N-3 polyunsaturated fatty acids in psychiatric diseases: Mechanisms and clinical data

Miroslav ZEMAN^{1*}, Roman JIRÁK^{2*}, Marek VECKA^{1*}, Jiří RABOCH², Aleš ŽÁK¹

¹ IVth Department of Internal Medicine, Charles University in Prague, First Faculty of Medicine and General University Hospital, in Prague Czech Republic

2 Psychiatric Clinic, Charles University in Prague, First Faculty of Medicine and General University Hospital in Prague, Czech Republic

* these authors contributed equally to the article

Correspondence to: Assoc. Prof. Miroslav Zeman, MD., PhD. IVth Department of Internal Medicine, Charles University in Prague, First Faculty of Medicine and General University Hospital in Prague U Nemocnice 2, 128 08, Prague 2, Czech Republic. E-MAIL: mirozem@centrum.cz

Submitted: 2012-09-25 Accepted: 2012-11-17 Published online: 2012-12-28

Key words: docosahexaenoic acid; arachidonic acid; eicosanoids; docosanoids; depression; psychiatric disease

Neuroendocrinol Lett 2012; 33(8):736-748 PMID: 23391975 NEL331112R01 © 2012 Neuroendocrinology Letters • www.nel.edu

Abstract The lipids constitute majority of dry weight of mature human brain. From lipids, 35% is comprised of PUFA with long chain (LC-PUFA), especially docosahexaenoic acid (DHA) of n-3 family and arachidonic acid (AA) of n-6 family. Humans are dependent on dietary intake of both AA and DHA. Interestingly, the dietary n-6/n-3 ratio increased considerably during last century. LC-PUFAs play numerous roles in the brain, including structural (forming the physico-chemical properties in the lipid bilayer of cellular membranes) and signaling ones. Moreover, they influence neurogenesis and neurotransmission within the nervous tissue. The metabolites of PUFA modulate immune and inflammatory processes in the brain, oxidative stress as well as its consequences. Of high importance is also their connection with several metabolic factors involved in the proper function of the brain and/or were discovered to play a role in the pathogenesis of neuropsychiatric diseases - melatonin, homocysteine, leptin, and adiponectin. This review gives short view of the metabolism and possible mechanisms of PUFA n-3 action in the brain, and their role in the pathogenesis of psychiatric diseases.

Abbreviations:		k Ras	- Kirsten RAt Sarcoma viral oncogene homolog
PUFA	 polyunsaturated fatty acids 	PI3-kinase	- phosphatidylinositol-3 kinase
AA	- arachidonic acid	MMSE	- Mini Mental State Examination
DHA	- docosahexaenoic acid	βFGF	- Fibroblast growth factor-beta
EPA	- eicosapentaenoic acid	iL	- interleukin
CNS	- central nervous system	MB-COMT	- membrane-bound catechol-O-methyltransferase
LCPUFA	- long-chain polyunsaturated fatty acids	Erk 1/2	- extracellular signal-regulated kinase-1/2
TRP channels - transient receptor potential channels		sorLA	- sorting protein-related receptor
GSH-Px	- glutathione peroxidase	5-HIAA	- 5-Hydroxyindoleacetic acid
MARCKS	- myristoylated alanine-rich C kinase substrate	RCT	- randomized clinical trial
VMAT2	- vesicular monoamine transporter 2	O&NS	 oxidative and nitrosative stress
raf-1	- the acronym RAF is derived from Rapidly Accelerated Fibrosarcoma	ME/CFS	 myalgic encephalomyelitis/chronic fatigue syndrome

1. INTRODUCTION

In the last two centuries, the human nutrition has considerably changed in the sense of higher total energy intake. Moreover, dietary content of saturated fatty acids (SFA) has increased at the expense of polyunsaturated fatty acids (PUFA), especially those of n-3 family (Simopoulos 1999). These changes are thought to be related with significant increase of prevalence of obesity, type 2 diabetes mellitus and cardiovascular diseases (Reddy & Katan 2004; Simopoulos 2008). Furthermore, there is growing evidence that the pathogenesis of some neuropsychiatric disorders (depression, schizophrenia, Alzheimer disease) also involves (apart from genetic) the nutritional factors, such as the fat intake, the content of PUFA n-3 and their ratio to PUFA n-6 (Young & Conquer 2005).

2. POLYUNSATURATED FATTY ACIDS – OVERVIEW AND METABOLISM

Fatty acids (FA) are higher monocarboxylic acids, most of them with even number of carbons in the molecule. According to the number of double bonds in hydrocarbon chain, FA are traditionally divided into saturated FA (SFA, without double bond), monounsaturated FA (MFA, one double bond), and PUFA (with two and more double bonds). The principal metabolic pathways for changes in FA chain are presented on Figures 1a and 1b.

In humans, the PUFA n-3 and n-6 bound in lipids form an integral part of biological membranes. These acids modulate membrane fluidity, the interactions between lipid and protein part and also modulate the function of various enzymes, transporters as well as membrane receptors (Murphy 1990). The PUFA can change the cellular response on the extracellular signals with acylation of intracellular proteins (Pike 2004), they are also structural parts of second messengers, such as diacylglycerol (DAG), the functions of which they can influence in this way (Hichami et al. 2005). The last, but not the least, issue is the possible action of PUFA on the transcription of many genes (Deckelbaum et al. 2006). The enzymatic oxidation of twenty-carbon PUFA [dihomo-gammalinolenic (DGLA), 20:3n-6; arachidonic (AA), 20:4n-6; eicosapentaenoic (EPA), 20:5n-3] (Fitzpatrick & Soberman 2001) gives rise to the eicosanoids, the compounds with many important biological effects. Other oxidative metabolites of PUFA include resolvins (resolutionphase interaction products) with immunoregulatory and antiinflammatory effects, which are synthesized from EPA and DHA: series E resolvins (RvE) and D (RvD), and (neuro)protectins from DHA in glial cells of CNS; these derivatives downregulate the expression of inflammatory cytokines (see Serhan et al. 2011 for review).

3. PUFA IN CENTRAL NERVOUS SYSTEM

<u>3.1 PUFA content in brain and the relation to other body</u> <u>compartments</u>

The lipids constitute app. 50-60% of dry weight of mature human brain. From these, 35% is comprised of PUFA with long chain (LC-PUFA), especially AA and DHA. The analyses of FA profile in the fat from frontal cortex revealed that the PUFA n-6 content reached 17% (mainly AA) and the content of PUFA n-3 was as much as 14% (only DHA, other PUFA n-3 - EPA, ALA and 22:5n-3 represented less than 1%) (McNamara & Carlson 2006). The arachidonic acid can be found in all tissues, whereas DHA prevails in grey matter, retina and testes. Interestingly, the content of AA and DHA increases during the period of maximal brain growth, i.e. three months before delivery and for the short period of time in the postnatal life (Wainwright 2000). The postmortem analyses of FA content in brain tissues of individuals with neuropsychiatric diseases are not consistent. Many confounding factors can bias the results, such as undetected exposition to antidepressants, mood stabilizers, antipsychotics, long-time alcohol abuse, or the fat content/quality (McNamara & Jandacek 2011). The high dietary intake of LCPUFA n-3 was selectively associated with higher volume of gray matter in the region taking part in corticolimbic circuitry (subgenual anterior cingulate cortex, right hippocampus and right amygdala), the dysfunction of which represents pathologic marker of mood disorders such as depression (Conklin et al. 2007). The dietary intake of EPA and DHA correlates with their raised concentrations in plasma as well as erythrocyte phospholipids and it also correlates with decreased content of AA (Arterburn et al. 2006). Carver et al. analyzed FA content in the samples of brain cortex of 58 individuals aged 2-82 years; the authors found statistically significant correlations between AA as well as DHA content in erythrocytes and brain cortex in samples of those older than 18 years (Carver et al. 2001).

3.2 The effects of PUFA in CNS

PUFA were found to exhibit their effects in CNS through various mechanisms connected with (i) characteristics of neuronal membranes, (ii) intracellular signalization, gene expression and transcription, (iii) modulation of immune and inflammatory processes, (iv) effects on neurogenesis, (v) neurotransmission, especially of serotonin and dopamine type, (vi) crosstalk with metabolic factors functioning in CNS.

