by Statgraphics Centurion XV, using a one-way analysis of variance (ANOVA). Statistical differences between tested values were determined using the Least Significant Difference (LSD) test. Data was presented as the means  $\pm$  SEM. Optical density (OD) in control groups was assumed to be 100 percentage and OD of examined compounds were presented as a percentage of the control group. Differences were considered significant if p<0.05.

## RESULTS

Melatonin at both examined concentrations  $10^{-7}$  M and  $10^{-9}$  M significantly decreased the viability of cancers cells (Figures 1 and 2). The selective antagonist of MT<sub>2</sub> membrane receptor – 4P-PDOT and luzindole given alone at the concentrations  $10^{-6}$ ,  $10^{-7}$ ,  $10^{-8}$  and  $10^{-9}$  M did not influence the growth of Colon 38 cells. The addition of 4P-PDOT to MLT did not change the inhibitory effect of melatonin (Figure 1), whereas luzindole given together with MLT diminished, but failed to block totally, the oncostatic properties of melatonin (Figure 2).

## DISCUSSION

The obtained data confirm our earlier observation that melatonin at the physiological concentration  $(10^{-9} \text{ M})$  and also at the pharmacological level  $(10^{-7} \text{ M})$  inhibits the growth of murine colonic cancer (Melen-Mucha *et al.* 1998; Winczyk *et al.* 2002; 2001). It is also compat-

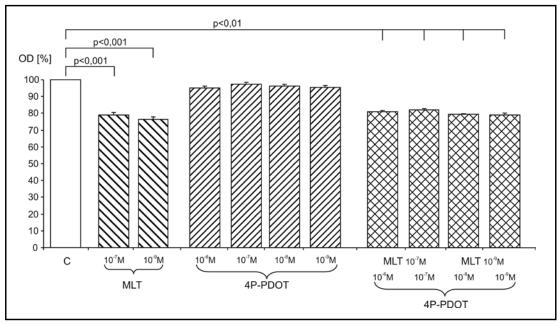


Fig. 1. The effects of melatonin (MLT) and 4-phenylo-2-propionamidotetralin (4-P-PDOT), applied alone and together, on the growth of Colon 38 cancer cells in vitro. Bars represent means ± SEM. C – control

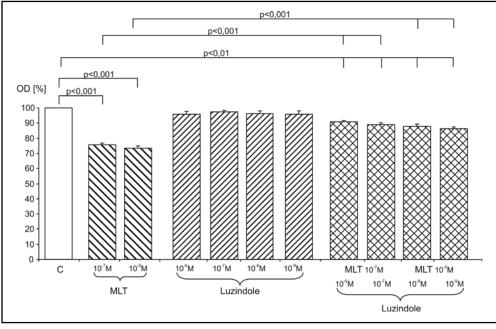


Fig. 2. The effects of melatonin (MLT) and luzindole, applied alone and together, on the growth of Colon 38 cancer cells in vitro. Bars represent means ± SEM. C - control

ible with the findings of Anisimov et al. (1997; 2000) showing that MLT inhibits the intestinal carcinogenesis and decreases the growth of carcinogen-induced colon tumors in rats. Melatonin also reduced the proliferation of the cells of murine CT-26 adenocarcinoma and human HT-29 colon cancer but the hormone was applied at high milimolar concentrations (Farriol et al. 2000; García-Navarro et al. 2007). The specific membrane melatonin receptors have been detected in animal and human intestines (Soták et al. 2006; Poon et al. 1996; 1997). We have shown that murine Colon 38 cancer possesses both membrane - MT<sub>1</sub> and MT<sub>2</sub> receptors (Karasek et al. 2002). The expression of MT<sub>1</sub> receptors and the involvement of this receptor subtype in oncostatic action of MLT have been documented in MCF-7 breast cancer, androgen-dependent LNCaP prostate cancer, PC12 pheochromocytoma and NIE-115 mouse neuroblastoma (Ram et al. 2002; Xi et al. 2001; Roth et al. 2001; Bordt et al. 2001). Our earlier study showed that N-[(4-methoxy-1H-indol-2-yl) methyl] propanamide - UCM 386, which is an antagonist of membrane MT<sub>1</sub> receptor and a weak agonist of membrane MT<sub>2</sub> receptor, did not change the inhibitory effect of MLT on Colon 38 cells (Winczyk et al. 2002). This observation suggested the participation of MT<sub>2</sub> receptor subtype in oncostatic action of the pineal hormone. The predominant involvement of MT<sub>2</sub> receptors in antiproliferative effects of melatonin has been indicated in ovarian carcinoma, endometrial cancer, choriocarcinoma and melanoma (Shiu et al. 1999; Kanishi et al. 2000; Petranka et al. 1999; Roberts et al. 2000. However, the present study shows that the antagonist of membrane MT<sub>2</sub> receptor 4P-PDOT neither has effect

alone nor changes the inhibitory action of melatonin on the viability of Colon 38 cells. Similar results have been obtained by other authors (García-Navarro et al. 2007). It has been shown that 4P-PDOT does not alter the antiproliferative effect of the pineal hormone on HT-29 human colon cancer cells. Thus, the abovementioned data indicate that the oncostatic action of MLT on human and murine colon cancer is rather independent of subtype MT<sub>2</sub> receptor. Nonetheless, we have observed that luzindole, the antagonist of both membrane melatonin receptors, clearly weakens but fails to block the inhibitory action of MLT on Colon 38 cells. Moreover, we cannot exclude that luzindole may interfere with melatonin action involving some other mechanisms that remain unknown. It is suggested that MLT exerts its anti-cancer effects also via other mechanisms than the modulation of membrane receptors. Melatonin easily enters inside the cells and may act in cytosol via the interaction with calmodulin and also modulates the expression genes by influencing the nucleus directly (Wiesenber et al. 1998; Benítez-King et al. 1991). In our previous study, the involvement of nuclear RZR/RORa receptors in oncostatic and immunomodulatory effects of melatonin were considered (Wiesenber et al. 1998; Garcia-Mauriño et al. 1998). The RZR/ROR receptors were cloned in two groups and received the respective names of: retinoid Z receptor (RZR) and retinoid acid receptor-related orphan receptor (ROR) (Becker-André et al. 1993; Giguère et al. 1994). In human and mammals the  $\alpha$ -subtype of these receptor are widely expressed outside the brain, in peripheral tissues. Our data showed that the cells of murine Colon 38 cancer possess the RORa receptors (Karasek et al. 2002). Besides, in some

of human cancer cells such as: MCF-7 breast cancer, melanoma, Du-145 androgen-impendent and LNCaP androgen-dependent prostate cancer, RORa transcripts have been also identified (Dai et al. 2001; Moretti et al. 2000; 2001a; Fischer et al. 2006). Although the interaction of melatonin with RZR/RORa receptors is still under discussion, some data supports the participation of these nuclear receptors in oncostatic action of MLT. Melatonin and a thiazolidinedione derivative, CGP 52608, which was characterized as a selective ligand for RZR/RORa receptors, in similar manner inhibit the proliferation of several human cancers cells lines: LNCaP and Du-145 prostate cancers, BG-1 ovarian adenocarcinoma, MCF-7 breast cancer, HT-29 colon cancer (García-Navarro et al. 2007; (Dai et al. 2001; Moretti et al. 2000; 2001a; 2001b; Petranka et al. 1999). Moreover, we have found that both compounds exerted comparable anti-proliferative effects on Colon 38 cancer and on rat experimental pituitary tumor (Karasek et al. 1998; 1999). Our earlier study also showed that melatonin and CGP 52608 enhance apoptosis in the transplantable murine colon adenocarcinoma (Winczyk et al. 2001). The results of the experiments conducted in our laboratory with CGP 55644 – an antagonist of RZR/RORa receptors also support the participation of nuclear receptors in the oncostatic action of the hormone. We have documented that CGP 556444 diminishes the antiproliferative effect of MLT and blocks its proapoptotic action in Colon 38 tumors (Winczyk et al. 2002). Moreover, other studies have shown that antagonist of RZR/RORa receptors blocks the inhibitory effect of MLT on the growth of diethylstilbestrolinduced rat prolactin-secreting pituitary tumor cells in vitro (Karasek et al. 2003). However, regardless of the outcome reported, the independent effects of both membrane and nuclear receptors of MLT on cancer growth should be considered. It has been shown that the high level of nitric oxide synthase (NOS) correlates with invasiveness and progression of colon carcinoma in humans (Lagares-Garcia et al. 2001). Melatonin binding with calmodulin decreases the activity of NOS and in this way may exert its oncostatic action (Pozo et al. 1997). The other receptor-independent mechanism in which melatonin may affect cancer growth is its ability to scavenge free radicals and to protect cells from oxidative damage.