3.2.1. Physical-chemical characteristics of neuronal membranes

PUFA considerably influence the physical-chemical characteristics of membranes and consequently the function of the structures connected with membranes (ion channels, receptors, cellular transporters etc.) (Murphy 1990; Mourek *et al.* 2009). The increased membrane fluidity enhances lateral movements of

Miroslav Zeman, Roman Jirák, Marek Vecka, Jiří Raboch, Aleš Žák

proteins within the lipid bilayer and enables easier interactions between the proteins located in the membrane. Sufficient content of DHA in phospholipids of retinal rods supports the activation of rhodopsin and the conformational changes leading to the transformation into the active form, metarhodopsin II (Stinson *et al.* 1991). Rhodopsin, which belongs to the G-protein coupled receptors (GPCR), is responsible for night vision. DHA increases the activity of Na⁺/K⁺-ATPase, which is located in the neuronal membranes and keeps the resting membrane potential (Gerbi *et al.* 1998). On the contrary, DHA inhibits Ca²⁺-ATPase (Kearns & Haag 2002). DHA accumulates in neuronal synaptic membranes mostly in phosphatidylethanolamines (PE) and phosphatidylserines (PS). The decreased content of DHA in PS due to DHA deficiency conduces to low



Fig. 1a 1b. Docosahexaenoic acid (DHA, 22:6n-3) is fatty acid of n-3 family with 22 carbon atoms in the chain and 6 double bonds. The n-3 stands for the position of the first double bond from the methyl end of the chain, i.e. the bond is located between the third and the fourth carbons. The humans can synthesize SFA and MUFA (see next Figure), but they cannot introduce the double bond nearer than to the ninth carbon to the carboxyl end (D9 desaturase). Therefore, humans are not able to biosynthesize double bonds on the positions n-3 and n-6. The linoleic (18:2n-6) and α-linolenic (18:3n-3) acids, which are the precursors of respective PUFA families, must be ingested in the diet and are referred to as essential FA (EFA).

expression of membrane protein MARCKS (*myris-toylated alanine-rich C kinase substrate*) and disturbed homeostasis of intracellular calcium (McNamara & Carlson 2006). PUFA n-3 can modulate either directly or indirectly the function of TRP channels (*transient receptor potential channels*), which take part in neurotransmitter release, redox signalization and modulation of Ca²⁺ level in mitochondria (Leonelli *et al.* 2011). The changes in function of the abovementioned channel proteins could participate in the pathogenesis of neurodegenerative and psychiatric diseases (Chahl 2011).

3.2.2. Intra- and intercellular signalization, gene expression and transcription The membrane phospholipids are potential source of signaling molecules with both intra- and extra-cellular mode of action. The phospholipase A_2 liberates AA, EPA and DHA from molecules of PL. These acids are metabolized with cyclooxygenases, lipoxygenases or cytochrome P450 monooxygenases transforming the respective FA into eicosanoids (AA and EPA) or docosanoids (DHA) (See Figure 2). Eicosanoids and docosanoids easily diffuse through neuronal membranes and can act as autocrine or paracrine signaling molecules. The PUFA molecules themselves can be intracellular second messengers or their content in signaling structures influences the function of transcriptional factors and gene expression (Deckelbaum et al. 2006). For example, DHA released from membrane with $iPLA_2$ (calcium-independent phospholipase A_2) takes part in the modulation of signal transduction or as a part of the diacylglycerol (DAG) molecule, and DHA enhances DAG-dependent activation of protein kinase C. DHA can also be transformed into docosanoids that have antiinflammatory effects and protect neurons from oxidative damage (Bazan 2009, Shimazawa et al. 2009). PUFA n-3 influence the gene expression not only in liver and adipose tissue (Deckelbaum et al. 2006), but also in brain. These genes govern synaptic plasticity, signal transduction, the interactions of cytoskeleton with cellular membrane, the formation of ion channels, regulatory proteins and other factors (expression of synuclein α and γ , calmodulins, actin, ras oncogenes and other) (Kitajka et al. 2002).



Fig. 2. Production of oxylipins from polyunsaturated fatty acids.

3.2.3. Modulation of inflammation and oxidative stress

The twenty-carbon derivatives of AA, EPA and DGLA play an important role in the human metabolism (Fitzpatrick & Soberman 2001). In general, the metabolites derived from AA are proinflammatory, whereas the EPA and DHA derivatives are antiinflammatory (see Figure 2). In membrane PL, AA is bound preferentially in PE, phosphatidylcholine and phosphatidylinositol. Arachidonic acid and its metabolites are involved in synaptic signalization, neurotransmitter release; they also influence brain perfusion and expression of neuronal genes (Bosetti 2007). The disturbed metabolism of AA in brain was related to many neurodegenerative and psychiatric diseases, such as (e.g.) Alzheimer disease, and depressive disorders (Bazan et al. 2002, Sublette et al. 2004). The inflammatory cytokines (IL-1 β , IL-2, IL-6, interferon- γ and TNF α), activate hypothalamo-hypophysary axis, decrease the concentrations of neurotransmitter precursors and further change their metabolism (Maes & Smith 1998). The AA molecules, released from PL in cellular membranes, are transformed by cyclooxygenases (COX), which are also referred to as prostaglandin H synthases (PGHS). PGHS produces PGH₂, the precursor for prostanoids, which are compounds with high tissue specificity. In brain, PGH₂ is a precursor for PGE₂ (Bosetti 2007). EPA can inhibit the production of eicosanoids from AA by direct inhibition of COX. Moreover, EPA and DGLA compete with AA for many eicosanoid forming enzymes, which further underlines their antiinflammatory effects. Arachidonic acid stimulates the biosynthesis of glutamate, the excess of which potentiates the neuronal destruction resulting from the overproduction of oxygen radicals. On the contrary, DHA increases the level of serotonin and acetylcholine (Minami et al. 1997, Vancassel et al. 2008, Mc Namara et al. 2010) and may act as an antioxidant (Shimazava et al. 2009).

3.2.4. Neurotransmission

Both the inter- and intra-cellular signal transduction is influenced by the concentration of neurotransmitters, their storage in vesicles and the function of presynaptic and postsynaptic receptors. The pathophysiology of neuropsychiatric diseases involves mainly neurotransmitters of catecholamine type (dopamine, noradrenaline, and serotonine), amino acids (glutamate, y-aminobutyric acid) or acetylcholine. The animal models give us some insight into the interaction between the content of PUFA n-3 and neurotransmitters. The PUFA n-3 deficient rats had by 40-60% lower content of dopamine in frontal cortex (Delion et al. 1994), and lower content of dopamine in synaptic vesicles, higher expression of D2 receptors and lower content of vesicular monoamine transporter 2 (VMAT2) in nucleus accumbens (NAC), whereas the number of D2 receptors decreased in frontal cortex (Zimmer et al. 2002). NAC belongs to mesolimbic dopaminergic system that plays a role in cognitive and affective functions The animals deficient in PUFA n-3 exhibit dysbalance between dopaminergic pathways, which is shifted to mesolimbic one. This dysbalance can participate in observed changes in attention, motivation and emotion motivation, reward answer and learning skills (Chalon 2006).

3.2.5. Neurogenesis and neuronal survival

Many studies proved the significance of PUFA n-3 (especially DHA) for the proper brain development in children; the importance for adult brain is currently under discussion. The experimentally induced lower intake of PUFA n-3 led to substantial decrease of DHA content in neurons, which was accompanied with higher content of PUFA n-6. In the nervous tissue, DHA is abundant in phospholipids, such as PE and PS. Higher intake of DHA causes increased synthesis of PS that is involved in cellular signalization. Elevated content of PS leads to enhanced translocation of raf-1 and PI₃ kinases to plasma membrane, increased activity of PI₃ kinase, which consequently diminishes the activation of caspase-3 and inhibits apoptosis (Marszalek and Lodish 2005). Interestingly, in some other cell types DHA can trigger apoptosis, as was shown for CaCo-2 colon cancer cell lines (Narayanan et al. 2001).

PUFA n-3 can favorably influence neoneurogenesis. The changes in the content (and the fluidity) of the neuronal membrane bring about the changes in quaternary structure of membrane proteins, including transporters and receptors (Bourre et al. 1991). The membrane fluidity influences the serotonin binding to the membranes and, secondarily, also the neurogenesis. The positive influence of PUFA n-3 is exerted via their effect on inflammatory cytokines (decreased concentrations of IL-1β and TNFα) (Shahbakhti *et al.* 2004). The activation of microglial cells during chronic inflammation disrupts the neurogenetic processes in hippocampus, whereas the inhibition of inflammation by tetracycline restores the neurogenesis in experimental rats (Ekdahl et al. 2003). Another possible mechanism, by which PUFA n-3 favorably influence the brain neurogenesis, includes higher expression of brain-derived neurotrophic factor (BDNF)(Cysneiros et al. 2010).

3.2.6. Metabolic pathways in CNS

3.2.6.1. Homocysteine (Hcy)

Hyperhomocysteinemia (HHcy) is not only risk factor for atherosclerosis and thrombosis, but also higher levels of Hcy are connected with inborn defects of neuronal crest, autoimmune diseases, osteoporotic fractures and neuropsychiatric diseases. HHcy was observed in euthymic patients with bipolar disorders (Dittmann *et al.* 2007), schizophrenics (Henderson *et al.* 2006), and patients with depressive disorders (Folstein *et al.* 2007). The concentration of homocysteine (Hcy) is the result of the interaction between dietary and genetic factors; most of the mildly elevated levels of Hcy is caused by the deficiency in vitamins B, especially

folate, B12 and B6 (pyridoxal phosphate), which are the cofactors of metabolic pathways keeping stable level of Hcy (Hankey & Eikelboom 1999). The concentration of Hcy can be decreased with administration of PUFA n-3 (Zeman et al. 2006); the underlying mechanism is probably connected with the expression of enzymes taking part in Hcy metabolism (Huang et al. 2012). The Hcy concentration negatively correlated with the content of DHA both in serum platelet PL (Li et al. 2007) and plasma PC (Zeman et al. 2006). HHcy in depressive patients was accompanied with low levels of folate and S-adenosylmethionine (SAM) in serum and cerebrospinal fluid, also with low 5-HIAA and homovanilic acid (Bottiglieri et al. 2000). On the contrary, the treatment with SAM had antidepressive effects (Miller 2008). In HHcy states, the enzymes catalyzing methylation of catecholamines are inhibited. The brains of the patients with Alzheimer disease and HHcy had lower activity of phenylethanolamine N-methyltransferase (PNMT) and catechol O-methyltransferase (COMT) (Kennedy et al. 2004).

Other possible connection between HHcy and psychiatric diseases is the relationship of HHcy with dysregulated methylation of DNA in the gene promotor regions. In the study by Abdomaleky et al. (2006), it was found that MB-COMT promotor DNA is often hypomethylated in schizophrenics as well as in the bipolar disorder patients, which leads to overexpression of MB-COMT and increased degradation of dopamine in the frontal lobe. The experiments carried out on rats proved that high levels of Hcy attenuate the neurogenesis in hippocampus via inhibition of FGF-dependent activation of Erk1/2 (Rabaneda et al. 2008). Ravaglia et al. (2003) described in healthy individuals independent and gradual association of high level of serum Hcy with cognitive disorder measured with MMSE. HHcy is also known to raise the content of β -amyloid due to overexpression of y-secretase and phosphorylation of amyloid precursor protein in experimental animals (Zhang et al. 2009).

3.2.6.2 Melatonin

Melatonin, (N-acetyl-5-methoxytryptamine) is produced mainly in the pineal gland, but also in other tissues. In humans, it plays an important role in the regulation of circadian biorhythms. The production of melatonin is higher in the dark periods, whereas its production decreases in the daylight. The regulation of melatonin biosynthesis is influenced by hepoxilins (derivatives of PUFA n-3 and PUFA n-6). PUFA n-3 and PUFA n-6 probably regulate the formation of hepoxilins and melatonin in pineal gland (Catalá 2009). Melatonin acts as an effective scavenger of hydroxyl and peroxyl radicals and also induces the production of antioxidant enzymes, such as GSH-Px (Reiter et al. 2008). The antioxidative effects of melatonin could be of high importance for favorable influence on neurodegenerative and psychiatric diseases (Alzheimer disease,

schizophrenia, affective disorders), in the pathogenesis of which the oxidative stress is highly involved (Kodydková *et al.* 2009; Maes 2011a; Anderson *et al.* 2012). One study found neuroprotective effect of melatonin administration in the model of hypoxic-ischemic encephalopathy in rats (Signorini *et al.* 2009). However, the studies examining the antidepressive effect of melatonin gave contradictory results. One derivative of melatonin, agomelatonin, has been reported to be an effective antidepressant therapy (Fornaro *et al.* 2010).

3.2.6.3 Leptin and adiponectin

Brain leptin significantly influences neurogenesis, axon growth and synapsis formation in the hypothalamus, hippocampus and other brain regions including the cortex (Paz-Filho et al. 2010). Furthermore, the interaction between leptin and serotonergic system was proved (Charnay et al. 2000). Leptin modulates various brain functions, e.g. the learning and memory skills. It is supposed that the dysregulation of leptin is involved in the pathogenesis of depression and Alzheimer disease; the leptin resistance cannot be excluded as well (Lu 2007). The mechanisms of action of adiponectin in brain have not been elucidated yet. Several studies described lower serum adiponectin in depressive disorder (Zeman et al. 2009), and the concentration of adiponectin negatively correlated with the stage of depression expressed as HAMD score (Zeman et al. 2009).

PUFA n-3 can modulate the levels of adipokines (including adiponectin and leptin) and consequently their effect in brain tissue. In the study by Peyron-Caso (Peyron-Caso et al. 2002), the administration of PUFA n-3 in insulin resistant rats increased leptin levels, other study (Rossi et al. 2005) found that PUFA n-3 counteracted the inhibition of secretion of leptin that was induced by long-time saccharose rich diet. However, another study did not find any effect of PUFA n-3 on leptin level (Flachs et al. 2006). Adiponectin levels are reported to be increased (Flachs et al. 2006; Rossi et al. 2005), but not always (Lorente-Cebrián et al. 2006) after the PUFA n-3 administration. The principal metabolic effects of PUFA n-3 include their apparent hypotriglyceridemic action (Harris et al. 1989; Žák et al. 1990). Hypertriglyceridemia leads to deteriorated transport of leptin through blood brain barrier (Banks et al. 2004), which can contribute to the peripheral leptin resistance.

3.2.7 Brain PUFA deficiency

The animal experiments showed that young individuals are more prone to EFA deficiency than the adults (Sardesai 1992). The western type of diet usually contains sufficient amounts of PUFA; therefore, the EFA deficiency in adults is limited to those being on total parenteral nutrition (TPN) (Richardson & Sgoutas 1975) that causes higher level of insulin, which slower the fat (EFA) mobilization from the adipose tissue depots (Sardesai 1992). The administration of fat-free TPN led after 3 months to alopecia, brittle nails, squamous dermatitis and increased susceptibility to infections (Richardson & Sgoutas 1975; Sardesai 1992). Deficiency of LCPUFA can also result from disturbed metabolic pathways transforming LA and ALA to more unsaturated PUFA n-6 and PUFA n-3 analogues (See Figure 1a). The deficiencies in D6 (Willard *et al.* 2001) and D5 (Moore *et al.* 1995) desaturases were described. The patients had signs of multineuronal degeneration and mental retardation. The indices of lacking D6 desaturase activity were also observed in patients with Sjögren-Larsson syndrome, genetic disorder with frequent progression to paraplegia and mental retardation (Hernell *et al.* 1982). Deficiency of n-6 and n-3 PUFA have different neurologic signs (Table 1)

The diet deficient in PUFA n-3 given to Rhesus monkeys and rats led to disturbed vision, abnormal electroretinogram and polydipsia. (Reisbick *et al.* 1990a; Reisbick *et al.* 1990b; Faldella *et al.* 1996). The EFA deficiency in humans is often connected with disturbed learning and behavioral skills (Stevens *et al.* 1996; Maes *et al.* 1996); the signs and symptoms of this deficiency depend on the extent and general nutritional status [simultaneous carency of vitamins and/or minerals (Siguel 1994)]. Siguel and Lerman (1996) proposed the term EFA insufficiency for the patients, who exhibit in the plasma FA profile the same type of abnormalities as the patients with EFA deficiency (high conversion of n-3, n-6 and n-9 FA to respective derivatives and accumulation of MUFA).

4. PUFA IN SELECTED PSYCHIATRIC DISEASES

<u>4.1 Schizophrenia</u>

The patients with schizophrenia have abnormal metabolism of PL, more specifically, lower concentrations of PS, PC and PE in cellular membranes as a consequence of PUFA (AA, EPA, and DHA) deficiency. The deficiency was proved in erythrocyte membranes and in post mortem brain specimens (Horrobin et al. 1991, Peet et al. 2004). The membrane PL are important for signal transduction via neurotransmitters, which is mediated by neuronal receptors; therefore, their abnormal metabolism is sometimes linked with the disturbed information handling in schizophrenics (Mahadik & Evans 2003). Some authors (Fukuzako et al. 1999; Keshavan et al. 2000), but not all (Puri et al. 2008), found using P³¹ magnetic resonance spectroscopy (MRS) low concentrations of phosphomonoesters and higher levels of phosphodiesters in temporal and frontal lobes in schizophrenic brains. This could point at higher intensity of PL metabolization and further corresponds with high concentrations of Ca++ independent PLA₂, the enzyme cleaving the FA from sn-2 position of PL, in blood and brains of schizophrenics (Ross et al. 1999). The decreased content of PUFA can result from higher oxidative stress or improper nutritional habits in schizophrenia. The arachidonic acid, after

Tab. 1. Neurologic characteristics of n-6 and n-3 EFA deficiency.

n-3	
Worsened learning ability	
Abnormal electroretinogram	
Blurred vision	
Polydipsia	
Decreased sensitivity	

being released from PL molecule, can be metabolized into many metabolites, some of them interfering with the homeostasis of neurotrasmitters, PI signalization cascade and neuromodulation by endocannabinoids (Yao & van Kammen 2004). Recently, the conceptualization has been introduced that schizophrenia similarly as depression (see below in paragraph 4.2.1.) can be viewed as a condition, in which the dysregulation of the immune system, oxidative and nitrosative stress (O&NS) as well as the tryptophan catabolite pathway play a role (Anderson *et al.* 2012). This novel concept, the neuroprogression, links neuroanatomical dysfunctions, lowered neurotrophic factors as well as a neurocognitive decline. The frame includes a possible role of PUFA n-3 being a part of the antioxidant system, which was used both in preventive and adjuvant therapy of schizophrenia (Pandya et al. 2012).

There were conducted many placebo-controlled studies which aimed at PUFA n-3 supplementation to the schizophrenic patients. However, the results are inconsistent, often comparable to the placebo group (Ross *et al.* 2007). In RCT lasting for 12 weeks and enrolling 76 individuals with subtreshold psychosis aged 13-25 years, the supplementation with PUFA n-3 (1.2 g/day) decreased the cumulative risk for the progression into manifesting psychosis by 22.6 % (p=0.007) (Ammringer *et al.* 2010).

4.2 Affective disorders

4.2.1. Depression

Major depression

It is now becoming evident that in major depression, progressive neuropathological processes are involved. Neuroprogression is a phase associated and potentially deteriorating process including the combination of impaired neuroplasticity, reduced neurogenesis coupled to increased apoptosis and neurodegeneration (Maes *et al.* 2012). The recurrent episodes of depression are connected with the decline of cognitive functions together with increased risk of Alzheimer disease. The probable factors involved include oxidative stress leading to modification of DNA and lipid peroxidation. The antidepressive treatment was shown to improve the parameters of oxidative stress (Maes *et al.* 2011b). Possible beneficial effects of PUFA n-3 in depression are

in link with the conception of neuroprogressive theory of depression (Moylan *et al.* 2012), as PUFA n-3 have the antiapoptotic, anti-inflammatory and antioxidative effects in CNS (Pandya *et al.* 2012).