Summing up, the present results and our previous study conducted on Colon 38 adenocarcinoma cells indicate that membrane melatonin receptors are not indispensable for the oncostatic action of melatonin and other pathways such as nuclear signaling and receptorindependent mechanism may be also involved.

#### ACKNOWLEDGEMENTS

This study was supported by the Medical University of Lodz, project No. 502-15-503.

#### REFERENCES

- 1 Anisimov VN, Popovich IG, Shtylik AV, Zabezhinski MA, Ben-Huh H, Gurevich P, *et al* (2000). Melatonin and colon carcinogenesis: Effect of melatonin on proliferative activity and apoptosis in colon mucosa and colon tumors induced by 1,2-dimethylhydrazine in rats. Exp Toxicol Pathol. **52**: 71–76.
- 2 Anisimov VN, Popovich IG, Zabezhinski MA (1997). Melatonin and colon carcinogenesis: I. Inhibitory effect of melatonin on development of intestinal tumors induced by 1,2-dimethylhydrazine in rats. Carcinogenesis. **18**: 1549–1553.
- 3 Becker-André M, André E, DeLamarter JF (1993). Identification of nuclear receptor mRNAs by RT-PCR amplification of conserved zinc-finger motif sequences. Biochem Biophys Res Commun. 194: 1371–1379.
- 4 Benítez-King G, Huerto-Delgadillo L, Antón-Tay F (1991). Melatonin modifies calmodulin cell levels in MDCK and N1E-115 cell lines and inhibits phosphodiesterase activity in vitro. Brain Res. **557**: 289–292.
- 5 Blask DE, Sauer LA, Dauchy RT, Holowachuk EW, Ruhoff MS, Kopff HS (1999). Melatonin inhibition of cancer growth in vivo involves suppression of tumor fatty acid metabolism via melatonin receptor-mediated signal transduction events. Cancer Res. 59: 4693–701.
- 6 Bordt SL, McKeon RM, Li PK, Witt-Enderby PA, Melan MA (2001). N1E-115 mouse neuroblastoma cells express MT1 melatonin receptors and produce neurites in response to melatonin. Biochim Biophys Acta. **1499**: 257–264.
- 7 Bubenik GA (2008). Thirty four years since the discovery of gastrointestinal melatonin. J Physiol Pharmacol. 59 Suppl 2: 33–51.
- 8 Calberg C, Wiesenberg I (1995). The orphan receptor family RZR/ ROR, melatonin and 5-lipooxygenase: An unexpected relationship. J Pineal Res. 18: 171–178.
- 9 Corbett TH, Griswold DP, Jr, Roberts BJ, Peckham C, Schabel FM, Jr (1975). Tumor induction relationship in development of transplantable cancers of the colon in mice for chemotherapy assays, with a note on carcinogen structure. Cancer Res. 35: 2434–2439.
- 10 Dai J, Ram PT, Yuan L, Spriggs LL, Hill SM (2001). Transcriptional repression of RORalpha activity in human breast cancer cells by melatonin. Mol Cell Endocrinol. **176**: 111–120.
- 11 Dubocovich ML, Cardinali DP, Guardiola-Lemaitre B, Hagan RM, Krause DN, Sugden D, Vanhoutte PM, *et al* (1998). Yocca FD. Melatonin receptors. In: The IUPHAR Compendium of receptor characterisation and classification. London: IUPHAR Media. pp 187–193.
- 12 Farriol M, Venereo Y, Orta X, Castellanos JM, Segovia-Silvestre T (2000). In vitro effects of melatonin on cell proliferation in a colon adenocarcinoma line. J Appl Toxicol. **20**: 21–24.
- 13 Fischer TW, Zmijewski MA, Zbytek B, Sweatman TW, Slominski RM, Wortsman J, *et al* (2006). Oncostatic effects of the indole melatonin and expression of its cytosolic and nuclear receptors in cultured human melanoma cell lines. Int J Oncol. **29**: 665–672.
- 14 Garcia-Mauriño S, Gonzalez-Haba MG, Calvo JR, Goberna R, Guerrero JM (1998). Involvement of nuclear binding sites for melatonin in the regulation of IL-2 and IL-6 production by human blood mononuclear cells. J Neuroimmunol. **92**: 76–84.
- 15 García-Navarro A, González-Puga C, Escames G, López LC, López A, López-Cantarero M, et al (2007). Cellular mechanisms involved in the melatonin inhibition of HT-29 human colon cancer cell proliferation in culture. J Pineal Res.43: 195–205.
- 16 García-Santos G, Antolín I, Herrera F, Martín V, Rodriguez-Blanco J, del Pilar Carrera M, et al (2006). J Pineal Res. Melatonin induces apoptosis in human neuroblastoma cancer cells. 41: 130–135.