Epidemiological data show that the fish consumption negatively correlates with the prevalence of depression in various populations (Hibbeln 1998). Lower levels of PUFA n-3 are connected with higher incidence of depression (Tiemeier et al. 2003). The depressive patients have lower content of PUFA n-3 in PL of various tissues - erythrocytes (Edwards et al. 1998, Peet et al. 1998), serum (Maes et al. 1996; Maes et al. 1998), adipose tissue (Mamalakis et al. 2006), and brain (McNamara et al. 2007) probably as a consequence of increased oxidative stress. Moreover, the concentrations of PUFA n-3 negatively correlate with the severity of depressive symptomatology (Conklin et al. 2007; Feart et al. 2008; Tiemeier et al. 2003). The lower content of PUFA n-3 is often accompanied with higher values of AA/EPA and AA/DHA ratios and/or PUFA n-6/n-3 ratio (Adams et al. 1996, Frasure-Smith et al. 2004). Restricted intake of PUFA n-3 in animals increases the depression and aggression scores (DeMar et al. 2006). Concomitantly, it was observed that the central serotonergic and dopaminergic transduction decreased together with lower number of hippocampal neurons. The dietary supplementation of DHA to rats brings about higher content of brain noradrenaline, serotonine, and acetylcholine and promotes neurite growth in hippocampal neurons (Calderon & Kim 2004; De Mar Jr. et al. 2006). The studies on healthy volunteers taking high PUFA n-3 doses proved (with the help of structural MR) increased amounts of gray matter in brain regions important for emotive and cognitive functions - the right hippocampus, the right amygdale and anterior cingulum (Conklin et al. 2007).

Recently, a new metaanalysis on PUFA n-3 in the treatment of depression was published (Appleton *et al.* 2010). The metaanalysis supports the hypothesis about the beneficial therapeutic effects of PUFA n-3, but it states that the results are difficult to summarize and evaluate due to considerable heterogeneity. Pronounced effect of PUFA n-3 was found in the patients with more severe symptomatology, whereas in the individuals without diagnosed depressive disorder, the effects on mood were weaker. The heterogeneity of the published results is probably caused by many factors, such as variable content of PUFA n-3, n-6 in the diet, placebo composition, age, gender, the accessory treatment, duration of the study, the dosage and type of PUFA n-3, or in the way the depression is evaluated (Appleton *et al.* 2010).

4.2.2 Bipolar affective disorder

It is supposed that PUFA n-3 play a role also in the bipolar disorder. Stoll and coll. compared in bipolar disorder the effect of olive oil (active placebo) and fish oil (equaling daily dosages of EPA 6.2 g/day and DHA 3.4 g/day). In the group taking fish oil, the authors

observed significant decrease in number of relapses of the disorder. PUFA n-3 improved the depressive, but not the manic signs of the disorder (Stoll et al. 1999). Similarly, Frangou et al. (2006) conducted double-blind, placebo controlled study in 74 patients with bipolar disorder. They found that 12-week administration of 1 or 2 g/day of EPA had significant effect on the depressive, but not manic symptoms, when both EPA groups were compared to the placebo group. PUFA n-3 were also tried as the treatment of bipolar disorder in children and adolescents. In juvenile bipolar disorder, the dosage of EPA+DHA only slightly alleviated manic symptoms (Wozniak et al. 2007). Recent metaanalysis including only randomized designs lasting ≥ 4 weeks found that adjunctive administration of PUFA n-3 significantly corrected depressive symptoms without an effect on manic ones (Sarris et al. 2011).

4.2.3 Postpartum depression

Postpartum depression is symptomatic depressive disorder with the same symptoms as major depressive disorder or depressive disorder in a part of bipolar affective disorder. The populations with higher fish consumption have higher concentrations of DHA in breast milk and lower incidence of postpartum depressive disorder, whereas low fish intake during pregnancy is connected with higher incidence of depression (Golding et al. 2009). The hypothesis about the deficiency of PUFA n-3 in women with postpartum depression was supported by some authors (De Vriese et al. 2003), but later study did not find lower PUFA n-3 (Browne et al. 2009). During pregnancy, the PUFA n-3 are transferred into the fetus (the biomagnification effect), thus the mother is under higher risk of PUFA n-3 deficiency (Holman et al. 1991). The supplementation of PUFA n-3 to pregnant women was proposed to prevent from this type of deficiency, but newly published metaanalysis (Jans et al. 2010), with 309 women supplemented with PUFA n-3 and 303 women on placebo, did not prove the beneficial effect of the supplementation.

4.3 Alzheimer's disease

Epidemiological data indicate that the sufficient dietary intake of PUFA n-3 (DHA) is associated with a significant reduction in dementia risk (Johnson & Schaefer 2006). One prospective study found that the individuals with the upper quartile concentration of DHA in plasma PC had (in comparison with those in the lower quartile) during 9 year follow-up significantly lower risk for AD (Schaefer et al. 2006). The patients with sporadic form of AD have lower content of protein SorLA (LR11) and its mRNA. This protein decreases the cleavage of amyloid precursor protein (APP) by α - and β -secretases, henceforward the formation of β -amyloid. The administration of DHA to transgenic mice caused higher production of LR11 in neurons and lower production of β-amyloid (Ma et al. 2007). The influence of PUFA n-3 on the oxidative stress and inflammation, both of which

playing an important role in the pathogenesis of AD, can be a part of the beneficial effect of PUFA n-3 in the prevention of AD. DHA and its derivative, neuroprotectin D1, enhance antioxidant defense of neurons by induction of antioxidant enzymes (Hashimoto *et al.* 2002) and antiapoptotic proteins (Jicha and Markesbery 2010). Moreover, DHA attenuates the production of toxic oligomers A beta (Hashimoto *et al.* 2008), which damage the synapses. PUFA n-3 support the formation of BDNF (*brain-derived neurotrophic factor*), that is important for neurogenesis and synaptic neuronal plasticity, both being impaired in AD (Bousquet *et al.* 2009).

PUFA n-3 deficiency is involved in the progression of other dementias and neurodegenerative diseases (Kidd 2008). In vascular dementia, the antiatherogenic and antithrombogenic effects play a role. It is supposed that PUFA n-3 act through their antiinflammatory and neuroprotective properties, as well as the ability to decrease the toxicity of α -synuclein (degenerative neuronal protein). Other protective effects include higher production of endogenous cannabinoids, the specific class of PUFA metabolites. Endogenous cannabinoids modulate dopaminergic activity in basal ganglia (Fernández-Ruiz *et al.* 2010).

Supplementation of PUFA n-3 to the patients with AD improves cognitive performance probably only in the subgroup with mild cognitive dysfunction (equaling to so-called amnestic form of mild disorder in cognitive functions, usually considered as a preclinical form of AD) (Freund-Levi *et al.* 2008). The reduction of risk for the development of AD was described in several studies (Barberger-Gateau *et al.* 2007). It seems that apoE is involved, because the beneficial effects of PUFA n-3 were not observed in the carriers of the e4 allele (Barberger-Gateau *et al.* 2007).

4.4 Other psychiatric diseases

4.4.1 Attention deficit hyperactivity disorder (ADHD)

Attention deficit hyperactivity disorder is very common disorder with onset in childhood, which inflicts 5-7% of children on average; the disorder often fades away, but in many cases, it persists to adolescent age and in 40–60% cases, it is still diagnosed in adults. Prevalence of ADHD in adults reaches 3-5% (Drtílková & Theiner 2008). About one quarter of ADHD patients suffers also from learning disorders, especially from reading and writing ones (Doggett 2004). It is supposed that the etiopathogenesis of ADHD includes both genetic and extrinsic factors. Both dopaminergic and noradrenergic dysfunctions were described - decreased transmission in the mesolimbic and frontal cortex areas. Some signs of ADHD can be accentuated with the PUFA deficiency, which often results from improper dietary habits (Antalis et al. 2006). Colter et al. (2008) found in adolescents with ADHD higher energy intake in comparison with the control group, but there was no difference in PUFA n-3 and n-6 intake; in the erythrocyte PL, the ADHD group had lower content of DHA, PUFA n-3 and lower PUFA n-3/n-6 ratio. These changes correlated with severity of ADHD symptoms evaluated with the Conner scale (Colter et al. 2008). Sorgi and coll. treated the children with ADHD, for 8 weeks with high dose of EPA + DHA (16.2 g/day from the beginning, in the 4th week, the dosage was titrated to reach the AA/EPA ratio in plasma PL similar to Japanese population). The observed findings included improved behavior of the patients and significant positive correlation between the decrease in AA/EPA ratio and global decrease in burden of disorder (Sorgi et al. 2007). In the new metaanalysis of the studies, it was found that the supplementation with PUFA n-3, especially with high doses of EPA, is modestly effective in the treatment of ADHD (Bloch & Quawasmi 2011).