- 17 Giguère V, Tini M, Flock G, Ong E, Evans RM, Otulakowski G (1994). Isoform-specific amino-terminal domains dictate DNAbinding properties of ROR alpha, a novel family of orphan hormone nuclear receptors. Genes Dev. **8**: 538–553.
- 18 Kanishi Y, Kobayashi Y, Noda S, Ishizuka B, Saito K (2000). Differential growth inhibitory effect of melatonin on two endometrial cancer cell lines. J Pineal Res. 28: 227–233.
- 19 Karasek M, Carrillo-Vico A, Guerrero JM, Winczyk K, Pawlikowski M (2002). Expression of melatonin MT(1) and MT(2) receptors, and ROR alpha(1) receptor in transplantable murine Colon 38 cancer. Neuro Endocrinol Lett. **23** Suppl 1: 55–60.
- 20 Karasek M, Gruszka A, Lawnicka H, Kunert-Radek J, Pawlikowski M (2003). Melatonin inhibits growth of diethylstilbestrol-induced prolactin-secreting pituitary tumor in vitro: possible involvement of nuclear RZR/ROR receptors. J Pineal Res. 34: 294–296.
- 21 Karasek M, Pawlikowski M (1999). Antiproliferative effects of melatonin and CGP 52608. Biol Signals Recept. 8: 75–78.
- 22 Karasek M, Winczyk K, Kunert-Radek J, Wiesenberg I, Pawlikowski M (1998). Antiproliferative effects of melatonin and CGP 52608 on murine Colon 38 adenocarcinoma in vitro and in vivo. Neuro-endocrinol Lett. **19**: 71–78.
- 23 Lagares-Garcia JA, Moore RA, Collier B, Heggere M, Diaz F, Qian F (2001). Nitric oxide synthase as a marker in colorectal carcinoma. Am Surg. 67: 709–713.
- 24 Lee PP, Pang SF (1993). Melatonin and its receptors in the gastrointestinal tract. Biol Signals. 2:181–193.
- 25 Marelli M, Limonta P, Maggi R, Motta M, Moretti RM (2000). Growth-inhibitory activity of melatonin on human androgenindependent DU 145 prostate cancer cells. Prostate. 45: 238–244.
- 26 Martín-Renedo J, Mauriz JL, Jorquera F, Ruiz-Andrés O, González P, González-Gallego J (2008). Melatonin induces cell cycle arrest and apoptosis in hepatocarcinoma HepG2 cell line. J Pineal Res. 45: 532–540.
- 27 Melen-Mucha G, Winczyk K, Pawlikowski M (1998). Somatostatin analogue octreotide and melatonin inhibit bromodeoxyuridine incorporation into cell nuclei and enhance apoptosis the transplantable murine Colon 38 cancer. Anticancer Res. **18**: 3615–3620.
- 28 Moretti RM, Marelli MM, Maggi R, Dondi D, Motta M, Limonta P (2000). Antiproliferative action of melatonin on human prostate cancer LNCaP cells. Oncol Rep. 7: 347–351.
- 29 Moretti RM, Marelli MM, Motta M, Polizzi D, Monestiroli S, Pratesi G, *et al* (2001a). Activation of the orphan nuclear receptor ROR alpha induces growth arrest in androgen-independent DU 145 prostate cancer cells. Prostate. **46**: 327–335.
- 30 Moretti RM, Montagnani Marelli M, Motta M, Limonta P (2001b). Oncostatic activity of a thiazolidinedione derivative on human androgen-dependent prostate cancer cells. Int J Cancer. 92: 733–737.
- 31 Pajtasz-Piasecka E, Szyda A, Rossowska J, Krawczenko A, Indrová M, Grabarczyk P, *et al* (2004). Loss of tumorigenicity of murine colon carcinoma MC38/0 cell line after transduction with a retroviral vector carrying murine IL-12 genes. Folia Biol (Praha). 50: 7–14.
- 32 Pawlikowski M, Winczyk K, Karasek M (2002). Oncostatic action of melatonin: facts and question marks. Neuroendocrinol Lett. 23 Suppl 1: 24–29.

- 33 Petranka J, Baldwin W, Biermann J, Jayadev S, Barrett JC, Murphy E (1999). The oncostatic action of melatonin in an ovarian carcinoma cell line. J Pineal Res. 26: 129–136.
- 34 Poon AM, Chow PH, Mak AS, Pang SF (1997). Autoradiographic localization of [125I]iodomelatonin binding sites in the gastrointestinal tract of mammals including humans and birds. J Pineal Res. **23**: 5–14.
- 35 Poon AM, Mak AS, Luk HT (1996). Melatonin and 2[125l]iodomelatonin binding sites in the human colon. Endocr Res. 22: 77–94.
- 36 Pozo D, Reiter RJ, Calvo JR, Guerrero JM (1997). Inhibition of cerebellar nitric oxide synthase and cyclic GMP production by melatonin via complex formation with calmodulin. J Cell Biochem. 65: 430–442.
- 37 Ram PT, Dai J, Yuan L, Dong C, Kiefer TL, Lai L, et al (2002). Involvement of the mt1 melatonin receptor in human breast cancer. Cancer Lett. 179: 141–150.
- 38 Roberts JE, Wiechmann AF, Hu DN (2000). Melatonin receptors in human uveal melanocytes and melanoma cells. J Pineal Res. 28: 165–171.
- 39 Roth JA, Rosenblatt T, Lis A, Bucelli R (2001). Melatonin-induced suppression of PC12 cell growth is mediated by its Gi coupled transmembrane receptors. Brain Res. **919**: 139–146.
- 40 Shiu SY, Li L, Xu JN, Pang CS, Wong JT, Pang SF (1999). Melatonininduced inhibition of proliferation and G1/S cell cycle transition delay of human choriocarcinoma JAr cells: possible involvement of MT2 (MEL1B) receptor. J Pineal Res. 27: 183–192.
- 41 Shiu SY, Xi SC, Xu JN, Mei L, Pang SF, Yao KM, *et al* (2000). Inhibition of malignant trophoblastic cell proliferation in vitro and in vivo by melatonin. Life Sci. **67**: 2059–2074.
- 42 Soták M, Mrnka L, Pácha J (2006). Heterogeneous expression of melatonin receptor MT1 mRNA in the rat intestine under control and fasting conditions. J Pineal Res. **41**: 183–188.
- 43 Wiesenber I, Missbach M, Carlberg C (1998). The potential role of the transcription factor RZR/ROR as a mediator of nuclear melatonin signalling. Restr Neurol Neurosci. 12: 143–150.
- 44 Winczyk K, Pawlikowski M, Guerrero JM, Karasek M (2002). Possible involvement of the nuclear RZR/ROR-alpha receptor in the antitumor action of melatonin on murine Colon 38 cancer. Tumour Biol. **23**: 298–302.
- 45 Winczyk K, Pawlikowski M, Karasek M (2001). Melatonin and RZR/ ROR receptor ligand CGP 52608 induce apoptosis in the murine colonic cancer. J Pineal Res. **31**: 179–182.
- 46 Winczyk K, Pawlikowski M, Lawnicka H, Kunert-Radek J, Spadoni G, Tarzia G, et al (2002). Effects of melatonin and melatonin receptors ligand N-[(4-methoxy-1H-indol-2-yl)methyl]propanamide on murine Colon 38 cancer growth in vitro and in vivo. Neuro Endocrinol Lett. 23 Suppl 1: 50–54.
- 47 Winczyk K, Pawlikowski M, Lawnicka H, Kunert-Radek J, Spadoni G, Tarzia G, *et al* (2002). Effects of melatonin and melatonin receptors ligand N-[(4-methoxy-1H-indol-2-yl)methyl]propanamide on murine Colon 38 cancer growth in vitro and in vivo. Neuro Endocrinol Lett. **23** Suppl 1: 50–54.
- 48 Xi SC, Siu SW, Fong SW, Shiu SY (2001). Inhibition of androgensensitive LNCaP prostate cancer growth in vivo by melatonin: association of antiproliferative action of the pineal hormone with mt1 receptor protein expression. Prostate. **46**: 52–61.