4.4.2 Chronic fatigue syndrome

Myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS), medical disorder or group of disorders (Sanders 2008), characterized by persistent fatigue accompanied by at least four of the following symptoms: substantial impairment in short - term memory or concentration; sore throat; tender cervical and axillary lymph nodes; muscle pain; multi - joint pain without selling or redness; headache of new type; unrefreshing sleep; and post exertion malaise lasting more than 24 hours for a minimum of six months in adults. Inflammation and oxidative and nitrosative pathways and a lowered antioxidant status are important pathophysiological mechanisms underpinning ME/CFS (Maes et al. 2009). Chronic fatigue syndrome shows a high degree of comorbidity with depression; many CFS patients develop depression during their illness, while fatigue is one of the key symptoms of major depression (Maes 2005). Some studies reported finding reduced levels of PUFA n-6 or PUFA n-3 in cell membranes or serum in patients diagnosed with CFS (Liu et al. 2003), while Warren (Warren et al. 1999) did not find any differences in red blood cell membrane lipids. Treatment of patients with CFS solely with a high-EPA containing supplement led to an improvement in their symptomatology within eight to 12 weeks (Puri et al. 2004).

ACKNOWLEDGEMENTS

The study was supported by Research project of Charles University in Prague, 1st Faculty of Medicine – PRVOUK-P25/LF1/2, the grant NT/13199-4, IGA Ministry of Health, The Czech Republic and Research Project MSMT 0021620849.

REFERENCES

- Abdolmaleky HM, Cheng KH, Faraone SV, Wilcox M, Glatt SJ, Gao F, Smith CL, Shafa R, Aeali B, Carnevale J, Pan H, Papageorgis P, Ponte JF, Sivaraman V, Tsuang MT, Thiagalingam S (2006). Hypomethylation of MB-COMT promoter is a major risk factor for schizophrenia and bipolar disorder. Hum Mol Genet. **15**(21): 3132–45.
- 2 Adams PB, Lawson S, Sanigorski A, Sinclair AJ (1996). Arachidonic acid to eicosapentaenoic acid ratio in blood correlates positively with clinical symptoms of depression. Lipids **31** Suppl: S157–61
- 3 Amminger GP, Schäfer MR, Papageorgiou K, Klier CM, Cotton SM, Harrigan SM, Mackinnon A, McGorry PD, Berger GE (2010). Long-chain omega-3 fatty acids for indicated prevention of psychotic disorders: a randomized, placebo-controlled trial. Arch Gen Psychiatry 67(2): 146–54.
- 4 Anderson G, Maes M (2012). Schizophrenia: Linking prenatal infection to cytokines, the tryptophan catabolite (TRYCAT) pathway, NMDA receptor hypofunction, neurodevelopment and neuroprogression. Prog Neuropsychopharmacol Biol Psychiatry. http://dx.doi.org/10.1016/j.pnpbp.2012.07.016.
- 5 Antalis CJ, Stevens LJ, Campbell M, Pazdro R, Ericson K, Burgess JR (2006). Omega-3 fatty acid status in attention-deficit/hyperactivity disorder. Prostaglandins Leukot Essent Fatty Acids. 75(4–5): 299–308.
- 6 Appleton KM, Rogers PJ, Ness AR (2010).Updated systematic review and meta-analysis of the effects of n-3 long-chain polyunsaturated fatty acids on depressed mood. Am J Clin Nutr 91(3): 757-70.
- 7 Arterburn LM, Hall EB, Oken H (2006). Distribution, interconversion, and dose response of n-3 fatty acids in humans. Am J Clin Nutr. 83(6 Suppl): 1467S–1476S
- 8 Banks WA, Coon AB, Robinson SM, Moinuddin A, Shultz JM, Nakaoke R, Morley JE (2004). Triglycerides induce leptin resistance at the blood-brain barrier. Diabetes 53(5): 1253–60.
- 9 Barberger-Gateau P, Raffaitin C, Letenneur L, Berr C, Tzourio C, Dartigues JF, Alpérovitch A (2007). Dietary patterns and risk of dementia: the Three-City cohort study. Neurology. 69(20): 1921–30.
- 10 Bazan NG (2009). Neuroprotectin D1-mediated anti-inflammatory and survival signaling in stroke, retinal degenerations, and Alzheimer's disease. J Lipid Res 50: S400–S405.
- 11 Bazan NG, Colangelo V, Lukiw WJ (2002). Prostaglandins and other lipid mediators in Alzheimer's disease. Prostaglandins Other Lipid Mediat **68–69**: 197–210.
- 12 Bloch MH, Qawasmi A (2011). Omega-3 fatty acid supplementation for the treatment of children with attention-deficit/ hyperactivity disorder symptomatology: systematic review and meta-analysis. J Am Acad Child Adolesc Psychiatry **50**(10): 991–1000.
- 13 Bosetti F (2007). Arachidonic acid metabolism in brain physiology and pathology: lessons from genetically altered mouse models. J Neurochem. **102**(3): 577–86.
- 14 Bottiglieri T, Laundy M, Crellin R, Toone BK, Carney MW, Reynolds EH (2000). Homocysteine, folate, methylation, and monoamine metabolism in depression. J Neurol Neurosurg Psychiatry. 69(2): 228–32
- 15 Bourre JM, Dumont O, Piciotti M, Clément M, Chaudière J, Bonneil M, Nalbone G, Lafont H, Pascal G, Durand G (1991). Essentiality of omega 3 fatty acids for brain structure and function. World Rev Nutr Diet 66: 103–17.
- 16 Bousquet M, Gibrat C, Saint-Pierre M, Julien C, Calon F, Cicchetti F (2009). Modulation of brain-derived neurotrophic factor as a potential neuroprotective mechanism of action of omega-3 fatty acids in a parkinsonian animal model. Prog Neuropsychopharmacol Biol Psychiatry. **33**(8): 1401–8.
- 17 Browne JC, Scott KM, Silvers KM (2006). Fish consumption in pregnancy and omega-3 status after birth are not associated with postnatal depression. J Affect Disord **90**(2–3): 131–9. Epub 2005 Dec 1.
- 18 Calderon F, Kim HY (2004). Docosahexaenoic acid promotes neurite growth in hippocampal neurons J Neurochem 90(4): 979–88.

- 19 Carver JD, Benford VJ, Han B, Cantor AB (2001). The relationship between age and the fatty acid composition of cerebral cortex and erythrocytes in human subjects. Brain Res Bull 56(2): 79–85.
- 20 Catalá A (2010). The function of very long chain polyunsaturated fatty acids in the pineal gland. Biochim Biophys Acta. **1801**(2): 95–9.
- 21 Colter AL, Cutler C, Meckling KA (2008). Fatty acid status and behavioural symptoms of attention deficit hyperactivity disorder in adolescents: a case-control study. Nutr J. **14**;7: 8.
- 22 Conklin SM, Gianaros PJ, Brown SM, Yao JK, Hariri AR, Manuck SB, Muldoon MF (2007). Long-chain omega-3 fatty acid intake is associated positively with corticolimbic gray matter volume in healthy adults. Neurosci Lett. **421**(3): 209–12.
- 23 Conklin SM, Manuck SB, Yao JK, Flory JD, Hibbeln JR, Muldoon MF (2007). High omega-6 and low omega-3 fatty acids are associated with depressive symptoms and neuroticism. Psychosom Med. 69(9): 932–4.
- 24 Cysneiros RM, Ferrari D, Arida RM, Terra VC, de Almeida AC, Cavalheiro EA, Scorza FA (2010). Qualitative analysis of hippocampal plastic changes in rats with epilepsy supplemented with oral omega-3 fatty acids. Epilepsy Behav **17**(1): 33–8
- 25 De Vriese SR, Christophe AB, Maes M (2003). Lowered serum n-3 polyunsaturated fatty acid (PUFA) levels predict the occurrence of postpartum depression: further evidence that lowered n-PUFAs are related to major depression. Life Sci **73**(25): 3181–7.
- 26 Deckelbaum RJ, Worgall TS, Seo T (2006). n-3 fatty acids and gene expression. Am J Clin Nutr **83**(6 Suppl): 15205–15255
- 27 Delion S, Chalon S, Hérault J, Guilloteau D, Besnard JC, Durand G (1994). Chronic dietary alpha-linolenic acid deficiency alters dopaminergic and serotoninergic neurotransmission in rats. J Nutr **124**: 2466–76.
- 28 DeMar JC Jr, Ma K, Bell JM, Igarashi M, Greenstein D, Rapoport SI (2006). One generation of n-3 polyunsaturated fatty acid deprivation increases depression and aggression test scores in rats. J Lipid Res. **47**(1): 172–80. Epub 2005 Oct 6
- 29 Dittmann S, Seemüller F, Schwarz MJ, Kleindienst N, Stampfer R, Zach J, Born C, Bernhard B, Fast K, Grunze H, Engel RR, Severus E (2007). Association of cognitive deficits with elevated homocysteine levels in euthymic bipolar patients and its impact on psychosocial functioning: preliminary results. Bipolar Disord 9(1–2): 63–70.
- 30 Doggett AM (2004). ADHD and drug therapy: is it still a valid treatment? J Child Health Care **8**: 69–81.
- 31 Drtílková I., Theiner P (2008). Clinical and Biological Markers of Persistent Form of Hyperkinetic Disorder (ADHD). Čes. a slov. Psychiat. **104**: 167–171.(In Czech)
- 32 Edwards R, Peet M, Shay J, Horrobin D (1998). Omega-3 polyunsaturated fatty acid levels in the diet and in red blood cell membranes of depressed patients. J Affect Disord **48**(2–3): 149–55.
- 33 Ekdahl CT, Claasen JH, Bonde S, Kokaia Z, Lindvall O (2003). Inflammation is detrimental for neurogenesis in adult brain. Proc Natl Acad Sci U S A **100**(23): 13632–7.
- 34 Faldella G, Govoni M, Alessandroni R, Marchiani E, Salvioli GP, Biagi PL, Spano C (1996). Visual evoked potentials and dietary long chain polyunsaturated fatty acids in preterm infants. Arch Dis Child **75**: F108–F112.
- 35 Féart C, Peuchant E, Letenneur L, Samieri C, Montagnier D, Fourrier-Reglat A, Barberger-Gateau P (2008). Plasma eicosapentaenoic acid is inversely associated with severity of depressive symptomatology in the elderly: data from the Bordeaux sample of the Three-City Study. Am J Clin Nutr. 87(5): 1156–62.
- 36 Fernandez-Ruiz J, Hernández M, Ramos JA (2010) Cannabinoid– Dopamine Interaction in the Pathophysiology and Treatment of CNS Disorders. CNS Neuroscience & Therapeutics 16 : e72–e91
- 37 Fitzpatrick FA, Soberman R (2001). Regulated formation of eicosanoids J Clin Invest **107**: 1347–1351.
- 38 Flachs P, Mohamed-Ali V, Horakova O, Rossmeisl M, Hosseinzadeh-Attar MJ, Hensler M, Ruzickova J, Kopecky J (2006). Polyunsaturated fatty acids of marine origin induce adiponectin in mice fed a high-fat diet. Diabetologia. 49(2): 394–7.

- 39 Folstein M, Liu T, Peter I, Buell J, Arsenault L, Scott T, Qiu WW (2007). The homocysteine hypothesis of depression. Am J Psychiatry. **164**(6): 861–7.
- 40 Fornaro M, Prestia D, Colicchio S, Perugi G (2010). A Systematic, Updated Review on the Antidepressant Agomelatine Focusing on its Melatonergic Modulation. Current Neuropharmacology 8: 287–304.
- 41 Frangou S, Lewis M, McCrone P (2006). Efficacy of ethyl-eicosapentaenoic acid in bipolar depression: randomised doubleblind placebo-controlled study. Br J Psychiatry **188**: 46–50.
- 42 Frasure-Smith N, Lespérance F, Julien P (2004). Major depression is associated with lower omega-3 fatty acid levels in patients with recent acute coronary syndromes. Biol Psychiatry 55(9): 891–6.
- 43 Freund-Levi Y, Basun H, Cederholm T, Faxén-Irving G, Garlind A, Grut M, Vedin I, Palmblad J, Wahlund LO, Eriksdotter-Jönhagen M (2008). Omega-3 supplementation in mild to moderate Alzheimer's disease: effects on neuropsychiatric symptoms. Int J Geriatr Psychiatry. 23(2): 161–9.
- 44 Fukuzako H, Fukuzako T, Hashiguchi T, Kodama S, Takigawa M, Fujimoto T (1999). Changes in levels of phosphorus metabolites in temporal lobes of drug-naive schizophrenic patiens. Am J Psychiatry. **156**(8): 1205–8.
- 45 Gerbi A, Maixent JM, Barbey O, Jamme I, Pierlovisi M, Coste T, et al (1998). Alterations of Na, K-ATPase isoenzymes in the rat diabetic neuropathy: protective effect of dietary supplementation with n-3 fatty acids. J Neurochem **71**: 732–740.
- 46 Golding J, Steer C, Emmett P, Davis JM, Hibbeln JR (2009). High levels of depressive symptoms in pregnancy with low omega-3 fatty acid intake from fis. Epidemiology. **20**(4): 598–603.
- 47 Hankey GJ, Eikelboom JW (1999). Homocysteine and vascular disease. Lancet **354**: 407–13.
- 48 Harris WS (1989). Fish oils and plasma lipid and lipoprotein metabolism in humans: a critical review. J Lipid Res. 30(6): 785–807.
- 49 Hashimoto M, Hossain S, Shimada T, Sugioka K, Yamasaki H, Fujii Y, Ishibashi Y, Oka J, Shido O (2002). Docosahexaenoic acid provides protection from impairment of learning ability in Alzheimer's disease model rats. J Neurochem. 81(5): 1084–91.
- 50 Hashimoto M, Shahdat HM, Yamashita S, Katakura M, Tanabe Y, Fujiwara H, Gamoh S, Miyazawa T, Arai H, Shimada T, Shido O (2008). Docosahexaenoic acid disrupts in vitro amyloid beta(1-40) fibrillation and concomitantly inhibits amyloid levels in cerebral cortex of Alzheimer's disease model rats. J Neurochem. **107**(6): 1634–46.
- 51 Henderson DC, Copeland PM, Nguyen DD, Borba CP, Cather C, Eden Evins A, Freudenreich O, Baer L, Goff DC (2006). Homocysteine levels and glucose metabolism in non-obese, nondiabetic chronic schizophrenia. Acta Psychiatr Scand **113**(2): 121–125.
- 52 Hernell O, Holmgren G, Jagell SF, Johnson SB, Holman RT (1982). Suspected faulty essential fatty acid metabolism in Sjogren-Larsson syndrome. Pediatr Res **16**: 45–49.
- 53 Hibbeln JR (1998). Fish consumption and major depression. Lancet **351**: 1213.
- 54 Hichami A, Morin C, Rousseau E, Khan NA (2005).: Diacylglycerol-Containing Docosahexaenoic Acid in Acyl Chain Modulates Airway Smooth Muscle Tone. Amer J Resp Cell Mol Biol 33: 378–386.
- 55 Holman RT, Johnson SB, Ogburn PL (1991). Deficiency of essential fatty acids and membrane fluidity during pregnancy and lactation. Proc Natl Acad Sci U S A. **88**(11): 4835–9.
- 56 Horrobin DF, Manku MS, Hillman H, Iain A, Glen M. Fatty acid levels in the brains of schizophrenics and normal controls. Biol Psychiatry. 1991 Oct 15;30(8): 795–805.
- 57 Huang T, Wahlqvist ML, Li D (2012). Effect of n-3 polyunsaturated fatty acid on gene expression of the critical enzymes involved in homocysteine metabolism. Nutr J. **11**(1): 6. [Epub ahead of print]
- 58 Chahl LA (2011). TRP channels and psychiatric disorders. Adv Exp Med Biol **704**: 987–1009.

- 59 Chalon S (2006). Omega-3 fatty acids and monoamine neurotransmission. Prostaglandins Leukot Essent Fatty Acids. **75**(4–5): 259–69.
- 60 Charnay Y, Cusin I, Vallet PG, Muzzin P, Rohner-Jeanrenaud F, Bouras C (2000). Intracerebroventricular infusion of leptin decreases serotonin transporter binding sites in the frontal cortex of the rat. Neurosci Lett **283**(2): 89–92.
- 61 Jans LA, Giltay EJ, Van der Does AJ (2010). The efficacy of n-3 fatty acids DHA and EPA (fish oil) for perinatal depression. Br J Nutr **104**(11): 1577–85.
- 62 Jicha GA, Markesbery WR (2010). Omega-3 fatty acids: potential role in the management of early Alzheimer's disease. Clin Interv Aging. **5**: 45–61.
- 63 Johnson EJ, Schaefer EJ (2006). Potential role of dietary n-3 fatty acids in the prevention of dementia and macular degeneration. Am J Clin Nutr. **83**(6 Suppl): 14945–14985.
- 64 Kearns SD, Haag M (2002). The effect of omega-3 fatty acids on Ca-ATPase in rat cerebral cortex. Prostaglandins Leukot Essent Fatty Acids **67**: 303–8.
- 65 Kennedy BP, Bottiglieri T, Arning E, Ziegler MG, Hansen LA, Masliah E (2004). Elevated S-adenosylhomocysteine in Alzheimer brain: influence on methyltransferases and cognitive function. J Neural Transm. **111**(4): 547–67.
- 66 Keshavan MS, Stanley JA, Pettegrew JW (2000). Magnetic resonance spectroscopy in schizophrenia: methodological issues and findings--part II. Biol Psychiatry **48**(5): 369–80.
- 67 Kidd PM (2008). Alzheimer's disease, amnestic mild cognitive impairment, and age-associated memory impairment: current understanding and progress toward integrative prevention. Altern Med Rev **13**(2): 85-115.
- 68 Kitajka K, Puskás LG, Zvara A, Hackler L Jr, Barceló-Coblijn G, Yeo YK, Farkas T (2002). The role of n-3 polyunsaturated fatty acids in brain: modulation of rat brain gene expression by dietary n-3 fatty acids. Proc Natl Acad Sci U S A. **99**(5): 2619–24.
- 69 Kodydková J, Vávrová L, Zeman M, Jirák R, Macášek J, Stanková B, Tvrzická E, Žák A (2009). Antioxidative enzymes and increased oxidative stress in depressive women. Clin Biochem. 42(13–14): 1368–74.
- 70 Leonelli M, Graciano MFR. Britto LRG (2011). TRP channels, omega-3 fatty acids, and oxidative stress in neurodegeneration: from the cell membrane to intracellular cross-links. Braz J Med Biol Res 44(11): 1088–1096.
- 71 Li D, Yu XM, Xie HB, Zhang YH, Wang Q, Zhou XQ, Yu P, Wang LJ (2007). Platelet phospholipid n-3 PUFA negatively associated with plasma homocysteine in middle-aged and geriatric hyperlipaemia patients. Prostaglandins Leukot Essent Fatty Acids. 76(5): 293–7.
- 72 Liu Z, Wang D, Xue Q, Chen J, Li Y, Bai X, Chang L (2003). Determination of fatty acid levels in erythrocyte membranes of patients with chronic fatigue syndrome. Nutritional neuroscience 6: 389–92.
- 73 Lorente-Cebrián S, Pérez-Matute P, Martínez JA, Marti A, Moreno-Aliaga MJ (2006). Effects of eicosapentaenoic acid (EPA) on adiponectin gene expression and secretion in primary cultured rat adipocytes. J Physiol Biochem **62**(2): 61–9.
- 74 Lu XY (2007). The leptin hypothesis of depression: a potential link between mood disorders and obesity? Curr Opin Pharmacol. 7(6): 648–52.
- 75 Ma QL, Teter B, Ubeda OJ, Morihara T, Dhoot D, Nyby MD, Tuck ML, Frautschy SA, Cole GM (2007). Omega-3 fatty acid docosahexaenoic acid increases SorLA/LR11, a sorting protein with reduced expression in sporadic Alzheimer's disease (AD): relevance to AD prevention. J Neurosci. 27(52): 14299–307.
- 76 Maes M (2009). Inflammatory and oxidative and nitrosative stress pathways underpinning chronic fatigue, somatization and psychosomatic symptoms. Curr Opin Psychiatry 22: 75–83.
- 77 Maes M, Fišar Z, Medina M, Scapagnini G, Nowak G, Berk M (2012). New drug targets in depression: inflammatory, cellmediated immune, oxidative and nitrosative stress, mitochondrial, antioxidant, and neuroprogressive pathways. And new drug candidates-Nrf2 activators and GSK-3 inhibitors Inflammopharmacol **20**: 127–150.

- 78 Maes M, Galecki P, Chang YS, Berk M (2011b). A review on the oxidative and nitrosative stress (O&NS) pathways in major depression and their possible contribution to the (neuro) degenerative processes in that illness. Prog Neuropsychopharmacol Biol Psychiatry **35**: 676–692.
- 79 Maes M, Christophe A, Delanghe J, Altamura C, Neels H, Meltzer HY (1998). Lowered omega3 polyunsaturated fatty acids in serum phospholipids and cholesterol esters of depressed patients. Psychiatry Res. 85(3): 275–91.
- 80 Maes M, Kubera M, Obuchowiczwa E, Goehler L, Brzeszcz J (2011a). Depression's multiple comorbidities explained by (neuro)inflammatory and oxidative & nitrosative stress pathways. Neuro Endocrinol Lett **32**(1): 7–24.
- 81 Maes M, Mihaylova I, Leunis JC (2005). In chronic fatigue syndrome, the decreased levels of omega-3 poly-unsaturated fatty acids are related to lowered serum zinc and defects in T cell activation. Neuro Endocrinol. Lett. **26**: 745–51.
- 82 Maes M, Smith R, Christophe A, Cosyns P, Desnyder R, Meltzer H (1996). Fatty acid composition in major depression: Decreased omega 3 fractions in cholesteryl esters and increased C20: 4 omega 6/C20: 5 omega 3 ratio in cholesteryl esters and phospholipids. J Affective Disord **38**: 35–46.
- 83 Maes M, Smith RS (1998). Fatty acids, cytokines, and major depression. Biol Psychiatry **43**(5): 313–4.
- 84 Mahadik SP, Evans DR (2003). Is schizophrenia a metabolic brain disorder? Membrane phospholipid dysregulation and its therapeutic implications. Psychiatr Clin N Am 26: 85–102
- 85 Mamalakis G, Kalogeropoulos N, Andrikopoulos N, Hatzis C, Kromhout D, Moschandreas J, Kafatos A (2006). Depression and long chain n-3 fatty acids in adipose tissue in adults from Crete. Eur J Clin Nutr **60**(7): 882–888.
- 86 Marszalek JR, Lodish HF (2005). Docosahexaenoic acid, fatty acid-interacting proteins, and neuronal function: breastmilk and fish are good for you. Annu Rev Cell Dev Biol **21**: 633–57.
- 87 McNamara RK, Carlson SE (2006). Role of omega-3 fatty acids in brain development and function: potential implications for the pathogenesis and prevention of psychopatology Prostaglandins Leukot Essent Fatty Acids **75**(4–5): 329–49.
- 88 McNamara RK, Hahn CG, Jandacek R, Rider T, Tso P, Stanford KE, Richtand NM (2007). Selective deficits in the omega-3 fatty acid docosahexaenoic acid in the postmortem orbitofrontal cortex of patients with major depressive disorder. Biol Psychiatry. 62(1): 17–24.
- 89 McNamara RK, Jandacek R (2011). Investigation of postmortem brain polyunsaturated fatty acid composition in psychiatric disorders: limitations, challenges, and future directions. J Psychiatr Res **45**(1): 44–6.
- 90 McNamara RK, Jandacek R, Rider T, Tso P, Cole-Strauss A, Lipton JW (2010). Omega-3 fatty acid deficiency increases constitutive pro-inflammatory cytokine production in rats: relationship with central serotonin turnover. Prostaglandins Leukot Essent Fatty Acids. **83**(4–6): 185–91.
- 91 Miller AL (2008). The methylation, neurotransmitter, and antioxidant connections between folate and depression. Altern Med Rev. **13**(3): 216–26.
- 92 Minami M, Kimura S, Endo T, Hamaue N, Hirafuji M, Togashi H, Matsumoto M, Yoshioka M, Saito H, Watanabe S, Kobayashi T, Okuyama H (1997). Dietary docosahexaenoic acid increases cerebral acetylcholine levels and improves passive avoidance performance in stroke-prone spontaneously hypertensive rats. Pharmacol Biochem Behav 58: 1123–1129.
- 93 Moore SA, Hurt E, Yoder E, Sprecher H, Spector AA (1995). Docosahexaenoic acid synthesis in human skin fibroblasts involves peroxisomal retroconversion of tetracosahexaenoic acid. J Lipid Res 36: 2433–2443.
- 94 Mourek J, Langmeier M, Pokorny J (2009). Significance of the plasma membrane for the nerve cell function, development and plasticity. Neuro Endocrinol Lett **30**(6): 694–699.
- 95 Moylan S, Maes M, Wray NR, Berk M (2012). The neuroprogressive nature of major depressive disorder: pathways to disease evolution and resistance, and therapeutic implications. Mol Psychiatry. 2012 Apr 24. doi: 10.1038/mp.2012.33.

- 96 Murphy MG (1990). Dietary fatty acids and membrane protein function. J. Nutr. Biochem **1**: 68–79.
- 97 Narayanan BA, Narayanan NK, Reddy BS (2001). Docosahexaenoic acid regulated genes and transcription factors inducing apoptosis in human colon cancer cells. Int J Oncol **19**(6): 1255–62.
- 98 Pandya CD, Howell KR, Pillai A (2012). Antioxidants as potential therapeutics for neuropsychiatric disorders. Prog Neuropsychopharmacol Biol Psychiatry. http: //dx.doi.org/10.1016/j. pnpbp.2012.07.017
- 99 Paz-Filho G, Wong ML, Licinio J (2010). The procognitive effects of leptin in the brain and their clinical implications. Int J Clin Pract **64**(13): 1808–12.
- 100 Peet M, Murphy B, Shay J, Horrobin D (1998). Depletion of omega-3 fatty acid levels in red blood cell membranes of depressive patients. Biol Psychiatry; 43(5): 315–9.
- 101 Peet M, Shah S, Selvam K., Ramchand CN (2004). Polyunsaturated fatty acid levels in red cell membranes of unmedicated schizophrenic patiens. World J Biol Psychiatry **5**: 92–99.
- 102 Peyron-Caso E, Taverna M, Guerre-Millo M, Veronese A, Pacher N, Slama G, Rizkalla S (2002). Dietary (n-3) polyunsaturated fatty acids upregulate plasma leptin in insulin-resistant rats J Nutr 132: 2235–2240.
- 103 Pike LJ (2004). Lipid rafts: heterogeneity on the high seas. Biochem J **384**: 281–292.
- 104 Puri BK (2004). The use of eicosapentaenoic acid in the treatment of chronic fatigue syndrome. Prostaglandins Leukot Essent Fatty Acids **70**: 399–401.
- 105 Puri BK, Counsell SJ, Hamilton G (2008). Brain cell membrane motion-restricted phospholipids: a cerebral 31-phosphorus magnetic resonance spectroscopy study of patients with schizophrenia. Prostaglandins Leukot Essent Fatty Acids. **79**(6): 233–5.
- 106 Rabaneda LG, Carrasco M, López-Toledano MA, Murillo-Carretero M, Ruiz FA, Estrada C, Castro C (2008). Homocysteine inhibits proliferation of neuronal precursors in the mouse adult brain by impairing the bFGF signaling cascade and reducing Erk1/2-dependent cyclin E expression. FASEB J **22**: 3823–3835.
- 107 Ravaglia G, Forti P, Maioli F, Muscari A, Sacchetti L, Arnone G, Nativio V, Talerico T, Mariani E (2003). Homocysteine and cognitive function in healthy elderly community dwellers in Italy. Am. J. Clin. Nutr 77: 668–673.
- 108 Reddy KS, Katan MB (2004). Diet, nutrition and the prevention of hypertension and cardiovascular diseases. Public Health Nutr. **7**(1A): 167–86.
- 109 Reisbick S, Neuringer M, Connor W, Iliff-Sizemore S (1990b). Increased intake of water and NaCl solutions in omega-3 fatty acid deficient monkeys. Physiol Behav **49**: 1139–1146.
- 110 Reisbick S, Neuringer M, Hasnain R, Connor W (1990a). Polydipsia in rhesus monkeys deficient in omega-3 fatty acids. Physiol Behav 47: 315–323.
- 111 Reiter RJ, Tan DX, Jou MJ, Korkmaz A, Manchester LC, Paredes SD (2008). Biogenic amines in the reduction of oxidative stress: melatonin and its metabolites. Neuro Endocrinol Lett. **29**(4): 391–8.
- 112 Richardson TJ, Sgoutas D (1975). Essential fatty acid deficiency in four adult patients during total parenteral nutrition. Am J Clin Nutr **28**: 258–263.
- 113 Ross BM, Seguin J, Sieswerda LE (2007). Omega-3 fatty acids as treatments for mental illness: which disorder and which fatty acid? Lipids in Health and Disease **6**: 21 doi: 10.1186/1476-511X-6-21
- 114 Ross BM, Turenne S, Moszczynska A, Warsh JJ, Kish SJ (1999). Differential alteration of phospholipase A2 activities in brain of patients with schizophrenia. Brain Res 821(2): 407–13.
- 115 Rossi AS, Lombardo YB, Lacorte JM, Chicco AG, Rouault C, Slama G, Rizkalla SW (2005). Dietary fish oil positively regulates plasma leptin and adiponectin levels in sucrose-fed, insulinresistant rats. Am J Physiol Regul Integr Comp Physiol **289**(2): R486–R494.
- 116 Sanders P, Korf J (2008). Neuroaetiology of chronic fatigue syndrome: an overview. World J Biol. Psychiatry **9**: 165–71

- 117 Sardesai VM (1992). The essential fatty acids. Nutr Clin Pract 7: 179–186.
- 118 Sarris J, Mischoulon D, Schweitzer I (2011). Adjunctive nutraceuticals with standard pharmacotherapies in bipolar disorder: a systematic review of clinical trials. Bipolar Disord. **13**(5–6): 454–65. doi: 10.1111/j.1399-5618.2011.00945.x.
- 119 Serhan CN, Krishnamoorthy S, Recchiuti A, Chiang N (2011). Novel Anti-Inflammatory -- Pro-Resolving Mediators and Their Receptors Curr Top Med Chem **11**(6): 629–647.
- 120 Shahbakhti H, Watson RE, Azurdia RM, Ferreira CZ, Garmyn M, Rhodes LE (2004). Influence of eicosapentaenoic acid, an omega-3 fatty acid, on ultraviolet-B generation of prostaglandin-E2 and proinflammatory cytokines interleukin-1 beta, tumor necrosis factor-alpha, interleukin-6 and interleukin-8 in human skin in vivo. Photochem Photobiol **80**(2): 231–5.
- 121 Shimazawa M, Nakajima Y, Mashima Y, Hara H (2009). Docosahexaenoic acid (DHA) has neuroprotective effects against oxidative stress in retinal ganglion cells. Brain Res **1251**: 269–75
- 122 Schaefer EJ, Bongard V, Beiser AS, Lamon-Fava S, Robins SJ, Au R, Tucker KL, Kyle DJ, Wilson PW, Wolf PA (2006). Plasma phosphatidylcholine docosahexaenoic acid content and risk of dementia and Alzheimer disease: the Framingham Heart Study. Arch Neurol 63(11): 1545–50.
- 123 Signorini C, Ciccoli L, Leoncini S, Carloni S, Perrone S, Comporti M, Balduini W, Buonocore G (2009).Free iron, total F-isoprostanes and total F-neuroprostanes in a model of neonatal hypoxic-ischemic encephalopathy: neuroprotective effect of melatonin. J Pineal Res **46**(2): 148–54.
- 124 Siguel E (1994). Essential Fatty Acids in Health and Disease. Nutrek Press: Brookline, MA.
- 125 Siguel E, Lerman R (1996). Prevalence of essential fatty acid deficiency in patients with chronic gastrointestinal disorders. Metabolism **45**: 12–23.
- 126 Simopoulos AP (1999). Essential fatty acids in health and chronic disease. Am J Clin Nutr. **70**(3 Suppl): 560S–569S.
- 127 Simopoulos AP (2008) Nutrition and fitness: cultural, genetic and metabolic aspects. World Rev Nutr Diet **98**: IX–XV.
- 128 Sorgi PJ, Hallowell EM, Hutchins HL, Sears B (2007). Effects of an open-label pilot study with high-dose EPA/DHA concentrates on plasma phospholipids and behavior in children with attention deficit hyperactivity disorder. Nutr J **6**: 16.
- 129 Stevens LJ, Zentall SS, Abate ML, Kuczek T, Burgess JR (1996). Omega-3 fatty acids in boys with behavior, learning, and health problems. Physiol Behav 59: 915–920.
- 130 Stinson AM, Wiegand RD, Anderson RE (1991). Fatty acid and molecular species compositions of phospholipids and diacylglycerols from rat retinal membranes. Exp.Eye Res 52: 213–218.
- 131 Stoll AL, Locke CA, Marangell LB, Severus WE (1999). Omega-3 fatty acids and bipolar disorder: a review.Prostaglandins Leukot Essent Fatty Acids. 60(5–6): 329–37.
- 132 Sublette ME, Russ MJ, Smith GS (2004). Evidence for a role of the arachidonic acid cascade in affective disorders: a review. Bipolar Disord 6: 95–105

- 133 Tiemeier H, van Tuijl HR, Hofman A, Kiliaan AJ, Breteler MM (2003). Plasma fatty acid composition and depression are associated in the elderly: The Rotterdam study. Am J Clin Nutr **78**: 40–46.
- 134 Vancassel S, Leman S, Hanonick L, Denis S, Roger J, Nollet M, Bodard S, Kousignian I, Belzung C, Chalon S (2008). n-3 polyunsaturated fatty acid supplementation reverses stress-induced modifications on brain monoamine levels in mice. J Lipid Res 49: 340–8.
- 135 Wainwright P (2000). Nutrition and behaviour: the role of n-3 fatty acids in cognitive function. Br J Nutr **83**(4): 337–9.
- 136 Warren G, Mckendrick M, Peet M (1999). The role of essential fatty acids in chronic fatigue syndrome. A case-controlled study of red-cell membrane essential fatty acids (EFA) and a placebo-controlled treatment study with high dose of EFA. Acta Neurol Scand **99**: 112–6.
- 137 Williard DE, Nwankwo JO, Kaduce TL, Hormon SD, Irons M, Moser HW, Raymond GV, Spector AA (2001). Identification of a fatty acid D6 desaturase deficiency in human skin fibroblasts. J Lipid Res 42: 501–508.
- 138 Wozniak J, Biederman J, Mick E, Waxmonsky J, Hantsoo L, Best C, Cluette-Brown JE, Laposata M (2007). Omega-3 fatty acid monotherapy for pediatric bipolar disorder: a prospective open-label trial. Eur Neuropsychopharmacol. **17**(6–7): 440–7.
- 139 Yao JK, van Kammen DP (2004). Membrane phospholipids and cytokine interaction in schizophrenia. Int Rev Neurobiol **59**: 297–326.
- 140 Young G, Conquer J (2005). Omega-3 fatty acids and neuropsychiatric disorders. Reprod Nutr Dev. **45**(1): 1–28.
- 141 Zeman M, Jirák R, Jáchymová M, Vecka M, Tvrzická E, Žák A (2009). Leptin, adiponectin, leptin to adiponectin ratio and insulin resistance in depressive women. Neuro Endocrinol Lett 30(3): 387–95.
- 142 Zeman M, Žák A, Vecka M, Tvrzická E, Písaříková A, Staňková B (2006). N-3 fatty acid supplementation decreases plasma homocysteine in diabetic dyslipidemia treated with statin-fibrate combination. J Nutr Biochem **17**(6): 379–84.
- 143 Zhang CE, Wei W, Liu YH, Peng JH, Tian Q, Liu GP, Zhang Y, Wang JZ (2009). Hyperhomocysteinemia Increases β -Amyloid by Enhancing Expression of γ -Secretase and Phosphorylation of Amyloid Precursor Protein in Rat Brain. Am J Pathol **174**: 1481–1491.
- 144 Zimmer L., Vancassel S., Cantagrel S., Breton P, Delamanche S, Guilloteau D, et al (2002). The dopamine mesocorticolimbic pathway is affected by deficiency in n-3 polyunsaturated fatty acids. Am J Clin Nutr **75**(4): 662–667.
- 145 Žák A, Hátle K, Mareš P, Vrána A, Zeman M, Šindelková E, Skořepa J, Hrabák P (1990). Effects of dietary n-3 fatty acids on the composition of cholesteryl esters and triglycerides in plasma and liver perfusate of the rat. J Nutr Biochem 1(9): 472–7.